

**Y-STR HAPLOTYPE DIVERSITY AND mtDNA
HVI SEQUENCE VARIATION AMONG MAJOR
TRIBES OF CHARSA DA AND MARDAN
DISTRICT**



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**DEPARTMENT OF GENETICS HAZARA
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Department of Genetics

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MARDAN DISTRICT**

BY

Sadia Tabassum

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2016**





DEDICATED

T

MY RESPECTED PARENTS

Who made me what I am today

(Ma Th Live Long)

AN

MY DAUGHTER FATIMA

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All the praise and glory is for almighty Allah who has showered on me his countless blessings and unbeatable strength to cope up with the challenges of life, without which the journey of life was impossible to go through.

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ABSTRACT

Substantial genetic diversity exists among the people of Pakistan and understanding evolution of this diversity is complicated due to several waves of migration from populations in the North and Northwest. Pathans, one of the largest ethnic groups of Pakistan residing mostly in the northwestern part which remained unexplored with respect to their biological affinity and haplotypic diversity, is the subject of this exploration. This dissertation reports the results of the PhD project wherein, we have assessed the extent of genetic diversity using Y-STR profiling and *mtDNA* sequence analyses of HVI control region of Pathan populations of Mardan and Charsada districts of KP province, Pakistan. Sum of 374 buccal swabs were collected from five major population of the two adjoining districts viz Charsada and Mardan. Y-STR profiling was carried out using Promega PowerPlex® Y23 System. Mitochondrial HVI data was also generated for 165 samples. Y-chromosomal and *mtDNA* haplogroups were assigned to each sample using a haplogroup predictor and phylotree, respectively. Principle Coordinate Analysis (PCoA) plots were generated by combining our data set with other published datasets from neighboring populations of central Asia, Middle East, Europe and South Asia. Our results revealed that the most frequent Y-STR haplogroup found was R1a (49.2%), followed by G2a (17.9%) and L (9.6%), while *mtDNA* HVI macro haplogroups R (63.4%), M (26.8%) and N (8.6%) were observed more frequent among Pathans of the area. Some novel mitochondrial haplogroups (*mtHgs*) of M and R subclades have also been recorded for the first time for Pathans. Analysis of our results

revealed that no significant genetic substructure was available among all the five populations with respect to 23 Y-STRs. Furthermore the Pathans of Charsada and Mardan are clustered together with little genetic difference. These people show homology with already studied Pathans from Pakistan, the Yousafzai tribe from northern proximity of Pakistan, as well as with the Pathans from Afghanistan and with some populations from Russian Federation, when compared on the basis of Y-STR profiles. The clustering pattern was different when compared on the basis of mitochondrial HVI data sets of different neighboring populations, which depicts more male mediated gene flow among populations than female mediated gene flow. In addition, the mitochondrial gene pool of these populations seems to be influenced by the Turkish invasion. The gene pool of the Pathan populations of Mardan and Charsada districts exhibit genetic affinities with other Pathans from Pakistan and Afghanistan, and exhibit shared haplogroups with Middle Eastern, Caucasian, South Asian and Central Asian people. The study provides a sound reference for collating molecular anthropology and genetic structure of the people of Pakistan.

Chapter 1

INTRODUCTION

A comprehensive rational interest, about the origin of mankind has always been compelled the human brain to seek logical answers. In fact the key inspiration behind this grilling and the search for human genetic variation has always been a challenge to cope up with and to explore the population structure in general and genetics of diseases in particular. Genetic variations available in the modern people also reflect the past of mankind, its origin and the spread of anatomically modern humans and their demographic history. It is increasingly more evident that knowledge about these processes is equally indispensable for a proper understanding of the genetics of complex diseases.

Decades long research of genetic variation has revealed that a significant degree of diversity and variation lies both within and between existing human populations around the globe. Despite the enormous amount of data gathered during this relatively long time span of “classical era”, has highlighted many basic problems but left them largely unresolved. Increasingly more informative “DNA era”, rapidly expanding during the last 35 years, took up the same list of problems and is formulating new ones. Largely irrespective of what kind of general questions related to demographic history are being asked, the present-day genetics investigates variation in three different systems i.e. bi-parental autosomal chromosomes, paternally inherited Y chromosome and maternally inherited mitochondrial DNA. In spite of having manifold large size of autosomal genes

than that of Y chromosome and mtDNA, the uniparentally inherited markers have powerful advantages, making them the favorite and handy tools of choice for the population geneticists. Besides their uniparental mode of inheritance, the advantage of their lacking of recombination is of prime importance that helps them to reconstruct genetic lineages back to the most recent common ancestors (MRCA). i.e. The Y chromosomal Adam and mitochondrial Eve. Furthermore in combination with information about the variation in autosomal genes, a promising synthesis is hopefully possible in the future. My thesis is about Y chromosomal STR haplotype and mtDNA HVI variation in major Pathan populations (tribes) of the two historically important geographically adjacent areas that remained under the influence of many foreign invaders. In order to understand the Y chromosomal STR variation and mtDNA HVI variation and its origin in these populations, additional background knowledge of those living outside this area is needed. Due to their scrupulous properties Y-STRs mtDNA allow to construct individual genealogies and connect them to a global phylogenetic tree of all humans. Basal branches of these Y Haplogroup and mtDNA trees have been shown to be highly continent specific, making these uniparentally inherited loci more valuable makers compared to other tools where their overall variation is hidden mainly within populations. The study of the spread and variability of Y-haplogroups and mtDNA clusters enable the scientists to address questions related to the early population movements of anatomically modern humans.

Through the collective effort of many labs and researchers around the globe, the Y-chromosomal and mitochondrial DNA clusters (haplogroups) have been investigated and explored in different continental regions.

South Asia comprising India, Pakistan, countries in the sub-Himalayan region and Myanmar was one of the first geographical regions to have been inhabited by modern humans. This region has served as a major route of dispersal to other geographical regions, including Southeast Asia. Populations of Pakistan show high genetic differentiation and extensive structuring as a result of evolutionary antiquity and commonly practiced endogamy. Linguistic variations of populations present the best account of genetic differences observed in this region of the world.

The progression of local population expansion and consequent dispersal to newer areas, coupled with evolution of cultural practices and geographical distance acting as a barrier to human contact, also resulted in the formation of intra-marrying groups; groups within which there was considerable gene exchange among individuals, but between which there was little genetic exchange. The development and evolution of language and culture embellished the formation of such groups.

Genetic diversity within a geographical area is a logical indicator of the age of the population's habitation in that specific area. However, it is also dependent on the population's effective sizes, thus entailing that the assessment of antiquity may not be simple. It is now well-established that Africa exhibits the highest genetic diversity among continental populations. The genetic diversity in Indian subcontinent, "the heartland of

south Asia", is second to that of Africa. Mitochondrial DNA (mtDNA) and the non-recombining region of the Y-chromosome (NRY) based Molecular genetic studies have supported a single southern dispersal route to Indian sub continent. The populations of Indian sub continent including Pakistan and India are all derivatives of mitochondrial M and N lineages, which themselves are the derivation of L3 lineage that now found only in Africa. The M and N lineages probably diverged from L3 shortly after their dispersal from Africa (Quintana *et al.*, 1999). Similarly Y-chromosomal data is also indicative of southern dispersal route and this is supported by the presence of C and D Y-haplogroups only in the Asian continent and Oceania (Kivisild *et al.*, 2003; Underhill *et al.*, 2001) and not in western Eurasia and North Africa.

Afghanistan being a landlocked nation in southwest Central Asia has central highlands of Hindu Kush Mountains extending from the northeast of the country to the southwest, separating Afghanistan into northern and southern provinces. Being at the intersection of Central Asia, South Asia, and the Middle East, Afghanistan has served as crossroads for human migrations and pilgrimages, including an important stop along the Silk Road. Mesolithic artifacts, Neolithic pottery of about 7.2kya, bones of domesticated animal and tools i.e. the sickle blades used to collect wild grasses, have been discovered during excavations in the Ghar-i-Mar site in north Afghanistan. These discoveries supported the early cultivation of wheat and barley around 9–11kya and domestication of animals around 7–9kya (Dupree *et al.*, 1972). Other recent archeological findings include Buddhist artifacts transported northward from India along the Silk Road, as well as captions and inscriptions engraved on rocks in ancient Hebrew dating from the eleventh to the

thirteenth centuries (Katzir, 2001). Historians speculated the onset of urbanization in Afghanistan around 4 to 4.5kya (Runion, 2007). According to Historical records Aryans from Iran were the first to occupy Afghanistan in eighth century B.C.E.), followed by Persians in sixth century B.C.E and Greeks in fourth century B.C.E. Later Mauryans from India and GrecoBactrian escorted Buddhism into this region while s Arabs and Mongols launched Islam (Runion, 2007). Dari and Pashto are two languages of Indo-European origin spoken by two major Ethnic groups of Afghanistan; the Tajiks and Pathans or Pathans, respectively. Pathans comprising the most prevalent ethnic group in Afghanistan (42%), inhabit primarily the south of the Hindu Kush mountains. According to legendary record and oral traditions, these

Pathans are considered to be the ancestors of Pathans in the Pakistan and Northern India. Different origins for Pathans, including Greek and Jewish ancestry, have been suggested (Runion, 2007; Caroe, 1976). These connections have not been confirmed and a few efforts characterizing the genetic structure of this group in Pakistan (Mohyuddin *et al.*, 2001; Qamar *et al.*, 2002; Mansoor *et al.*, 2004; Sengupta *et al.*, 2006; Firasat *et al.*, 2007), Afghanistan (Di Cristofaro, 2012) and India (Noor *et al.*, 2009) have been made. As previous studies were carried out without the tribal and subtribal status of the Pashtuns, so such study was of immense need to segregate them on the basis of their subtribal and caste level.

The present study was undertaken to ascertain, for the first time, the genetic diversity of major Pathan tribes of Pakistan residing in Charsada and Mardan district of KP province

such as Yousafzai, Mohmand, Muhammadzai and Kakakhel Mian, utilizing Y-chromosomal STR profile and mtDNA HVI sequence variation. The data produced were subsequently compared with formerly published geographically and ethnically connected indigenous and global populations to explore paternal and maternal signals of modern human's dispersals across this study area. In addition, we assessed genetic affinities between Pathans from our study area and Pathans from neighboring Afghanistan as being descendants of the same Y-chromosome, as well as their hypothesized phylogenetic relation to Greek and Jewish populations.

Based on the legendary information, it was hypothesized that the populations under investigation are the descendants and immigrants of Pathan ethnic group of Southern Afghanistan. So they must have close genetic ties with their suspected ancestors. On the basis of above mentioned hypothesis, several following queries were raised to be justified after this research.

1. What clusters (Haplogroups) are spread among populations of Charsada and Mardan?
2. How do these clusters connect to the global tree?
3. What is the diversity (%) of Y haplogroups and mtDNA haplogroups in these populations as compared to the populations who had invaded these areas and ruled for a long time?
4. What has been an impact of succeeding gene flow between these populations?

5. What has been a contribution of invading populations to the maternal and paternal gene pool of the populations under study?

The central focus of the literature overview of the current thesis will be therefore an overview of out of Africa migration and colonization of South Asian land mass by modern humans, the legendary ancestry of the populations under investigation and history of study area and finally the general skeleton tree connecting and characterizing the Y haplogroups and mtDNA haplogroups worldwide generally and in south Asia specifically.

Chapter2

REVIEW OF LITERATURE

Charles Darwin was the pioneer in suggesting the common African ancestry of human beings (Lafrenier, 2010). On the basis of some shared attributes he further proposed African apes to be their ancestors (Bowler *et al.*, 2003). Later on his proposition was rejected and after several genome analyses of people from different racial backgrounds, it was concluded that all modern humans evolved from one 'lucky mother' in Africa about 150,000 years ago (Cann *et al.*, 1987; Vigilant *et al.*, 1991). The recurrent development in molecular genetics and availability of analytical tools has revolutionized molecular anthropology which needs to be strengthened in Pakistan also.

2.1 Colonization of South Asia

According to Darwin human population started dispersing around 125,000 years before present (BP) using one route from the Nile Valley directed towards the Middle East, around 120,000–100,000 years BP and the second one through Bab-el-Mandeb Strait on the Red Sea, crossing through the Arabian Peninsula and finally heading towards the present-day United Arab Emirates around 125,000 years BP and Oman around 106,000 years BP (Lafrenier, 2010) and then consequently reaching the Indian Subcontinent at Jwalapuram around 75,000 years BP. Despite lacking any fossil record, the similarity among the stone tools collected from all these places, suggested that the modern humans to be their creator (Bower, 2011). These findings might give some support to the claim

that modern humans from Africa arrived at southern China about 100,000 years BP (Liu *et al.*, 2010), and the Liujiang hominid controversially dated at 139,000–111,000 years BP (Shena *et al.*, 2002). Dating results of the Lunadong (Buding Basin, Guangxi, southern China) teeth, which include a right upper second molar and a left lower second molar, indicate that the molars may be as old as 126,000 years.

For the previous exodus from Africa genetic analyses based on the Y chromosome and mitochondrial genome doesn't provide any clue, as they represent only a small part of the human genetic material. It could be assumed that those modern humans became extinct because of Toba catastrophe around 74,000 years BP. However, according to some arguments there was no remarkable impact of this catastrophe on human population (Balter, 2010).

Some other authorities claim that *Homo sapiens* first appeared around 200,000 years ago in Ethiopia (White *et al.*, 2003). After reaching Near East around 125,000 years ago, they moved back to Africa, as their settlements were replaced by Neanderthals. Now days it is strongly believed that the first modern human dispersal across Asia started about 75,000 years ago across the Bab el Mandib linking Ethiopia and Yemen. From there, some of these people moved further to South Asia around 50,000 years ago, and then to Australia by 46,000 years ago (Bowler *et al.*, 2003). *H. sapiens* reached Europe around 43,000 years ago (Wilford, 2011), replacing the Neanderthals around 24,000 years ago. They reached East Asia some 30,000 years ago.

Migration of several languages and cultures has also been noticed with these human migrations. The initial exodus out of Africa 125,000 years ago was followed by several waves of migration via the Arabian Peninsula into Eurasia around 60,000 years ago, where one group of migrants rapidly passes through coastal areas of the Indian Ocean and other group migrating north towards Central Asia.

Genetic evidence proved that modern humans have gone through genetic bottleneck resulting in reduced genetic diversity and then gone through a dramatic population growth from geographically dispersed populations around 50,000 years ago. And this idea of bottleneck was also supported by Henry Harpending's findings as well as geological and climatological evidences from Lake Toba explosion (Harpending *et al.*, 2009), that had resulted in overall population reduction up to 15,000 individuals that consequently leading to rapid racial differentiation as a possible consequence of founder effect and increased genetic drift (Ambrose, 1998).

According to Some genetic evidences the migration out of Africa had occurred along two different routes. However, other investigations revealed that a single migration occurred, followed by swift northern migration of a subset of the group. The group who followed southern route from West Asia spread generation by generation around the coast of Arabia and Persia finally reached Indo-Pak subcontinent and headed towards Australia around 55,000 to 40,000 years ago (fig.1). The group directed towards north (East Asians were the second group) ventured inland (Maca *et al.*, 2001) and exuded to

Europe, ultimately displacing the Neanderthals. They also radiated to India from Central Asia (Bowler *et al.*, 2003). Some recent evidences suggest that the human migration from Africa to Southeast Asia and Australia have occurred between 80,000 and 120,000 years ago (Ewen, 2015) following multiple routes and the *H. sapiens* interbred with other species like Neanderthals in Europe, Denisovans in Central Asia and *Homo erectus* (Dannel, 2012).

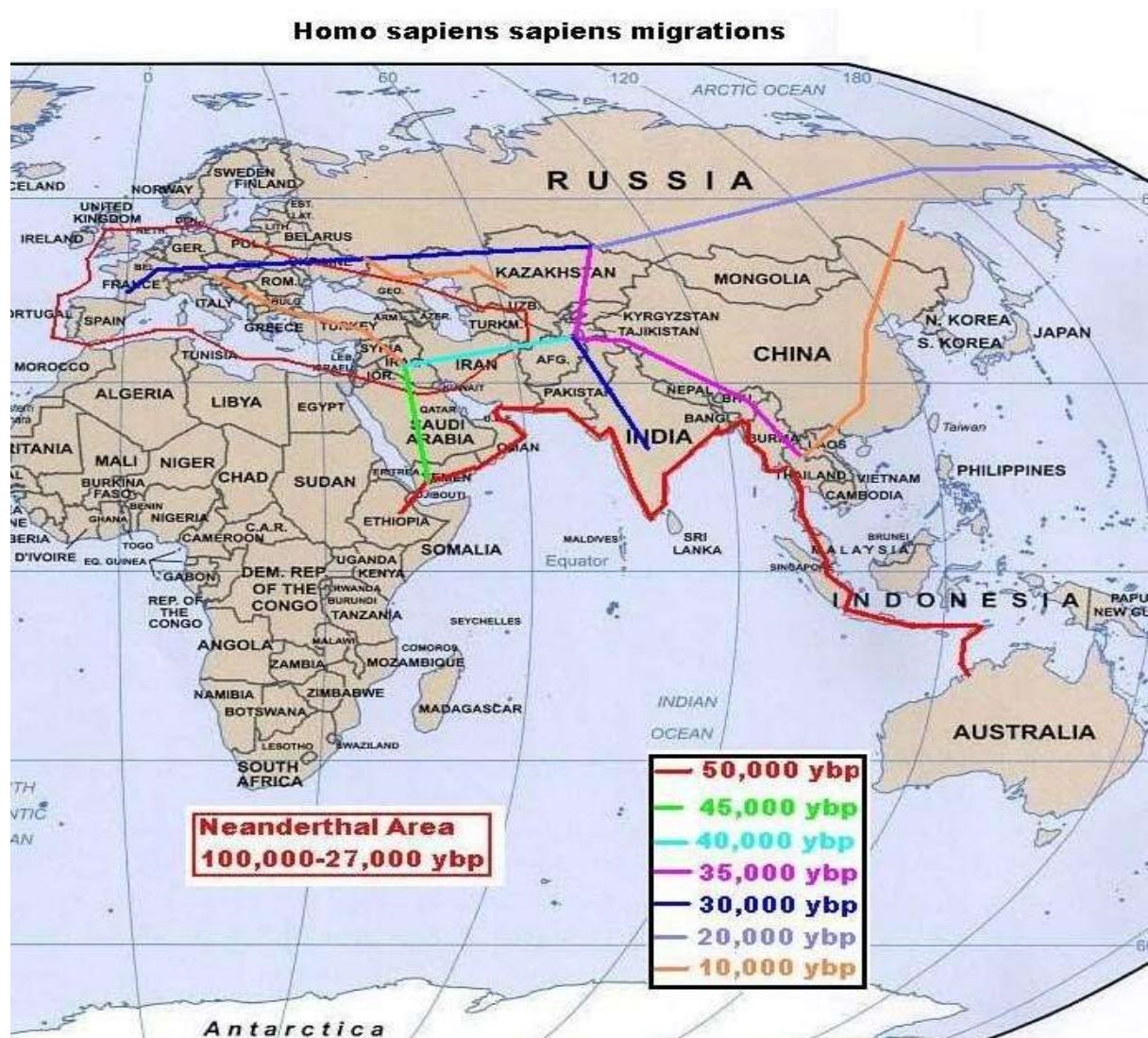


Fig.1. Probable human out of Africa migration time scale. (Source:

[http://www.roperld.com/YBiallelicHaplogroups.htm#routes\)](http://www.roperld.com/YBiallelicHaplogroups.htm#routes)

2.2 Pakistani civilization

Pakistan is a diverse country with a variety of cultures and ethnic standings available in its more than 180 million people. Historically it has been an important trading hub since the ancient Indus valley and Gandhara civilizations, Pakistan has had been exposed to many invaders and traders from around the world i.e. Africa, Middle East, Caucasus, Mediterranean and China. This influx has contributed to the vibrant mix of people in modern day Pakistan. The majority of Pakistani population belongs to an Indo-European ethnic groups comprising of various smaller ethnic groupings. Punjabis being the largest group constitute almost 44% of the population, Pathans being the second largest ethnic group constitute 15%, Sindhi constitute 14%, Saraiki 10%, Muhajir 7%, Baluchi 3% and the remaining 4% of the population consist of Hindkowans, Gujjars, Kashmiri, Potoharis, Chitralis, Hunzakuts, Dards, Balti and several other small groups.

2.3 The Pathans

Pathans, the Pashto speaking people also spelled as Pashtuns, Pakhtuns, Pukhtuns, broadly referred to as Pathans or ethnic Afghans (Banuazizi *et al.*, 1994) belong to an Eastern Iranian ethno-linguistic group residing for the most part of Eastern and southern Afghanistan and in Khyber Pakhtunkhwa Province, Federally Administered Tribal Areas and parts of Baluchistan Province of Pakistan. They are characterized by Pashto language, their culture, honor and the pre-Islamic code of conduct Pukhtunwali they practice

(Oliver, 1980). Their unified modern past is sometimes referred to the establishment of the rise of the Durrani Empire in 1747.

Majority of Pathan tribes live in an area stretching from western Pakistan to southwestern Afghanistan. Additional Pathan communities reside in the Northern Areas, Azad Kashmir and Sindh provinces of Pakistan as well as throughout Afghanistan and in the Iranian region of Khorasan. A large migrant worker community lives in the countries of the Arabian Peninsula and in smaller communities in Europe and North America. A sizable community of largely putative ancestry calls India home. Important metropolitan centers of Pathan culture include Mardan and Kandahar. In addition, Peshawar, Quetta and Kabul are ethnically mixed cities with large Pathan populations. With 1.5 million ethnic Pathans, Karachi stands the largest Pathan city in the world.

Pathans with total population of over 15.4 percent or 25.6 million people are widely dispersed in Pakistan inhabiting vast area from Khyber to Bolan including Peshawar, Quetta and Karachi (the largest Pathan city in the world) (Shehzad, 2007). They are also found in vast majority in Afghanistan with the estimated population of 42% or 12.5 million people, in Khorasan (Iran) and India (Lewis, 2009).

2.3.1 History and origin

The Pathan areas have faced many foreign invasions of Aryans, Medes, Persians, Mauryans, Scythians, Kushans, Greeks, Hapthalites, Turks, Mongols and Arabs. They

have an ancient history, much still unresearched. Various scholars and researchers have proposed different theories about the origin of Pathans.

2.3.2 Historical references

Herodotus, the famous Greek historian first referred to the people called Pactyans, inhabiting Arachosia, the eastern frontier of the Persian Satrapy in the 1st millennium B.C.E. (Rawlinson, 1858). Another reference about the existence of a tribe called Pakhtas residing in the region of Pakhat in Afghanistan, is given in the Rig-Veda, and some historians have hypothesized them to be the early ancestors of the Pathans. Some others linked Bactrians to the Pathans who used other similar Middle Iranian dialect.

Historically Pathans are referred to as ethnic Afghans. The term Pathan was used as synonym of Afghan until the dawn of modern Afghanistan and the establishment of Durand Line, a 250 Km long border line drawn by the British Mortimer Durand in 1893. According to various scholars it was believed, "The term Afghan was first used in 982 C.E." in Hudud-al-Alam (Minorsky, 1940). The Pathans consider Afghans to be their common renowned ancestor. According to Some historians Pathans believed to emerge in the region of Kandahar and the Suleiman Mountains, and started expanding 1000s of years ago. At the site of their origin they would used to be in close proximity with the Persians and Mauryans and may have been following Zoroastrians,

Buddhism, Hinduism or Judaism before to the advent of Arab Muslims in the seventh century (Dupree, 1977; Holdich, T. H. 1899).

2.3.3 Anthropology and linguistics

The Pathans have been considered as the modern day descendants of Scythians speaking Pashto, a sub-branch of the Indo-Iranian branch of the Indo-European family of languages. Pashto language has ancient origin and has similarity with Bactrian and Avestan language. Pamir dialects, such as Ossetic, Shughni and Wakhi are considered to be its closest modern relatives. Pashto has borrowed many words from other languages like Sanskrit and Persian. Foreign Invaders also have left their language foot prints on Pashto and it has borrowed many words from Arabic, Greek and Turkic languages (Awde *et al.*, 2002).

According to Yu V. Gankovsky, the Pathans emerged as a "union of East-Iranian tribes that provided the ethnic platform for the Pathan ethno genesis, in the middle of the first millennium C.E. (Gankovsky, 1982). Early precursors to the Pathans had been so the ancient Iranian tribes that stretched all through the eastern Iranian Plateau (Harvey, S. 2014). Based on the Southern or Northern Pashto dialect they speak, the Pathans either called Pathans or Pukhtuns respectively. Phenotypically, Pathans generally represent the Mediterranean people (Blood, 2001) having light hair tones and eye colors. Several Anthropologists documented the oral traditions about the origin of Pathans. Some anthropologists lend credence to the mythical oral traditions of the Pathan tribes

themselves. For instance, the Encyclopedia of Islam, highlighted the theory of Pathan descent from Israelites which could be traced from “Maghzan-e-Afghani” that was compiled in 17th century C.E. by a famous scholar in the court of Mughal emperor Jahangir, named Naimat ullah Haravi. Several other Pathan writers and historians followed the same theory in their writings. Same theory was also referred by Olaf Caroe in his writing “The Pathans”.

According to the scriptures of sir William Johns, Afghans are considered to be one of the lost tribes of Bani Israel who broke away from captivity of Babylonian king Bakht Nasr and took refuge in the area of Asarah in Afghanistan. Armia and Barkhia were the two of the six sons of Sawal or Saul, a successor of Yahuda, a son of Hazrat Yaqub (AWS). Armia had a son Afghan and Barkhia had a son named Asif. Afghan was selected as a commander of an army in the kingdom of Dawood and Suleiman, after the death of Sawal. Later when Palestine was captured by Bakht Nasr, Afghan and Asif migrated to Ghor and took control of the area from the local tribes. Similarly, according to *Taaqati-Nasiri*, people called Bani Israel settled in Ghor, Afghanistan, and then started migrating towards south and east. It is documented that in 722 BCE after the conquest of Neo-Assyrian empire, the descendants of Afghan and Asif who were expelled from their homeland Palestine and lost their way, were ten in number. One of the lost tribe moved towards Mecca and met an Islamic worrier and an Israelite tribes man, Khalid bin Waleed. He then sent a message to the Afghans settled in Ghor to embrace Islam. A delegation under the command of Qais moved to Madina where they met Prophet

Muhammad (PBUH) and embraced a new faith of Islam and Qais was given a new name "Abdur Rashid" by The Prophet. Later he was given a title of "Bathan" where B was replaced by P with the passage of time and all his descendants were called Bathans or Pathans.

Those Bani Israel references are in good promise with the traditional view held by Pathans, that the tribe of Joseph among other Hebrew tribes inhabited this region when twelve tribes of Israel from twelve sons of Jacob dispersed. Bani-Israel theory stated in *Maghzan-e-Afghani* has been interrogated due to lack of historical justifications and linguistic inconsistencies, for examples the exiled of ten lost tribes from Assyria. While in *Maghzan-e-Afghani* it is referred to the ruler of Persia, authorizing those tribes to go east to Afghanistan (Harawi *et al.*, 1960). That contradiction can be enlightened by Persian control over the Assyrian Empire's lands after taking control over Babylonia. Yet there is no record for mobility of these Israelites to further east, provided by ancient authors yet, that oral tradition has been widely accepted among

Pathans. The Pathans residence in the region of Afghanistan has also been mentioned in Rig Veda that was probably composed before 1200 B.C. There is no evidence of Israelite or Jewish connection provided by ancient authors before the conversion of the Pathans to Islam.

Another group of historians believe in the Aryan descent of Pathans. According to some historians, Pathans are considered to be of Northern European lineage. Some others traced their ancestry to the Southern Russia while some consider them to be of Mongolian

and Chinese Turkistani origin. A vast group of modern researchers consider the area of “Bakhtar” between Pamir and Oxus River to be their possible birth place.

After a gradual increase in number, they started moving out of this area and scattered towards the valley of Swat and Indus River after crossing Hindukush. This group of people was called Indo-Aryana. They kept on moving towards Punjab and finally reached the valleys of Ganga and Jumna overthrowing the local Dravidian tribes. Another group from Bakhtar migrated westward towards Iran. Some historians consider them to be the ancestors of Pathans. The modern researches like Fester Tytler proposed a “Mixed Race Theory” which documented the Aryan origin of Pathans having elements of Mongol, Turkish and other strains. This idea was strengthened by the opinion of Charles Miler about the amalgamation of Greek, Mongol, Persians, Scythians and Turks to the afghan stock. Several vestiges and leftovers of Syriac language were found during mining and excavation of the Gandhara, Kandhar, Taxila and Lughman areas, providing strong evidence of the presence of Syriac people in the area. Later in 5th century this land was invaded by Haphthalites, “white Huns”. In Umayyad supremacy, Arabs invaded the area and merged with the local population. In 13th century the “yellow race” of Chingiz Khan also stepped in and got mixed up with the local Afghan population.

Some historians connect the Pathan or Afghan lineage from Hazrat Ibrahim through his six sons who settled in the North West area of Iran from where they were expelled and inhabited “Parthia” or Pasht or Pakht. For which they were later called “Pashtuns” (Kakakhel, 1981).

These mythical oral traditions have been strengthened as a consequence of long cultural and political struggle between Pathans and the Mughals, elucidating the historical background for the formulation of the myth, the inconsistencies of the mythology, and the linguistic investigation that disproves any Semitic origins (Harawi *et al.*, 1960). Several other Pathan tribes claim to be descended from Arabs, including some i.e. Sayyid or Kakakhel Mian even claim their descent from the Muslim Prophet Muhammad (PBUH). Some other Pathan groups i.e. Afridi, Khattak and Sadozai from Peshawar and Kandahar also claim their descent from Alexander the Great of Greece (Mansoor *et al.*, 2004).

2.3.4 Ancestral alias

Pathan can only claim to be Pashtun if his father is Pashtun. The patrilineal definition is based on an important conventional law of Pashtunwali. That law has maintained the tradition of exclusively patriarchal tribal lineage intact. Under such definition, an ethnic Pathans have less concern with the language he speaks and more for his father. So, the Pathans who have lost both the language and seemingly many of the ways of their alleged ancestors can remain "Pashtun" by tracing their fathers' ethnic heritage back to the Pashtun tribes as they are said to be descended from Qais Abdur Rashid who has been considered as a possible Progenitor of Pathans (Allah *et al.*, 2013). According to ancient historians, Qais, travelled to Arabian Peninsula to meet Prophet Muhammad in Madina and after embracing Islam returned back to Afghanistan and present day

Pakistan. Then got married to Khalid bin waleed" s daughter and ostensibly had many children and four sons Sarban, Baitan, Ghourghusht and Karlan, who commenced towards the east i.e. each one moving its way towards Swat, Lahore, Multan and Quetta respectively.

2.4 Study Area

The study area includes two geographically adjacent districts of Khyber Pakhtunkhwa (KP) province of Pakistan. i.e. Charsada and Mardan. Location map of the study area is provided in Fig.2.

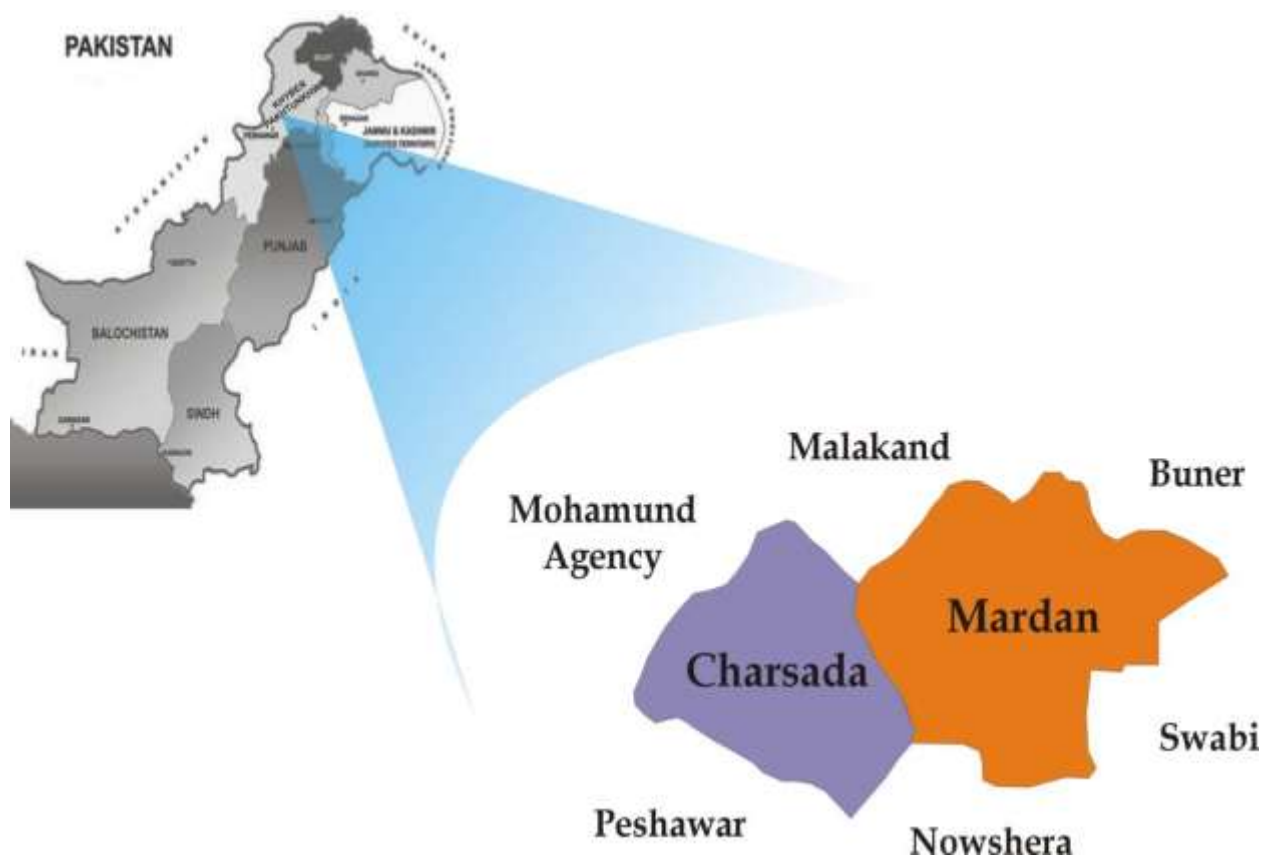


Fig.2. Location map of Charsada and Mardan district (Khyberpakhtunkhwa) Pakistan.

2.4.1 The Charsada District

Charsada is one of the 25 districts in the Khyber Pakhtunkhwa province of Pakistan. The town of Charsada was once part of the Peshawar region. Vast majority of the population of the area is Pathan.

2.4.1.1 Geography

It occupies total area of 996 km² with total population of almost 10.2 million individuals according to 1998 census report, and 16.2 million according to 2014 report. It is situated at 34° 03" and 34° 38" north latitude and 71° 28" to 71° 53" east longitude. It is bordered by Malakand district at its north, Mardan district at its east, Nowshera, Peshawar at south and federally administrated tribal areas (FATA) and Mohmand agency at its west (fig.2). Being geographically adjacent to Mardan district, the climate of Charsada is similar to that of Mardan.

Administratively it is divided into three tehsils (Charsada, Shabqadar and Tangi) collectively consisting of a total of 46 Union Councils.

2.4.1.2 History

It is considered as one of the most ancient and historical sites of Asia characterized by possessing several impressive mounds in the area. Charsada remained the part of the Gandhara kingdom that had been a part of 7th strappy of Achaemenid Empire around

516 BC until it was overthrown by Alexander the Great in the 4th century BC. The city was detained by Alexander" s army in 327 BC. The Indian sovereign Chandragupta Maurya took over the control of Gandhara after the death of Alexander the great in 323 BC. After this, Maurya" s grandson Ashoka became an emperor and in his reign he built one stupa there of 2.5 miles perimeter which was later revealed by Hieun Tsang., a legendry Chinese Buddhist pilgrim in 630. After the conquest of Mahmud of Ghazni and arrival of Islam in 1026, the name Gandhara was faded away (Muhammadzai, 2012; Ali, 1993). Between 250–125 BC, this area was also governed and ruled by the Bactrian Greeks which was later succeeded by the Indo-Greek Kingdom who ruled until 10 AD. Formerly, the town of Charsada was known as Pushkalavati (Lotus City). This name was first cited in the Hindu epic chronicle the "Ramayana". It was based on the cultivation of Lotus in this area that can be witnessed by archeological records and coins found from this area with the picture of goddess carrying a bunch of Lotus flower (Marshall and Vogel, 1903: 176). According to Ramanaya, Ramachandra" s brother Bharata founded two cities of Taksha and Pushkala on the names of his two sons, after the conquest of Indus valley (Dani, 1963). According to Vaidya, these two towns were named Taxila and Peukhlaoti in Greek history. This name was replaced by a Persian name "Hashtnagar" with the arrival of Muslims in this region in 11th century. Hasht meaning "eight", so its eight main villages Prang, Rajjar, Utmanzai, Umarzai, Tangi, Sherpao, Turangzai and CharsadaBazaar gave its name Hashtnagar. This name was first mentioned in Babur" s memories of 6th century (Babur, 1987: 55). Garrick was of the view that the name Hashtnagar was a combination of two words of two different languages

i.e. Persian and Sanskrit (Garrick, 1881-82). According to Cunningham, the name was derived from its original name "Hastinagra" that was named after the ruler of this area, Hasti or astes (Cunningham, 1963). Later on the name evolved into Charsada when Ilyas Khan Muhammadzai, an Afghan conqueror settled here in 1109 and divided the land among his four sons. As in Persian the share of land was called Rasad, so it became Char-Rasad or four shares (Gopaldas, 1889). This area had been extended over a large area, and the entire vicinity is covered with vast ruins. Excavation was accomplished in district Charsada for about two months in 1902. Bala Hisar, an archeological site was exhumed twice by the Sir John Marshall in 1902 and by Sir Mortimer Wheeler in 1958.

According to Wheeler, "Bala Hisar was established by the Persians in the 6th century BC as a camp protecting the eastern edge of their territory" (Wheeler, 1962). Several interesting remains of coins and pottery ornaments were found. A series of post holes and ceramic remains and other archeological deposits recovered from Charsada excavations are dated to 1400 BC (Hussain, 1992). Subsequent epochs signify that other more permanent structures including stone-lined pits were also constructed at Charsada. Charsada people started developing an iron-working industry and started using ceramics between 14th to 6th century BC distinctive for this period in the gorge of Peshawar, Dir and Swat.

From 6th century BC to 2nd Century CE, Charsada remained the capital of Gandhara Kingdom being its administrative centre. This area was ruled by many foreign invaders including Persians, Mauryans, Greeks (Alexander), Greco-Bactrians, Scythians,

Parthians, Kushans, Huns, Guptas and Turks (Gazetteer, 1997-98).

2.4.1.3 Tribes

80% of the population of district Charsada belongs to Muhammadzai tribe which is subdivided into groups locally called "khel". These sub-tribes (khels) have the status of khans of town. In addition several Sayyid families also exist. Mainly are Kakakhel Mian/Sayyid who claims to be the descendants of Sheikh Rehamkar Hazrat Kaka Sahib the most venerated Pir or Spiritual Figure among the entire Pathan population in Pakhtunkhwa Pakistan and Afghanistan. This tribe has gained special respect and honor among other Pathan tribes due to their virtuous nature and pious descent and ancestry. They are declared as land lords for having lands of prime importance in Charsada and neighboring districts.

2.4.2 The Mardan District

Pir Mardan Shah was a prominent religious figure of a small area which was named after him. Later a vast surrounding area merged into this small territory of Mardan.

This area was once a part of Gandhara kingdom being a fraction of Peshawar valley.

2.4.2.1 Geography and Topography

The district occupies total area of 1632 square kilometers at 34° 05" to 34° 32" north latitudes and 71° 48" to 72° 25" east longitudes. It is delimited by Swabi and Buner districts on east while Buner and Malakand protected area on north, Nowshera district on south and Charsada district and Malakand protected area on the west (fig.2). At

northern side the district is bounded by high mountains with highest points of Pajja or Sarka 2056m high and Garo or Pato at the height of 1816m above sea level, while there is a vast south western fertile plain with some low hills encumbered through it. There is a steep slope running through the Plain carrying rain water to the Kabul River from foot hills. Water bodies and streams flow from North to south direction. The important streams of the area are: Kalpani, Baghiari khawar, Muqam Khawar and Naranji khawar. Its climate is considerably hot and humid in summer with frequent dust storms at night. December and January are the coldest months with minimum temperature up to 0.5°C. According to 1998 census, its total population was recorded almost 14.6 million with approximately 3.01% growth rate.

2.4.2.2 History of Mardan district

The area comprising Mardan district is a part of the Peshawar valley, which first emerged on map as part of the Gandhara kingdom. After invasion of Alexander the Great, the mists of murkiness began to disperse. The Alexander" s armies invaded the Indus valley through two different routes, one via Khyber Pass and the other through Kunar, Swat, Bajaur and Buner in 326 B.C. After Alexander" s exodus, Chandragupta took over the control of this valley from 297 to 321 B.C. Buddhism was at its full swing in Peshawar valley for the period of the supremacy of the Buddhist emperor Asoka, Chandragupta" s grandson. In the reign of emperor Mehanda after the Greeks took over the valley, there was a revival of Brahmanism. After this, the Scythians and Indians followed and kept hold of the valley till the 7th century A.D.

By the end of 7th century, the Afghans invaded the valley. At that time Peshawar valley was governed by the rulers at Lahore. The Afghans united to the Gakkhars and overthrew the Lahore rulers and took over the control of the hill country south of the Kabul and west of the Indus River. The area came under command of the Sultan Sabuktgin after defeating the Hindu ruler of Lahore Raja Jaipal in 10th century. After this the area had become a rallying point of several incursions and foray to interior India, for Sultan Mahmud Ghaznavi, the son of Sabuktgin. Ghaznavid era came to an end when Pathans of Ghor took over in 15th century. The area was then invaded by Mughal emperor Babar via Khyber Pass in 1505 and remained under the authority of several Mughal emperors up to Aurangzeb. During his government the Pathan tribes revolted and in spite of continuous effort of two years from 1673-1675, to re-establish his authority Mughals were forced to agree with the terms set by Pathans for to retain their independence. All the territory west of the Indus including present day Mardan District was ceded to Nadir Shah by Mughals in 1738. Ranjit Singh after taking control of Attock in 1814 and Peshawar in 1818 handed over the command to Hari Singh Nalwa and moved further to Lahore. Sikhs ruled this valley until 1849 and were overpowered by the British Army in the Second Sikh War. Major Lawrence was appointed as the first Deputy Commissioner of Peshawar and Peshawar was declared an administrative district under the Government of Punjab. At that time the present Mardan district was merged with Peshawar district and later in 1909 Frontier Province was came into being. Peshawar district was branched off into Peshawar and Mardan districts in 1937 (Hastings, 1878).

2.4.2.3 Ethnicity

Yousafzai Pathans constitute the majority of Mardan population. But in Lundkhwar valley of Tehsil Takhtbhai sizeable Mohmand population also exist. The origin of the Yousafzai tribe is traced back to two brothers Khakai and Gori. They gave their names to the two divisions of the tribes inhabiting the area near Kandhar. In the middle of the 13th century, the Khakal were consequently expelled by the second group of Ghoris and they settled in the area near Kabul. After getting enough wealth and fame and getting increase in population size, they divided into three clans, the Turklays, Gigyanis and Yousafzai. By the end of 15th century, the Yousafzai and Gigyani tribes started moving towards the plains of Peshawar and ultimately overthrew Dalazaks and dispersed into Buner district. They finally settled into Mardan district and the area was known as “Yousafzai Plain”.

2.5 Tribes under Scientific investigation

2.5.1 Muhammadzai

The Muhammadzai also pronounced as Mohammadzai, Mohammedzai, Mohmandzai, Mamanzai (Murray, 1899). The Sarbani Pathan tribe should not be mystified with the Muhammadzai of the Barakzai Durrani lineage of Afghanistan. According to Pathan family history, the Muhammadzai Pathans claim to be descended from Qais Abdur Rashid who had three sons; Sarban, Bait and Ghourghusht. His son Sarban is known to be the ancestor of Muhammadzai of Charsada through his son Kharshban while Afghani Muhammadzai are said to be descendants of Sharkhbun, the Kharshbun's brother. One of the three sons of Kharshban named Zamand had a son named

Muhammad who is the actual founder of this tribe (Caroe, 1957; Rose, 1997).

Muhammadzai tribe is found primarily in Hashtnagar, an area in present day Charsada District. Initially it was said to have settled in Khorasan, but migrated to Pakistan and was given the Hashtnagar by the Yousafzai tribe (Elphinstone, 1815). In 9th century the area near Qandhar named Pusheen was inhabited by the Zaman tribe. The area was later invaded by Tareen tribe and zaman tribe was expelled from their home land. Muhammadzai tribe before this expulsion started scattering from Afghanistan and reached Eastern province of Afghanistan Ningharhar. Where they had battle with Yousafzai tribe but defeated by them. Even after defeat they were not expelled from Yousafzai area of Ningharhar and were asked for help from Yousafzai, when they planned attack against Dalazaks. Muhammadzai joined Yousafzai but asked for award of territory of Hashtnagar which they achieved after their victory. They still retain their hold. Mir panda Khan, Malik Khyzer Khan and Hegi Khan were declared the tribe leaders. In 1526-1530, uniting with Mughal emperor Babur conquered Peshawar and overpowered Muhammadzai and Yousafzai. After Babur, Muhammadzai took over the control of the area. The geography of tribes is fundamental to their internal organization, because the subdivisions of the tribe and their respective villages are known by the same names.

2.5.2 Mohmand

For centuries between the basins of the Oxus River and Tarnak River in Ghazni, there lived a Mohmand tribe. This tribe has purely Afghan origin and follows strict pre Islamic honorific code “Pashtunwali”. At the time of British invasion in Peshawar valley, the area comprising the present day Mohmand agency was under the influence of the local tribesmen and was also governed by Khan of Lalpura.

The Mohmand had many rebellions against the foreign invasions and specifically against British India. Many Khels of Mohmand tribe were given assurance in 1893 from the Emir of Afghanistan that they will not be endured severance because of their past association with Afghanistan, and they were known to be Assured Clans. Mohmand living in Charsada and Mardan district are said to be migrated from Mohmand area of Mohmand Agency, which is bounded by Charsada and the plains of Peshawar on East.

There are also several sub clans of Mohmand i.e. Khawezai, Tarakzai, Baizai and Halimzai which are further subdivided in several sub clans and divisions.

The famous Sanskrit grammarian and historian Panini also referred to the name of tribe the *Madhumants* (the modern day Mohmand) in his *Ashtadhyayi* (Grammar of Sanskrit) who colonized the northwestern areas, in the 5th century BC. (Roy *et al.* 2007).

2.5.3 Yousafzai

Yousafzai tribe is considered to be the largest Pathan tribe initially originated in

Qandhar, the present-day Afghanistan. Arrived and settled in Kabul in 1446 while migrating eastwards and there they remained under the influence of Turkic governor Ulugh Beg.

After further eastward movement towards Swat valley they faced rebellion with local Dilazak tribe. After a continuous effort of 20 years, under command of their chief Malik Ahmed Khan, the Yousafzai tribe along with some allied clans (Jadoon and Uthmankhel) ultimately succeeded to throw out Dalazaks from their home town and they were then pushed eastwards to the Hazara Mountains east of the Indus River, after the encounter of Katlang.

In 330 BC Alexander mentioned a tribe named "Isapzais". Similar name was also pointed out by Babur in 16th century. Because of their non cooperative attitude, Akbar tried to subdue them under the command of Raja Bir Bar. Consequently Raja Bir Bar was killed by Yousafzai. It was due to their infuriated nature that they could not be completely brought under dominion of Mughal Empire until 1690 (Richards, 1993). Pir Babawas their first emir. After Akbar Shah's death in 1857, Akhund Ghaffur took over the control of state and ruled the state until 20th century (Haroon, 2011).

The Yousafzai tribe is predominantly found in the districts of Swat, Shangla, Buner, Malakand, Mardan, Tor Ghar, Swabi, Lower Dir, Upper Dir in the Khyber Pakhtunkhwa province of Pakistan, besides some living in Battagram and Oghi (Mansehra).

They also inhabit areas other than Khyber Pakhtunkhwa in Pakistan e.g. a Brahui and Persian speaking Yousafzai clan has also been found in Mastung district Baluchistan.

The predominant language of this tribe is Pakhto (Pashto), the northern variant of Pashto with the hard “kh” replacing “sh” in soft southern variant. Some Yousafzai lineages are found in Andhra Pradesh, Uttar Pradesh, Madhya Pradesh, Gaya, Bihar, Gujrat and Banglore (India).

2.5.4 Kakakhel Mian

The Kakakhel Mian is considered to be a prominent Sayyidclan. Their roots reach to Hazrat Ali bin Ismail bin Imame-Jafer Sadiq. Sayyid Kastir Gul an Islamic Sufi or wali is their probable ancestor. Another probable ancestor is Sheikh Rehamkar, a student of Sheikh Hazrat Akhund Adeen/Adyan Seljuki. The title of Rehamkar was given by Sayyid Abdul Wahab Akhund Panju Baba.

Kastir Gul was affectionately called "Kaka Sahib", so his descendants are entitled Kakakhel. The probable place of origin of this clan is a small village named "Kakasaib" in Nowshera District, KhyberPakhtunkhwaProvince from where they spread throughout the province especially in the nearby area of Charsada district (District census report, 1998).

The word “Kaka” means “uncle” and “Khel” means, “sons” or “children”. So the word "Kakakhel" means the "Children of the Uncle". As Kakasaib was affectionately called “Kaka” by everyone in the village, so the name Kakakhel was given to his descendents.

Until late 13th century, Kakakhel Mian was least known clan because of their numerical scarcity beyond their borders. But they started getting prominence in 19th century by considerable increase in their population size. Their religious education and righteous nature contributed to high esteem among other Pathans. Kakakhel have been recognized as forefront fighters in battle against Sikhs (Official Gazette, 1870). During the British Raj, Kakakhel made a number of contributions to society. They proved to be highly competent civil contractors, soldiers, diplomats and police officers.

Kakakhel Mian has also made contribution to the independence movement and they have also served Pashto and Persian language for decades. Several writers and poets of Pashto literature belong to Kakakhel clan of Sayyid. The intellectuals and writers from Kaka sahib developed a union named "Milliah Rehamkaria" at the onset of 20th century where they donated a collection of about 4000 books, magazines and newspapers. Sayyid Bahadur Shah Zafar Kakakhel, a renowned intellectual compiled a dictionary comprising 45000 words. Members of this clan use "shah" as their surname (Gopaldas, 1874).

2.6 Mitochondrial genome

The length of Human mitochondrial DNA (mtDNA) is 16,569 base pairs that code for 13 proteins involved in oxidative phosphorylation, 2 rRNA and 22 tRNA molecules (Fig.3). Several other proteins i.e. RNA and DNA polymerases and RNA processing enzymes are of nuclear origin and are imported into mitochondria via double mitochondrial membrane (Cummins, 1998). Due to unsymmetrical distribution of Gs and Cs in both strands of the circular mtDNA, one strand is considered as Heavy (H) and other is Light

(L). Mitochondrial DNA is a circular molecule present in multiple copies per cell ranging from 100 to 1000. This attribute makes it useful in the presence of small quantity of sequence of interest, especially in case of fossil study. Because only an egg donates its mitochondria to the developing embryo with a few exceptions, in all humans mtDNA is maternally inherited (Giles *et al.* 1980; Cummins, 2000). The mitochondrion of sperm is degenerated by the enzymatic activity of ubiquitin (Thompson *et al.*, 2003). As the individual harbors only one type of multiple copies mitochondrial DNA, said to be homoplasmic but become Heteroplasmic when mutation occurs in any of the mtDNA copy during segregation (Cavalier *et al.*, 2000; Alonso *et al.*, 2002). Unlike nuclear genome there is no recombination mechanism in mtDNA (Meriwether *et al.* 1991). Similarly mtDNA is also lacking of repair mechanism that consequently leads to the much higher mutation rate than nuclear genome sequences (Brown *et al.* 1979, 1982; Saccone 2000). This attribute is the basis of sequence variation among individuals (Olivio *et al.*, 1983). Its high copy number renders it useful tool in DNA profiling for forensic investigations (Allen *et al.*, 1998).

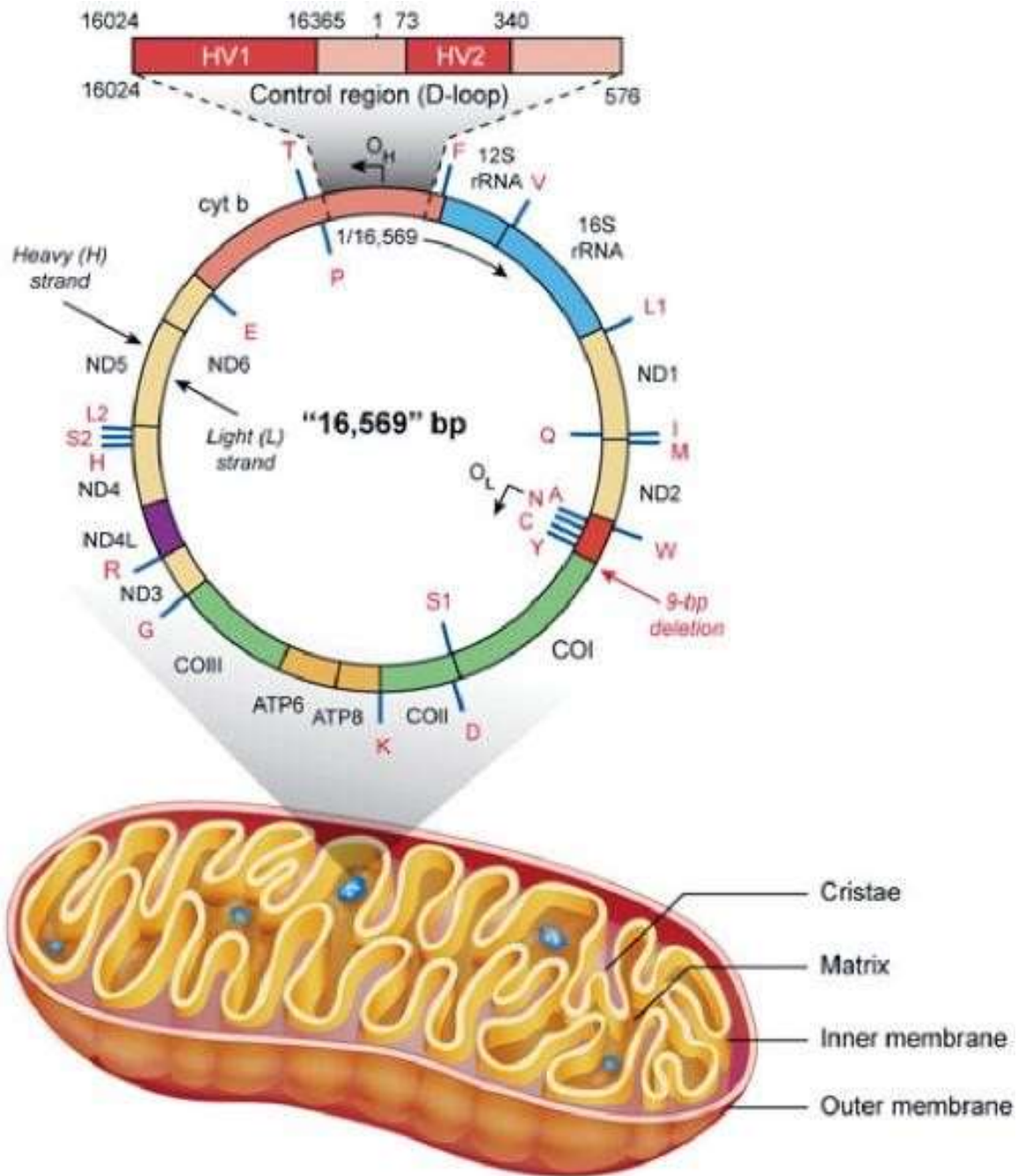


Fig.3. The 16,569 bp, circular human mitochondrial genome. The control region (D-loop) with the two hyper variable regions: HV1 and HV2 which are commonly used in Population genetic investigations (Lembering, M. 20013).

2.6.1 Tracing human migration from mtDNA

A few decades back, the only possible way of tracing human migration and to get insight into the evolution was to expose and uncover the skeletal remains. However, several breakthroughs have come up in the last two decades for the anthropologists with the advancement in the field of molecular biology. Until late 19th century, analysis of biomolecules like proteins was proved to be the useful tool for evolutionary studies but with less accuracy. With the discovery and introduction new genetic tool and molecular techniques, to get deep insight into the intra and inter population variation has been easier for the anthropologists to formulate new evolutionary and migration theories. DNA whether nuclear or extra nuclear has a tendency to record set of mutations in its sequences taking place with the passage of time so, act as a biological calendar. The role of mitochondria in several biological and physiological processes has long been known but their role in tracking evolutionary history has recently been studied in the last three decades. The rate of sudden changes/ mutations accumulated over time, defines the rate of evolution that can proves to be a useful tool to elucidate the time and age of divergence. e. g. Genetic and paleontological data suggested the common lineage of humans and Chimpanzee and their time of divergence was nearly 5million years ago (Ingman *et al.*, 2000). Whole mitochondrial genome doesn" t evolve simultaneously at same rate (Pesole *et al.*, 1999). The highest rate of mutation is in non coding triple stranded stretch of mtDNA called D-loop or displacement loop (Wallace *et al.*, 1995) and said to have three hyper variable regions, HVI, HVII and HVIII (Stoneking, 2000). This region comprises about 7% of mtDNA and has been studied and scrutinized in detail (Ingman *et al.*, 2000).

The rate of mutation in D-loop region is 10 times higher than in coding region (Howell *et al.*, 2007). On average 1-3 bp per 100bp varies among two unrelated individuals at random in their D-loop sequences (Piercy *et al.*, 1993). Although there are some kinds of mutation which make comparisons difficult e.g. back mutation and parallel mutation. Similarly the presence of hot spots for mutations also renders the comparison difficult but sequencing of complete human mitochondrial genome has offered many polymorphic sites for comparison. The HVI region ranges from 16024-16365 np and HVII ranges from 73-340 np while HVIII ranges from 438- 574 np. To annotate the population differences, mtDNA sequences are compared with (rCRS) Revised Cambridge Reference Sequence (Andrew *et al.*, 1999). It was discovered on the basis of Analysis of HVR1 and HVR2 of mtDNA that, around 500,000 years ago Human and Neanderthal split took place (Krings *et al.*, 1997) and it was also hypothesized that there is no contribution of Neanderthal in modern human mtDNA gene pool (Krings *et al.*, 1997, 1999; Ovchinnikov *et al.*, 2000). The first phylogenetic tree for mtDNA based on Restriction Site Polymorphism was constructed by Cann *et al* in 1987 and the maternal most recent common ancestor was called

Mitochondrial Eve originated around 200,000 years ago in Africa (Cann *et al.*, 1987). Like Y-STRs, being haploid markers mtDNA polymorphisms are best candidate for phylogenetic and population origin investigations including tracing human migrations. Such polymorphisms are geographically dependant as the mutations accumulated in mtDNA lineages during the course of time (Ingman *et al.*, 2001). These maternally inherited variants constitute haplotypes. A specific set of SNPs defines a haplogroup and

various haplotypes belong to same haplogroup on the basis of shared haplogroup defining SNPs (Torroni *et al.*, 1996). Haplogroup L0 - L7 are considered to be the most ancient haplogroups of African origin (Quintana *et al.*, 1999) while the haplogroup L3 is considered to be the first common ancestor of M and N haplogroup which later evolved into many variants during and after out of Africa human migration (Richards *et al.*, 2006; Chen *et al.*, 1999) A generalized sketch of human migration as reflected in mtDNA is provided in Fig.4.

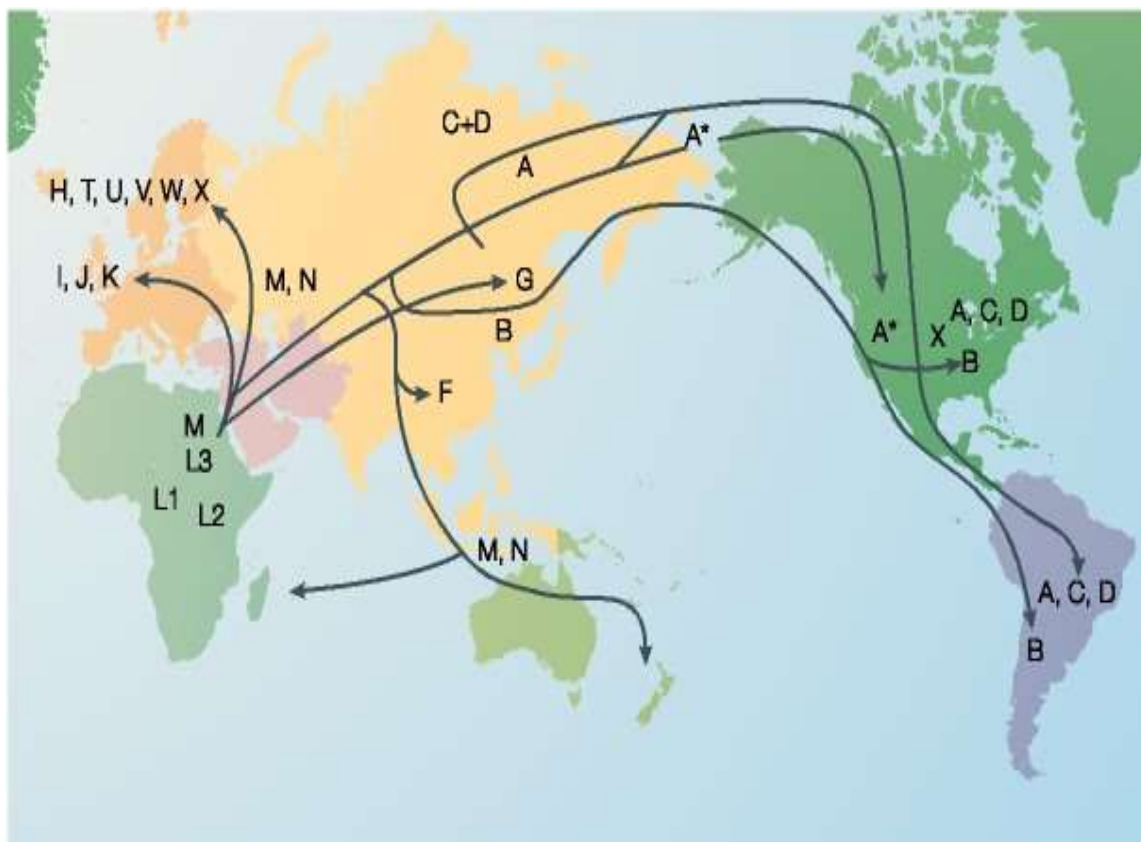


Fig.4. An Overview of worldwide distribution of mtDNA haplogroups (Schriver & Kittles, 2004).

2.6.2 mtDNA Haplogroups in South Asia

2.6.2.1 Haplogroup M

Macrohaplogroup M, along with its sibling N, account for all non-African mtDNAs.

This haplogroup originated during the human migration from Africa 57-75Kya (Chandrasekar *et al.*, 2009). An age estimate of 49,400Ky (95% CI; 39,000-62,200 years) is provided by Soares *et al.* (2009) in south Asia and in East Asia 60,600Ky (95% CI; 47,300-74,300 years). Other approximations include 55-73Ky for the lineage among African populations (Chen *et al.*, 1995) and 69.3 ± 5.4 Ky among Chinese populations (Kong *et al.*, 2003). Due to its significant contribution and distribution within the Indian population i.e. 60-70%, the haplogroup M is considered to be a south Asian lineage (Chandrasekar *et al.*, 2009; Disotell, 1999), but lower frequencies of this haplogroup has been estimated in Central Asia and western Eurasia. It is believed to have arrived in the Indian subcontinent via the Southern Route migration from Africa (Disotell, 1999; Macaulay *et al.*, 2005; Torroni *et al.*, 2006; Chandrasekar *et al.*, 2009; Kumar *et al.*, 2009). Sub-group M7 is a common lineage found in East Asian populations such as Korean-Chinese, the Han (Beijing) of China, Mongolians, Koreans (Jin *et al.*, 2009) and Japanese (Asari *et al.*, 2007). The Hazara, Balouch and Pathan populations of Afghanistan exhibit 15%, 13.3% and 7.1% frequencies respectively of this haplogroup (Whale *et al.*, 2012). within the Indian subcontinent its Frequencies have been found ranging from 26%-64% among different caste and tribal populations (Kivisild *et al.*, 1999; Quintana-Murci *et al.*, 2004) while the frequencies found within the Afghan populations seem to resemble

frequencies found elsewhere in Central Asia (Whale *et al.*, 2012).

2.6.2.2 Haplogroup N

The second macrohaplogroup to have diverged from the African lineage L3 is haplogroup N. Its age has been estimated at 64.6 ± 6.8 Ky (Kong *et al.*, 2003), 61,900 YBP in west Eurasia (95% CI; 49,200-75,000 years), 71,200YBP in South Asia (95% CI; 55,800-87,100 years) and in East Asia, 58,200YBP (95% CI; 44,100-72,800) (Soares *et al.*, 2009). It is considered to be the ancestor of many haplogroups found in Europe, Middle East, Asia and the Americas (among the Amerindians). The origin of this lineage occurred soon after or probably during the migration out of Africa, and is typically considered a southwest Eurasian lineage (Kivisild *et al.*, 1999; Quintana-Murci *et al.*, 2004; Nasidze *et al.*, 2006, 2007). This haplogroup is comparatively common in western Eurasia and is also present in Europe. A frequency of 5.3% has been reported in eastern Crete (Martinez *et al.*, 2008) while the combination of haplogroups N, I, W and X constitute approximately 9% of the Finnish population (Hedman *et al.*, 2007). This haplogroup is also found in the Near East and northeast Africa nearly ~13% in Egypt, ~10% in Israel, Syria and Jordan, ~5% in Iraq and ~23-44% among Iranian populations (Nasidze *et al.*, 2008). The Hazara population of Afghanistan exhibits a frequency of 7.5% and the Tajiks 10.5% of this haplogroup (Whale *et al.*, 2012). Elsewhere in Central Asia and western Eurasia, the frequency of N haplogroup ranges from 2.3% in the Tajiks of Tajikistan (Derenko *et al.*, 2007) to 20% in the South Caspian region in Iran (Comas *et al.*, 2004). The greater frequencies of this haplogroup appear to occur in the more western

populations rather than in Central Asian or South Asian. Haplogroup M is affluent in South Asia, however haplogroup N appears to be lacking from the mtDNA landscape with frequencies of 2.6% and 2.9% in the Brahui of Baluchistan and Gujarati of northwestern India, and 3% in both Pakistani and Makrani populations and 7.7% within the Han Chinese population (Yang *et al.*, 2011). The frequencies exhibited in Central Asia are similar to those found in the Hazara and Tajiks with Uzbeks harboring 7.1% (Quintana-Murci *et al.*, 2004) and the Turkmen population 10% (Comas *et al.*, 2004).

2.6.2.3 Haplogroup R

As a descendant of the macrohaplogroup N, the haplogroup R also diverged soon after the modern human migration out of Africa. Along with the macrohaplogroups M and N, R is one of the founder lineages for Eurasian settlement ~60-65Kya (Torroni *et al.*, 2006). It has an estimated age of 59,100 years in west Eurasia (CI; 47,100-74,100 YBP), 66,600 years in South Asia (CI; 52,600-81,000 YBP) and 54,300 years in East Asia (CI; 41,200-67,800 YBP) (Soares *et al.*, 2009), and 62.3 ± 6.3 Ky (Kong *et al.*, 2003). This haplogroup is a typical west Eurasian and South Asian lineage primarily due to its early divergence from haplogroup N in this region and can be characterized by a MboII site gain at np 12704 caused by a transition at np 12705. , The haplogroup R makes up for almost less than 3% of the maternal gene pool of Finnish population (Hedman *et al.*, 2007). It has often been recorded in Central Asia, South Asia and western Eurasia; however its distribution is not uniform throughout these regions (Whales *et al.*, 2012).

The Karakalpak population presents a frequency of 10% (Comas *et al.*, 2004), while in the Gujarati population of northwest India it appears in 8.8% of mtDNAs and in 1.8% of Georgians (Quintana-Murci *et al.*, 2004). Within the Afghan populations, the Pathans exhibit 28.6%, Tajiks 15.8% and the Hazara 7.5% of this haplogroup. However, the greatest frequency was found in the Uzbeks at a frequency of 20%, while in the south Caspian region, haplogroup X was found in 2.4% of Persians and 9.5% of Mazandrians. The presence of these three lineages within the Afghani populations and the adjacent populations from Iran, Central Asia and the Indian Subcontinent may be attributable to this region being the initial territory where haplogroups M, N and R settled following the human dispersal from Africa. Despite each lineage sharing similar coalescent ages, haplogroup M is prominent among South Asian populations, particular among those in southern India in Andhra Pradesh where its frequency has been recorded at 64% (Whales *et al.*, 2012). While the overall frequency of 61% for this haplogroup was recorded in Pathan Ethnic group of Pakistan (Rakha *et al.*, 2011).

An updated mtDNA haplogroup phylogenetic tree(Fig.5) is being used for population haplogrouping and classification on the basis of origin (Van *et al.*, 2009).

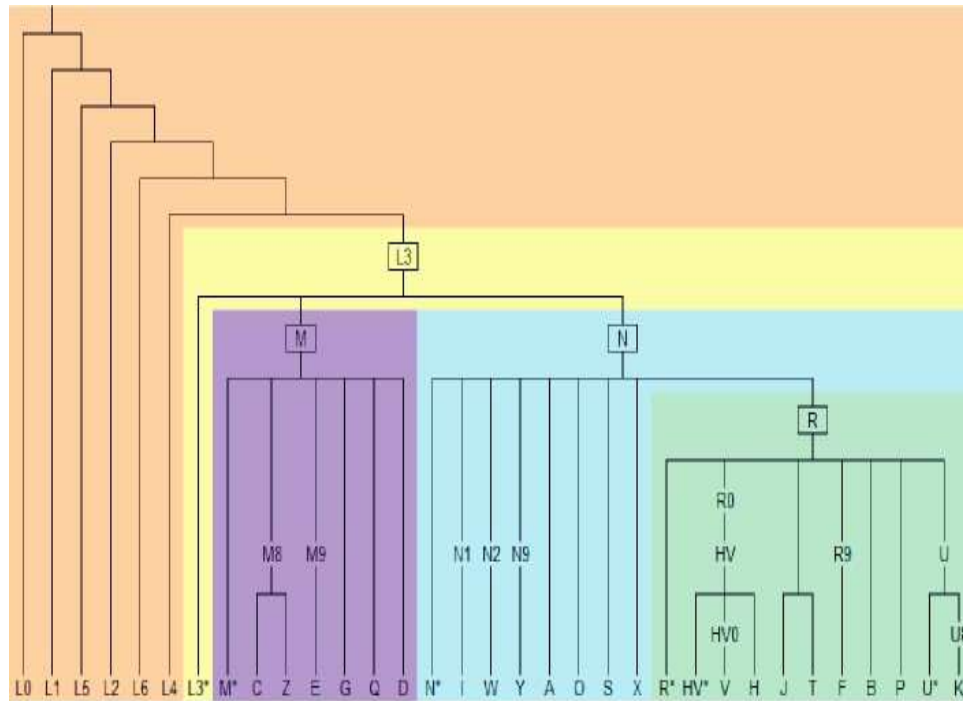


Fig.5. An Overview of the mtDNA haplogroups (Van *et al* 2009).

2.7 The Y-Chromosome

The human Y-chromosome is being considered as an evolutionary artifact of the X chromosome. Besides its ability to define gender, it has some other practical significance by carrying many functional genes essential for normal male development. The X and Y-chromosomes are considered to be the true homologues roughly around 300 million years ago (Lahn *et al.*, 2001) after successive mutations especially deletions reduced the Y chromosome to the size of 50 Mega bases, but sequence homology between to chromosomes still persist up to some extent.

2.7.1 The Structure of the Y-Chromosome

The smallest among all the chromosomes of human genome is the Y-Chromosome. Its average size is roughly around 60Mb. 24Mb of which comprises the euchromatin region and around 30Mb comprises the heterochromatin region, collectively they are referred to as the MSY (male specific region) or NRY (non-recombining region) constituting about 95% of the Y chromosome (Butler, 2003). Its large number of short tandem repeats (STRs) and single nucleotide polymorphisms (SNPs) contribute to the formation of a powerful and informative haplotyping system that has long been used in human identification (Jobling *et al.*, 2001). A sketch of the Y-chromosome is given in Fig. 6.

Several genealogical and evolutionary studies besides Forensic DNA typing have made use of continuously growing reliable Y- Short Tandem Repeats (STRs) and Single Nucleotide Polymorphism (SNPs) markers (Butler, 2003). Almost 99.99% of the Y-chromosome is inherited as a single unit from father to the son in paternal fashion (Butler, 2003). The entire chromosome remains intact and all its markers along its entire length remain strongly linked to each other. In this way a single chromosome acts as a single haplotype that can be used to trace the human migration and evolutionary events (Butler, 2003). The majority of Y chromosome exhibits lack of recombination and as a result of this attribute the haplotypes pass intact generation through generation and helps to preserve a simple historical record of the descent and phylogeny (Helena *et al.*, 2007). Unlike rest of the chromosomes mutation is the only source of variability in this case.

Due to small effective population size, i.e. one quarter to that of autosomes and one third to the X-chromosome, comparatively low sequence diversity is also expected on Y-chromosome than on any other (Hammer, 1995; Thomson *et al.*, 2000).

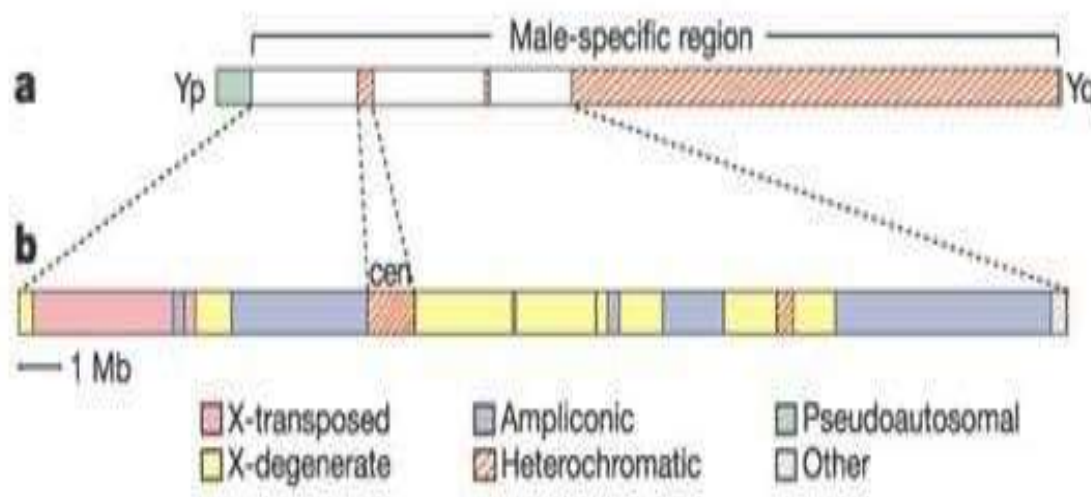


Fig.6. Structure of the various regions of the human Y-chromosome (Skaletsky *et al* 2003).

2.7.1.1 Pseudoautosomal regions (PARs)

The telomeric proximities positioned at the distal part of the short arm (Yp) and long arm (Yq) are called Pseudoautosomal region 1 (PAR1) and Pseudoautosomal region 2 (PAR2) respectively. The approximate length of PAR1 is 2.5Mb while PAR2 is less than 1Mb in length (Skaletsky *et al.*, 2003). The Y chromosome recombine with its counterpart the X chromosome only at these regions during meiotic division in a similar fashion to that of the autosomal loci (Butler, 2003).

2.7.1.2 Heterochromatic region

The 30 Mb regions on (Yq) long arm of Y chromosome comprising two repeat sequences i.e. DYZ1, DYZ2, alphoid sequences and several satellite sequences, is called heterochromatic region on Y chromosome (Skaletsky *et al.*, 2003; Gusmao *et al.*, 1999). These sequences are clustered tandemly near centromere and plays important role in spermatogenesis (Gusmao *et al.*, 1999). In addition to this, the Y-chromosome also carries two DYZ1 repeat fragments each 2.1 Kb in length and two other fragments i.e. the Y-specific (YS) and the non-Y-specific (NSY) (Gusmão *et al.*, 1999) each 3.4Kb in length. These fragments consist of a single array of pentameric satellite sequences with the repeat sequence of 5' TTCCA³' in 800 to 4000 copies (Gusmão *et al.*, 1999).

2.7.1.3 Male specific region (MSY) or non-recombining region (NRY) of Y- chromosome

Almost 95% of the Y chromosome is responsible for sex determination in males called NRY (Non recombining region on Y chromosome) or MSY (Male specific region on Y chromosome) region. It consists of a mosaic arrangement of Heterochromatic and Euchromatic regions (Skaletsky *et al.*, 2003). The euchromatic region is further divided into three categories, i.e. the X-Degenerative Region, the X-Transposed Region, and the Ampliconic Region. Ampliconic region carries the total of 156 transcription units (Skaletsky *et al.*, 2003).

2.7.1.4 X-Degenerative Region of Y chromosome

The 20% of the euchromatin consist of eight sequence blocks on Yp and Yq comprising almost 8.6Mb of sequences, called X-degenerative region on MSY. Almost 27 X-linked single copy genes are located on this region. Out of which 13 genes are nonfunctional Pseudo genes while remaining 14 are Y linked functional genes. Some of which are homologous to X linked genes (Skaletsky *et al.*, 2003).

2.7.1.5 X-Transposed Region of Y chromosome

X-transposed region of 3.4 Mb on the short arm of Y chromosome (Yp) comprises almost 15% of the MSY euchromatin and occurs in two sequence blocks (Skaletsky *et al.*, 2003). 99% of the X-transposed region sequences are homologous to the X chromosome long arm (Yq) sequences. This region possesses lowest gene density and highest interspersed repeat density and it doesn't recombine unlike PARs (Skaletsky *et al.*, 2003).

2.7.1.6 Ampliconic Region on Y chromosome

30% of the total MSY euchromatin constitutes the Ampliconic region of 10.2Mb, which consists of seven sequence hunks on both the short arm (Yp) and the long arm (Yq). These sequences possess the highest gene densities of the three classes of the MSY euchromatin and the lowest gene density of LINE1 and interspersed repeat elements (Skaletsky *et al.*, 2003). Nine protein coding gene families on MSY have been identified.

Among them, XKRY, PRY, VCY and HSFY have two copies, BPY2 exists in three copies while DAZ and CDY exist in four copies and six copies exist of RBMY gene. All of them are involved in male fertility.

Function	Copy number	Genes	PAR	Genes	Copy number	Function
Transcription factor - sex determination	1	<i>SRY</i>	1	<i>RPS4Y</i>	1	Protein of small ribosomal subunit
				<i>ZFY</i>	1	Zinc finger transcription factor
Testis transcript 1	m	<i>TTY1</i>	2	<i>PCDHY</i>	1	Protocadherin - cell adhesion
Cyclin B binding protein	m	<i>TSPY</i>	3	<i>PRKY</i>	1	Ser/Thr protein kinase
				<i>AMELY</i>	1	Tooth enamel formation
Protein tyrosine phosphatase	m	<i>PRY</i>	4A			
Testis transcript 1	m	<i>TTY1</i>	4B			
Testis transcript 2	m	<i>TTY2</i>				
Cyclin B binding protein	m	<i>TSPY</i>				
						Centromere
				<i>USP9Y</i>	1	Deubiquitinating enzyme
				<i>DBY</i>	1	DEAD-box - RNA helicase
				<i>UTY</i>	1	TPR-motif
				<i>TB4Y</i>	1	Actin sequestration
			5			
				<i>VCY</i>	2	Variable charged protein
Chromodomain protein	m	<i>CDY</i>				
Membrane transport protein	m	<i>XKRY</i>		<i>SMCY</i>	1	Transcription factor
				<i>EIF1AY</i>	1	Translation initiation factor
Protein tyrosine phosphatase	m	<i>PRY</i>		<i>RBMY</i>	30	RNA-binding protein
Testis transcript 2	m	<i>TTY2</i>	6			
				<i>RBMY</i>	30	RNA-binding protein
RNA-binding protein	4	<i>DAZ</i>				
Basic protein	m	<i>BPY2</i>				
Protein tyrosine phosphatase	m	<i>PRY</i>				
Chromodomain protein	m	<i>CDY</i>				
			7			Heterochromatin
Y-chromosome genes not found on the X			PAR			Y-chromosome genes with homologs on the X

Fig.7. Protein coding genes on the Male specific region on Y chromosome with homologues on the X-chromosome (right side) and genes not found on the Xchromosome (left side) (Skaletsky *et al.*, 2003).

2.7.1.7 Palendromic DNA sequences

There are eight long palendromic sequences in ampliconic region on Yq, denoted by P1-

P8 ranging in length from 9Kb to 1.45KB. These palindromes are separated by 2Kb to 170Kb spacer sequences (Rozen *et al.*, 2003). P1 is the largest palindrome of 2.9Mb. It consists of two secondary palindromes P1.1 and P1.2 within it, each 24Kb in length. The total length of all the eight palindromes is approximately 5.7Mb that is about 25% of the total MSY euchromatin (Skaletsky *et al.*, 2003).

2.7.1.8 Sex determining region on Y chromosome (SRY)

In the euchromatic region of the MSY, there is a region adjacent to the pseudoautosomal region on the short arm of Y chromosome called the sex determining region (SRY). It actually controls the male sexual development by encoding for (TDF) the testis determining factor (Sinclair *et al.*, 1990).

2.7.1.9 Amelogenin gene (AMEL)

The Amelogenin gene (AMELY) on the Y-chromosome and its homologue (AMELX) gene on the X-chromosome, plays important role in tooth bud growth by enamel production (amelogenesis) and dentine development (dentinogenesis) (Butler, 2005). This locus is primarily used in gender differentiation of male/female mixed samples which is not only a practical tool but of requirement of extreme importance in forensic caseworks particularly in cases of sexual assault, skeletal remains and old blood blots which require swift, sensitive, reliable and accurate means of exploration and analysis (Butler, 2005).

2.7.2 Polymorphisms on the Y-chromosome

In order to trace the human migration, evolutionary history and population admixture, the paternally inherited Non recombining region on Y chromosome (NRY) has extensively been used since last two decades. This region on Y chromosome holds various polymorphisms with varying mutation rates that could be used as a powerful tool for scientists to achieve their research goals. The genetic markers on NRY region of the Y-chromosome are physically linked and are transmitted to the males of next generation i.e. in patrilineal fashion. This linkage and lack of recombination results in the lack of independent assortment of these markers and increased susceptibility to genetic drift due to small effective population size as compared to autosomes. Drift enhance the differentiation and discrimination between Y-chromosomes in different populations and is useful to investigate the past events in a specific population. But genetic drift influences the haplotype frequency in a population over time. Clustering pattern is also influenced by the behavior of males bearing Y-chromosome, e.g. Patrilocality (Murdock, 1967; Burton, 1996). Patrilocality is mostly practiced in South Asia (Kayser *et al.*, 2001).

2.7.2.1 SNPs the Bi-allelic Markers

The most abundantly occurring markers are the Bi-allelic markers which include the ALU insertion (YAP-DYS287) and considerable number of SNPs (single nucleotide polymorphisms) (Underhill *et al.*, 1996). SNPs are SNPs are the single nucleotide change

in the base-pair sequence as consequence of substitution, transversion or transition. More than 33,000 Y-SNPs have been discovered and used so far for population phylogenetic studies and forensic investigations (Hallast *et al.*, 2015). The most potential use of SNPs is the construction of haplogroups in population genetics and evolutionary studies. The most important is the appropriate and careful selection of SNPs, as different SNPs can define the same haplogroup (Sanchez *et al.*, 2004).

2.7.2.2 Alu Polymorphisms on Y chromosome (YAP)

Allele variants are created by mutations as a consequence of insertions or deletions of usually one nucleotide at specific loci along the Y-chromosome. One such important insertion is Alu polymorphism on Y chromosome (YAP) consisting of approximately 300bp (Hammer *et al.*, 1994).

2.7.2.3 Multi-allelic markers (Micro and minisatellites)

Microsatellites are sequences with repeat units of 2 to 7 BP in length and mostly they are short tandem repeats (STRs) (Ellegren, 2000; Imad *et al.*, 2014). The type of DNA sequence is defined by the length of core repeat and number of repeat units (Butler, 2012). They may be di, tri, tetra, Penta or Hexa nucleotide repeats. Penta and Hexa nucleotide repeats are considered as microsatellites. STRs have potential to be used to study evolution and migration foot prints as well as for resolution of medico legal cases (Walkinshaw *et al.*, 1996).

Several hundreds of microsatellites or Y-STRs of potential importance in forensic caseworks and evolutionary studies have been identified on the Y-chromosome (Gil *et al.*, 2012). Number of repeats is highly variable among individuals so are best candidates for population genetic studies (Budowle, 1995; Butler *et al.*, 2009; Mohammed and Imad, 2013).

Similarly minisatellites are also present on Y chromosome that varies in length from 10 - 60 base pairs (Gusmão *et al.*, 2005). Number of repeats defines the allelenomenclature at specific locus. Changes occur occasionally in the number of repeats of these STRs or minisatellites. The most probable cause of intra allelic mutations in microsatellite is the replication slippage (Jeffreys *et al.*, 1998). Other than mutation, selection is another source of haplotype diversity in population. Paternal lineage can be traced by tracking these changes (Gusmão *et al.*, 2005). The rate of mutation is high in minisatellites loci, approximately equals to 6-11% per generation, while in microsatellites it is nearly equals to ~0.2% per generation for Y-STRs. On the other hand the mutation rate for SNPs is significantly low (Dupuy *et al.*, 2004). Multi-allelic markers have proved to be extremely useful in differentiating Y-chromosome haplotypes resulting in fairly high resolutions (Butler, 2003). Major fraction of the Y-STR loci is located on the long arm (Yq) of the Y-chromosome. Almost 25.3% of STRs are at Yq11.221, 16.6% of the STRs are at Yq11.222 and almost 18.4% of Y STRS are positioned at Yq11.223 (Hanson *et al.*, 2006). 22.1% are found on the short arm of the Y-chromosome at Yp11.2 and three loci

DYS716, DYS707 and DYS631 are located in the centromeric segment (Hanson *et al.*, 2006). There are several advantages of using Y-STR markers over autosomal markers with a few limitations.

2.7.2.4 Unique and recurrent event polymorphisms

Y chromosome markers are classified on the basis of their mutation rates. e.g. recurrent and unique event polymorphism (Hurles and Jobling, 2001). SNPs, YAP and indels exhibit low mutation rates while microsatellites and minisatellites exhibit high mutation rate and generally called “unique event polymorphisms” (UEPs). These are rarely occurring polymorphisms that are assumed to be population specific and occurs at specific loci as a consequence of evolution (Hurles and Jobling, 2001). Binary polymorphisms of distinctive origin (haplogroups) are pooled into monophyletic composite haplotypes and are phylogenetically branched off from the single parsimonious tree. Hurles and Jobling first time estimated the human ape divergence time on the genetic evolutionary clock. And the TMRCA (Time to most recent common ancestor) of existing human Y chromosome has been estimated to be between 50,000 and 200,000 years ago (Hurles and Jobling, 2001).

2.7.3 Applications of Y-chromosomal polymorphism

Several hundreds of the Y-STRs have been discovered so far are available as a traditional tool for forensic and population genetic investigations but only those with high genetic diversity and variance are proved to be more useful in such cases (Hanson *et al.*, 2006).

Rapidly mutating Y-STRs have ability to differentiate and discriminate between brothers in more than 60% of cases (Ballantyne *et al.*, 2012).

2.7.3.1 Y polymorphism, a tool in Forensic investigations

Y-chromosomal polymorphisms, primarily the STRs have been a useful discerning tool in forensic studies specifically in cases of sexual assault. They have been playing a substantial role in generation of valid genetic profiles acquired from male suspects of gang rapes, male /female or male/male sexual assault cases (Hanson *et al.*, 2006).

2.7.3.2 Paternity testing casework

The uniparental inheritance along the patriline makes the Y chromosome and Y-STRs useful for tracing the paternity of the male offspring. For this purpose the Y -DNA from any male relative in the lineage can be helpful (Gusmao *et al.*, 1999). A key feature in forensic investigation and paternity testing is the accurate construal of genetic profiles considering the probability % rates of possible STR mutations that could influence the inclusion or exclusion of paternity of an alleged father (Kayser *et al.*, 2001). Different sets of Y-STR markers are being used for this purpose. Y-STR haplotypes characterize information from the non-coding lineage of the Y-chromosome shared among several males along the paternal line. They do not provide individualization unlike that of autosomal STR loci and proved to be more valuable and conclusive in paternity tests of male subjects where autosomal and other STRs couldn't work (Gusmão *et al.*, 2005).

2.7.3.3 Implication of genealogical and Evolutionary investigations

Several sets of minisatellites, microsatellites or STRs and biallelic markers are the best candidates for evolutionary and genealogical investigations due to their typing simplicity and high level of diversity. Network analysis of the haplotypes based on these markers has proved to be more informative for evolutionary and phylogenetic studies (Gusmão *et al.*, 1999). Several hundreds of SNPs and Y-STRs are being used to investigate the biogeographical ancestry of different populations and sub-populations (Underhill *et al.*, 2001; Ali *et al.*, 2003).

Due to very low mutation rates and lack of recombination, SNPs are in strong agreement for the construction of maximum parsimony tree (Jobling *et al.*, 2001).

2.7.4 Y-Haplogroups

Human Y-chromosome haplogroups are defined by Y-chromosomal polymorphism specifically by SNPs. Each haplogroup represents the branch of Y-chromosome phylogenetic tree. The patrilineal most recent common ancestor of all humans is considered to be the “The Y-chromosomal Adam”.

A system of naming Y DNA haplogroups had been developed by (YCC) YChromosome Consortium. Initially 18 Y-haplogroups were defined by YCC but later with the discovery of some more SNPs, their number was increased to 20, ranging from A-T. The frequency and occurrence of these haplogroups varies in different geographical regions and some haplogroups are found to be population specific as well.

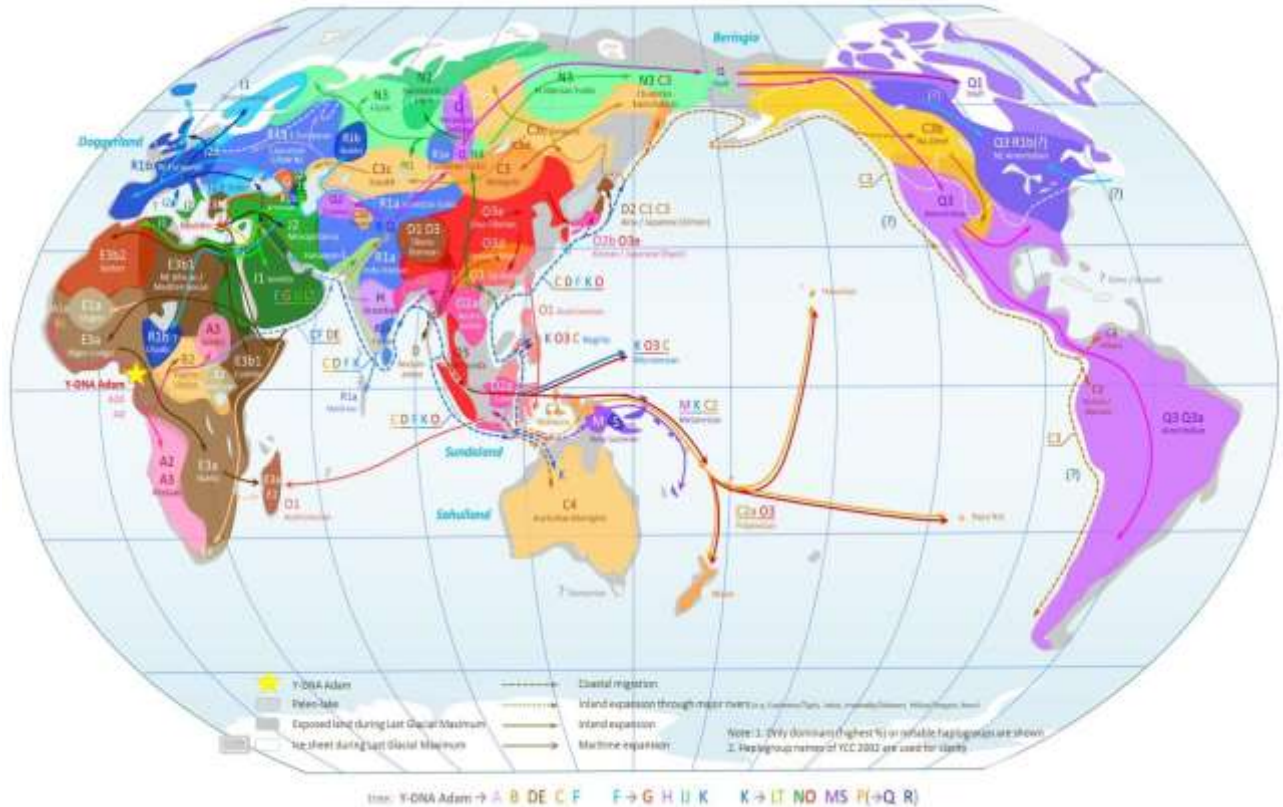


Fig.8. World map of Y- DNA Haplogroups around the globe with possible migration routes.([https:// en.wikipedia.org](https://en.wikipedia.org))

2.7.4.1 Y-Haplogroups in South Asia

South Asia includes present day Pakistan, India, Afghanistan, Bangladesh, Nepal, Bhutan, Maldives and Sri Lanka. Several Y-DNA haplogroups including C, F, G, H, J, L, O, P, Q, R1a, R1b, R2 and T prevailed with varying frequencies in different ethnic groups inhabiting South Asia.

2.7.4.1.1 Haplogroup R

Haplogroup R is said to be originated around 30,000 years ago in Central Asia as a branch of mega haplogroup P. There it divided into two branches, The R1a and R1b.

R1a moved towards Europe and R1b migrated towards South Asia. R1a is believed to be more likely instigated in South Asia and less probably in Eurasian steppes in the population of Kurgan culture practicing pit grave culture and horse domestication and the speakers of Indo European language. The high frequency of R1a haplogroup in south Asia is the consequence of Indo-Iranian migrations.

2.7.4.1.2 Haplogroup Q

The probable site of origin of this haplogroup is The South central Siberia or Altai Mountains around 17,000 to 31,700 years ago (Zegura *et al.*, 2004; Sharma *et al.*, 2007) and have been postulated to reach South Asia with Indo Aryan migrations. It is found in different populations of South Asian countries with varying frequencies.

2.7.4.1.3 Haplogroup F

This haplogroup has been found in south India and is considered to the parent of all other haplogroups (G-T) (fig.9). It prevails in almost 90% of the world" s population. Haplogroup G, H, IJ and K are considered to be the major sub-haplogroups of F. The probable place of its origin is either Eurasia around 48,000 years ago (Karafet *et al.*, 2008) or South Asia. According to other study it has been originated in Levant or the Arabian Peninsula around 50,000 years ago (Hammer *et al.*, 2002). Its descendant haplogroups depict pattern of expansion and radiation from South Asia or Middle East. And the presence of its sub clades in Africa might be the consequence of Back migration from south Asia or Southwest Asia towards Africa.

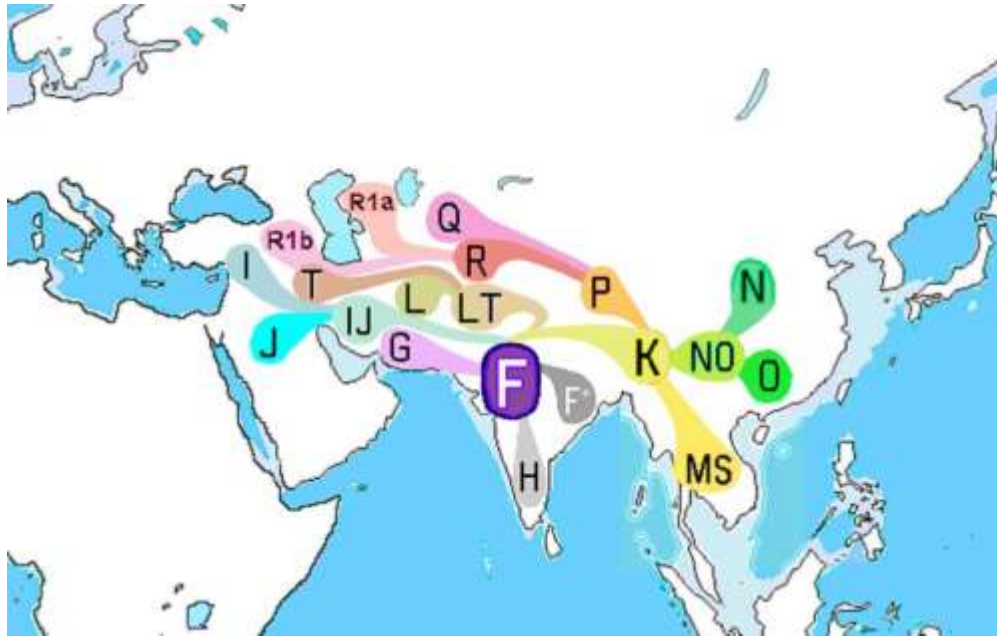


Fig.9. Y-DNA Haplogroup F and its descendent haplogroups from G to T
(<https://en.wikipedia.org>)

2.7.4.1.4 Haplogroup G, H, IJ

Haplogroup G was found to be originated approximately 17,000 - 21,000 years ago in the Middle East and is prevalent in several ethnic groups of Eurasia (Passarino *et al.*, 2002; Karlsson *et al.*, 2006) while haplogroup H is found primarily in South Asia. This haplogroup has been originated in South Asia around 30,000–40,000 years ago. Haplogroup I, is of European origin before Last Glacial Maximum from its ancestor haplogroup IJ which was said to be migrated there from middle East around 30,000–40,000 years ago. It bifurcated into I1 and I2 approximately 28,000 years ago (Battaglia *et al.*, 2009). Haplogroup J was derived from IJ in near east or West Asia around 42,000 years ago before present and it was scattered to India, Pakistan, Central Asia, Europe and

North Africa as a consequence of either Neolithic expansion or episodic migrations (Semino *et al.*, 2004; Shou *et al.*, 2010).

2.7.4.1.5 Haplogroup L

This haplogroup first descended from haplogroup K approximately 30,000 years ago. It is found in high frequencies in various South Asian populations including Pakistani and Indian populations. Three sub branches of this haplogroup: L1, L2 and L3, have been found in Irani and Pakistani populations while only L1 has been found in Indian population (Sangupta *et al.*, 2006).

2.7.4.1.6 Haplogroup O

This haplogroup was originated in East Asia approximately 35,000 to 40,000 years ago and expanded towards South East Asian populations (Yan *et al.*, 2011). Small percentage of this haplogroup is found in South Asian populations.

2.7.4.1.7 Haplogroup T

Previously it was known as K2 (Mendez *et al.*, 2011). It is said to be originated between Western and South Western Eurasian plate i.e. between Himalayas and Germania approximately 40,000 years ago (Hallast *et al.*, 2015). According to several other investigations, K2 or T has been originated in Asia and later migrated to North Africa (Underhill *et al.*, 2001, Cruciani *et al.*, 2002; Samino *et al.*, 2002; Luis *et al.*, 2004). It is present in low frequency in South Asian populations.

The area under investigation includes the major area of Peshawar valley and is the best representation of Pathan population of Pakistan. The area is unexplored land with respect to human biology, particularly molecular anthropology. Hence the PhD research project presented here was started to establish a base line for molecular anthropology with the objectives highlights given in the introduction section.

Chapter3

MATERIALS AND METHODS

3.1 Materials and Methods

Buccal swab samples were obtained from 374 unrelated healthy male donors. 75 each from Muhammadzai, Mohmand, Kakakhel Mian populations from Charsada District, and Yousafzai and Mohmand (74 sample) from Mardan District. All the unrelated donors who voluntarily donated their samples after properly signing the informed consent were included in this study. For obtaining the oral swab volunteers were asked to rinse their mouth before giving sample, in order to minimize the chance of contamination. Each individual was provided a collection cup with having 3mL of 5% sucrose solution. The purpose of sucrose was to induce the production of Ptyalin carrying saliva, in the buccal cavity. Ultimately sucrose was digested and loose cheek epithelial cells were mixed with the saliva after rinsing their mouth with the solution for 2-2.30 minutes. The resulting solution was collected in the collection cup. Samples were then stored at -20 °C before further processing.

3.2 DNA Extraction

Genomic DNA was isolated from buccal swab by using phenol chloroform method as described by Marisi and Sergio (2007).

The solutions and reagents were prepared as protocol given in Appendix: II.

Samples were transferred to the labeled 1.5mL eppendorf tubes. These tubes were then centrifuged at 7000 rpm for 5 minutes to pellet the buccal cells and debris. The

supernatant was poured off immediately to avoid pellet slippage. This step was repeated twice or thrice to get maximum quantity of pelleted cells.

In the next step, 300µL of cell lysis solution [10mM Tris, 0.5% SDS, 5mM EDTA (PH 8.0)] and 3 to 4 µL of Proteinase K (20 mg/ml) was added to each tube containing cell pellet. The mixture was vortexed at high speed for a few seconds to get the palette completely dissolved in the lysis buffer, and tubes were then incubated at 65°C for one hour. After incubation, 500 µL solution of Phenol chloroform in the ratio of 1:1 was added to each tube and was centrifuged at 10,000 rpm for 15 minutes. After that supernatant was transferred to the new eppendorf tube and 500 µL of Isopropanol (2propanol) was added and tubes were incubated for 10 minutes at -20°C. After that tubes were centrifuged at 12000rpm for 10 minutes and supernatant was discarded. 50 µL of 70% Ethanol was added to each tube and was centrifuged at 7000rpm for 5 minutes. This process was repeated after discarding the supernatant. Finally after palette washing with ethanol, tubes were kept inverted on clean absorbent paper overnight to let the palette dry. DNA palette was re-suspended in 50µl of TE buffer [12mM Tris (pH8) and 1mM EDTA] and was incubated for 10 minutes at 56°C to completely dissolve the palette.

3.3 Gel Electrophoresis

The isolated DNA from all the samples was run on agarose gel for qualitative analysis.

1% agarose gel was prepared by adding 1g of agarose in 100mL of TAE- buffer. The mixture was then heated in the microwave oven until it boiled. Solution was then cooled to 40°C and 12 µL of Ethidium bromide was added to it. The solution was then poured

into the gel cassette with the combs suspended in it to create wells and was kept at room temperature until solidified. After solidification, gel was transferred to the gel tank containing running buffer (TAE- buffer). 5 μ L of sample with 2 μ L of loading dye was loaded to each well. It was then run for 20 minutes at 80 Volts. The gel bands were visualized under the UV light to confirm the presence of genomic DNA in the samples (fig.10).

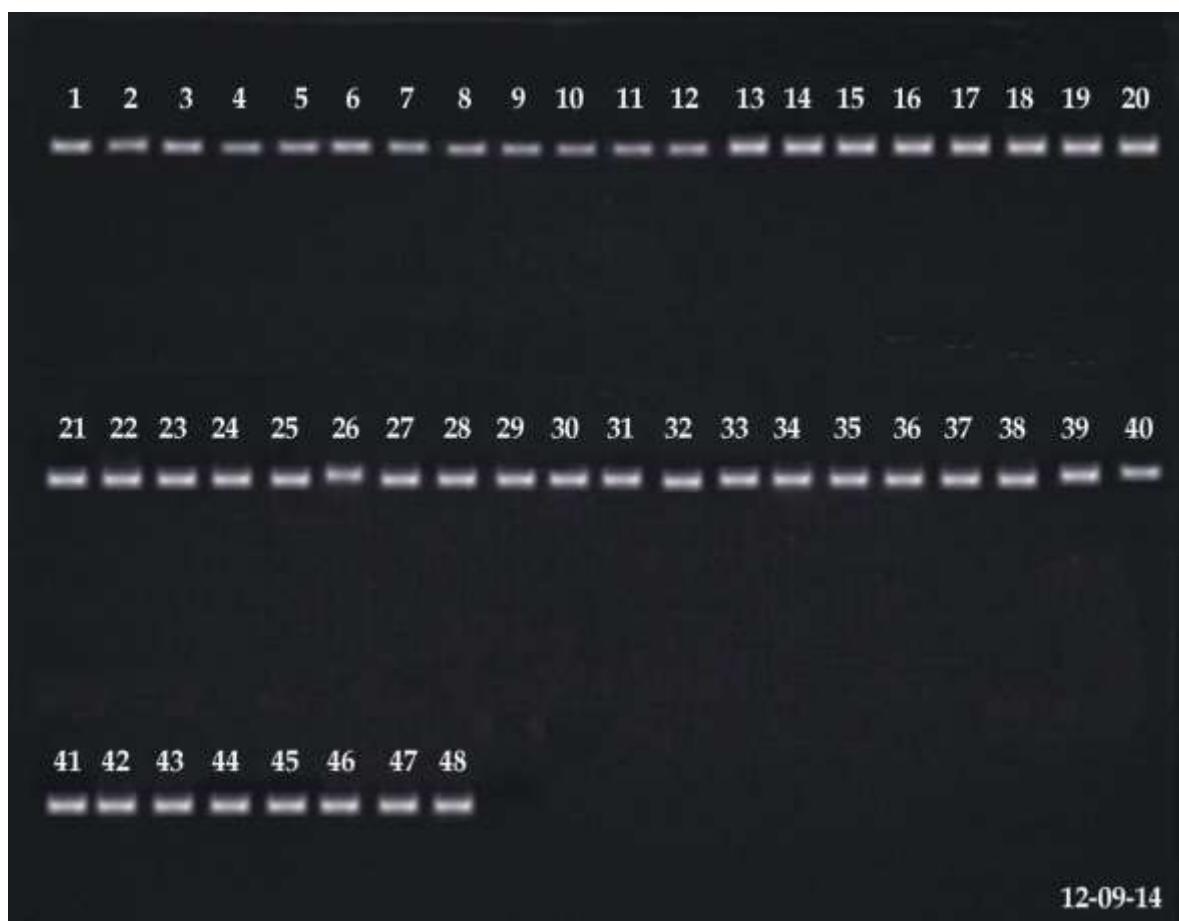


Fig.10. Agarose gel electrophoresis picture of the isolated Genomic DNA.

3.4 DNA Quantification

The DNA in each sample was quantified using Nano drop spectrophotometer before PCR amplification. And for those samples having highly concentrated DNA, serial dilutions were prepared before amplification.

3.5 Amplification of AMELY locus

All samples were amplified for AMELY locus confirmation before Y-STR amplification. 25 μ L of reaction mix was prepared for each 3 μ L of template DNA (table.1) and all the samples were amplified using the thermo cycling conditions given in fig.11

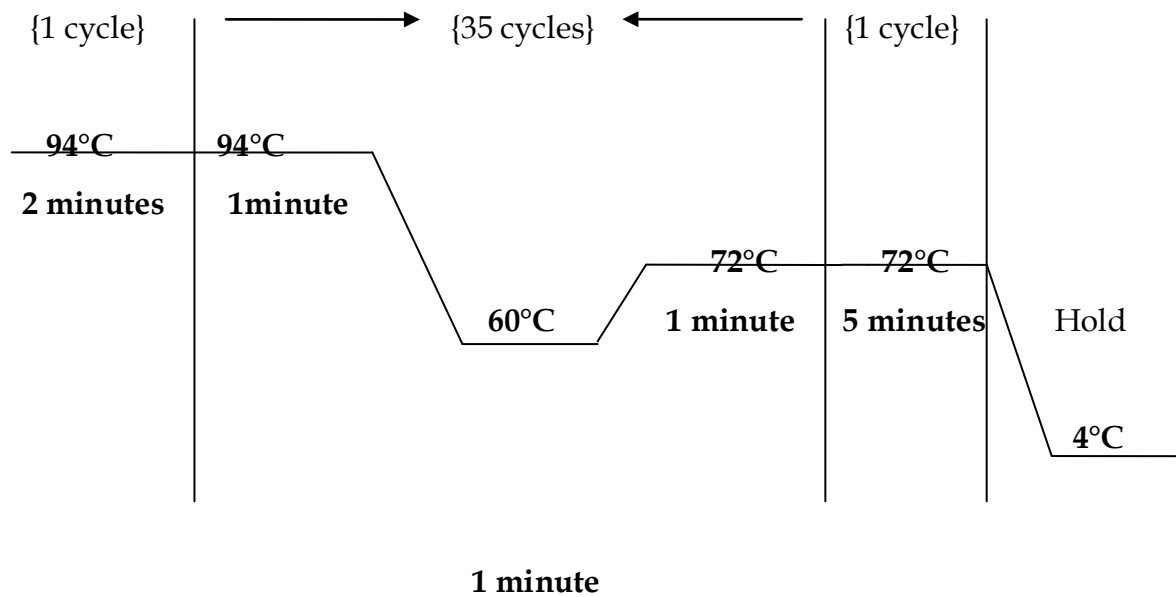
Table:1. Components of AMELY PCR amplification.

S. No	Components	Volume
1	Buffer	2.5 μ L
2	dNTPs	1 μ L
3	MgCl ₂	1 μ L
4	F primer	1 μ L
5	R primer	1 μ L
6	Plat Taq	0.2 μ L
7	5% Glycerol	5 μ L
8	Water, Amplification Grade	13.3 μ L
	Total volume of master mix	25 μ L

3.5.1 Thermo cycling parameters of the PCR

Thermal cycler was programmed with the following conditions as summarized in Fig.11.

- i. Initial hot start at 94°C for 2 minutes
- ii. Denaturation 94°C for minute (35 cycles)
- iii. Annealing at 60°C for 1 minute
- iv. Extension at 72°C for 5 minutes
- v. Final extension was performed at 72°C for 5 minutes
- vi. Storage soak indefinitely at 4°C



∞

Fig.11. Thermo cycling conditions of AMELY amplification.

3.6 Polyacrylamide gel electrophoresis (PAGE)

All the PCR products were run on Polyacrylamide gel to confirm the efficiency of PCR amplification. Polyacrylamide gel was prepared by adding 4.8mL dH₂O, 0.65mL 10X TBE, 975 µL of 40% Acryl mix, 16µL of 25% APS and 16µL of TEMED.

All the samples except two were found AMELY positive for further STR amplification with no risk of deletion of EMELY locus along with four important Y-STR loci i.e.

DYS570, DYS456, DYS576 and DYS481. A representation picture of PAGE results is provided in Fig.12.

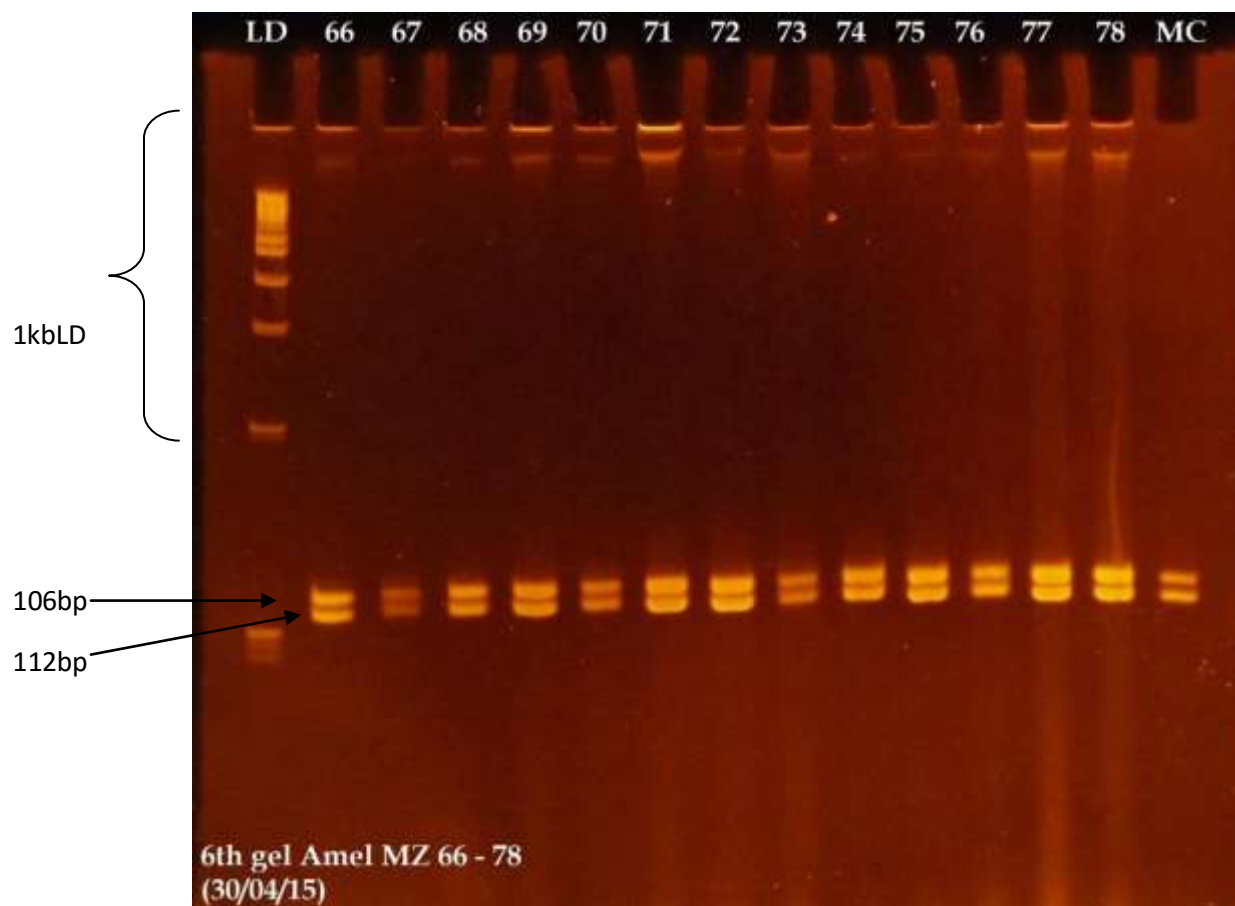


Fig.12. Polyacrylamide Gel Electrophoresis photograph of AMELY PCR product. LD: Ladder and MC: Male control

3.7 Y-STR amplification

Total of 374 samples from five different Pathan populations of Charsada and Mardan district were amplified for 23 Y chromosomal loci including (DYS576, DYS389I, DYS448, DYS389II, DYS19, DYS391, DYS481, DYS549, DYS533, DYS438, DYS437, DYS570, DYS635, DYS390, DYS439, DYS392, DYS643, DYS393, DYS458, DYS385ab, DYS456,

GATAH4) using Promega PowerPlex® Y23 System (PPY23, Promega Corporation, Madison, WI). Relative positions of 23 STRs on Y chromosome are provided in Fig.13.

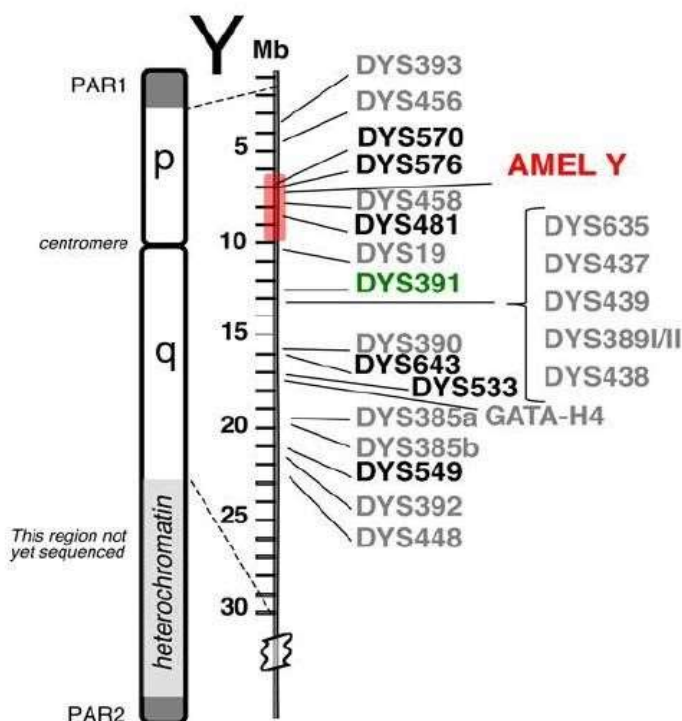


Fig.13. Relative positions of the 23 Y-STR loci used in the Promega PowerPlex® Y23 System.

3.7.1 Procedure

All the Pre-amplification components were thawed just prior to use. All the tubes were centrifuged and vortexed for 15 seconds before each use. Number of reactions was determined, including positive and negative control and 1 or 2 reactions were added to that number. PCR amplification mix was prepared by combining all the components of the kit using protocol established by Promega PowerPlex® Y23 System given in the table.2.

PCR Amplification mix was vortexed for 5-10 seconds and after that 9.4 μL of PCR was transferred to each labeled 0.2mL tubes. To each tube containing PCR amplification Mix, 1.5 μL of 0.5ng/ μL DNA template was added.

Table: 2. Components of Promega PowerPlex® Y23 amplification system.

Component	Volume per Reaction
Promega PowerPlex® Y23 5X. Master Mix	1.25 μL
Promega PowerPlex® Y23 10X Primer Pair Mix	0.625 μL
MgCl ₂	0.6 μL
Water, Amplification Grade	7 μL
Total Volume of Amplification Mix.	9.475 μL
Template DNA	0.5ng/ μL of 0.5 μL
Total Reaction volume	11 μL

3.7.2 Thermo cycling parameters of the PCR

Thermal cycler was programmed with the following conditions (Fig.14).

- i. Initial hot start at 96°C for 2 minutes
- ii. Denaturation 94°C for 10 seconds (28 cycles)
- iii. Annealing at 61°C for 1 minute
- iv. Extension at 72°C for 30 seconds
- v. Final extension was performed at 60°C for 20 minutes
- vi. Storage soak indefinitely at 4°C

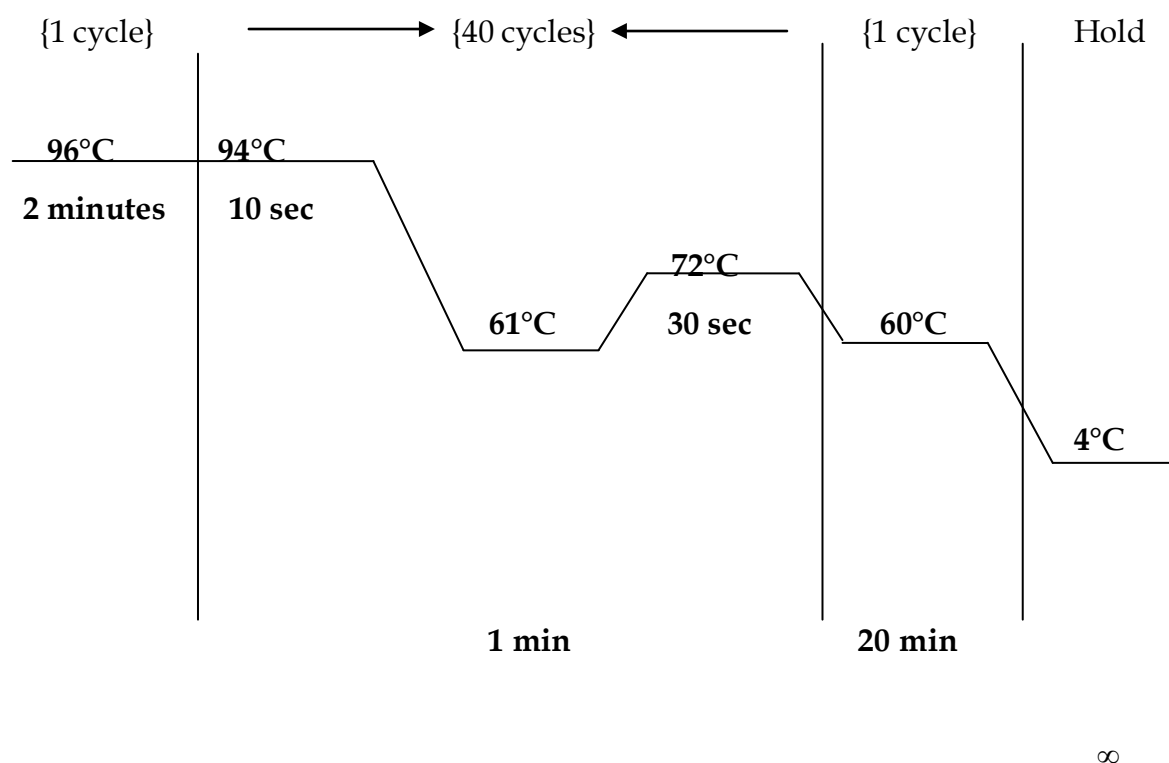


Fig.14. Thermal cycling protocol for PPY²³ System using Eppendorf mastercycler ep Gradient S thermal cycler.

3.8. Polyacrylamide Gel Electrophoresis

The next step was to check to make sure the PCR worked, before moving on to the next step. For this, Polyacrylamide gel electrophoresis (PAGE) was used. (Fig.15)

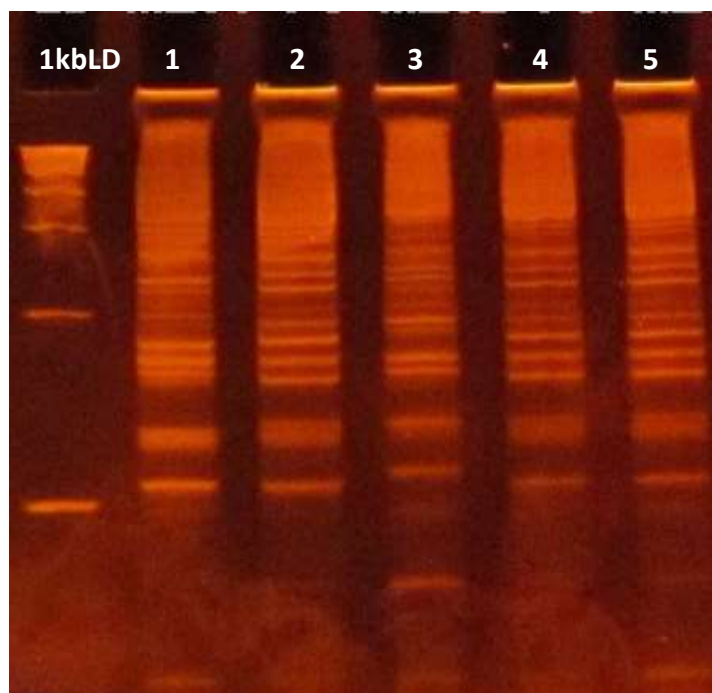


Fig.15. Polyacrylamide gel electrophoresis photograph of 23 Y-STR loci amplified with PPY²³ System.

3.9 Capillary Electrophoresis

After confirmation, capillary electrophoresis was carried out for fragment analysis using Applied Biosystem 3730 Genetic analyzer.

3.9.1 Sample Preparation

Sample preparation for ABI 3730 Genetic Analyzer was carried out initially by diluting each of the 5 dye matrix standard (Flourescein, JOE, TMR-ET, CXR-ET and CC5) in the ratio of 1:10 with Nuclease free water after thawing them. After this Fragment mix was prepared.

15 µL of each of the 5 dye matrix standards was taken from the initial dilution and was mixed with 1425 µL of Hi-Di™ formamide. 96 wells were used on 96 capillaries for matrix detection on ABI PRIMSM® 3730 Genetic Analyzer. 25 µL of fragment mix was loaded to each well containing 1 µL of sample in each well and 1 µL of PowerPlex® Y23 allelic

ladder mix to one of the wells, and plate was centrifuged for a few seconds to remove bubbles.

PCR product was denatured at 95°C for 3 minutes, and then was immediately chilled on crushed ice for 3 minutes, just before loading to the instrument.

3.9.2 Reading Alleles calls

The peak sizes and positions of the amplified alleles were compared with the PowerPlex® Y23 allelic ladder using Gene Mapper® Software 5 after capillary electrophoresis. The range of different alleles for 23 Y-STR loci is given in the fig.22.

3.10 Comparative populations

To investigate the genetic diversity of five major Pathan tribes of Charsada and Mardan districts and to compare with the other Pathan populations in the neighboring area of Mohmand agency and Swat valley and populations from neighboring countries, their Y chromosome genotype data was exploited.

For Y STR analysis, the comparative population data used was obtained from the published literature of Afghanistan[Pathan] (Lacau *et al.*, 2012), Iraq[Iraqi] (Purps *et al.*, 2014), Iran [Arabs, Bakhtiari, Galaki, Mazandarani, SouthernTalysh] (Rower *et al.* 2009), Turkey [Dogukoy, Eskikoy, Gocmenkoy, Merkez] (Alakoc *et al.*, 2010), Greece [Greeks] (Purps *et al.*, 2014), Israel [Muslim Arabs] (Farnandes *et al.*, 2011), Russian Federation [Archangelskaja, Brianskaja, Ivanoskaja, Lipezkaka, Ryasankaja, Smolenjkaja, Tamboskaja, Tverskaja, Wologodskaja] (Rower *et al.*, 2008), India [Tamil] (Balamurugan *et al.*, 2010), Xinjiang China [Uighur, Kazakh] (Shan *et al.*, 2014), Tibet China [Tibetan]

(Ye *et al.*, 2015), Pakistani Pathans (Lee *et al.*, 2014) and Yousafzai Pathans (KP) Pakistan (Ilyas *et al.*, 2012).

3.11 Mitochondrial DNA amplification

Out of 374 samples 165 samples from all five populations were amplified with 15971F (TTAACTCCACCATTAGCACC) and 484R (TGAGATTAGTAGTATGGGAG) primers and the sequence length of 1082bps was amplified using 25 μ L total reaction volume. The amplification was carried out using" Eppendorf Mastercycler ep Gradient S thermal cycler" of Applied Biosystem (Fig.16).

Recipe is given in the table: 3.

Table: 3. Reagents used in PCR reaction mixture

S. NO	Reagents	Volume of Reagents
1	10X buffer w/o MgCl ₂	2.5 μ L
2	10mM dNTPs	1 μ L
3	50mM MgCl ₂	1 μ L
4	(5 μ M) F Primer	1 μ L
5	(5 μ M) R Primer	1 μ L
6	5% Glycerol	1 μ L
7	Platinum Taq polymerase (5U/ μ L)	0.2 μ L
8	ddH ₂ O	14.3 μ L
9	Template DNA	3 μ L
	Final Volume	25 μ L

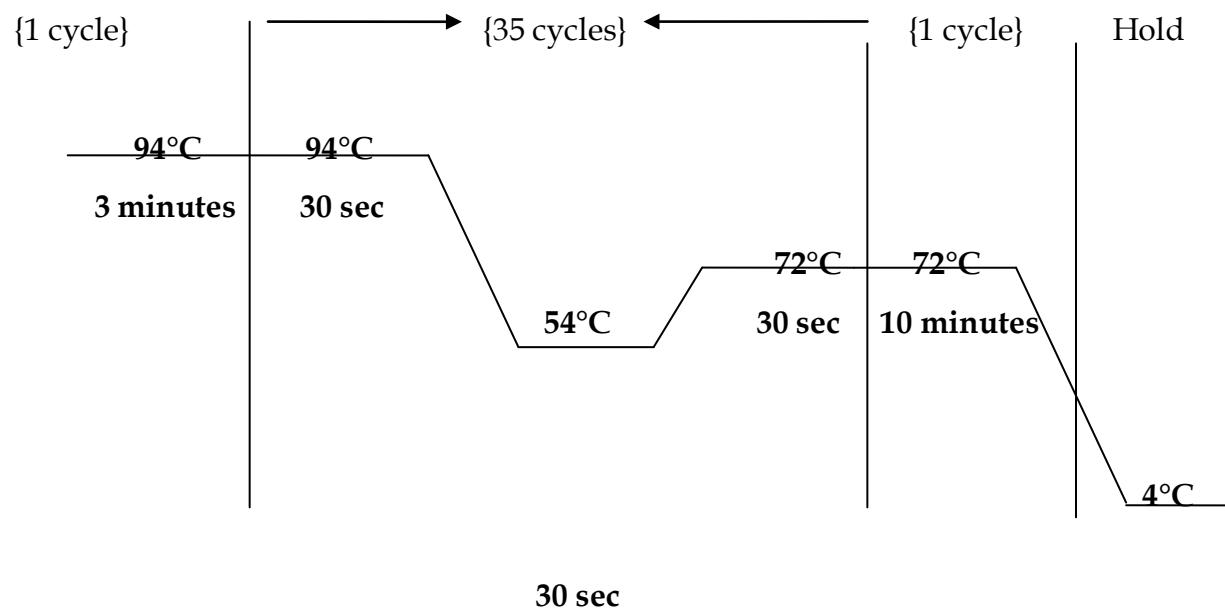


Fig.16. Thermo cycling conditions for mtDNA amplification

3.12 PCR product clean-up

5 μ L of the PCR product was mixed with 2 μ L of Exo-SAP-it in minitubes (SAP: Shrimp Alkaline Phosphatase and EXO: Exonuclease I) (Malhi *et al.*, 2010). After spinning, tubes were incubated at 37°C for 30 minutes followed by heat inactivation at 80°C for 15 minutes and were held at 25°C (Fig.17).

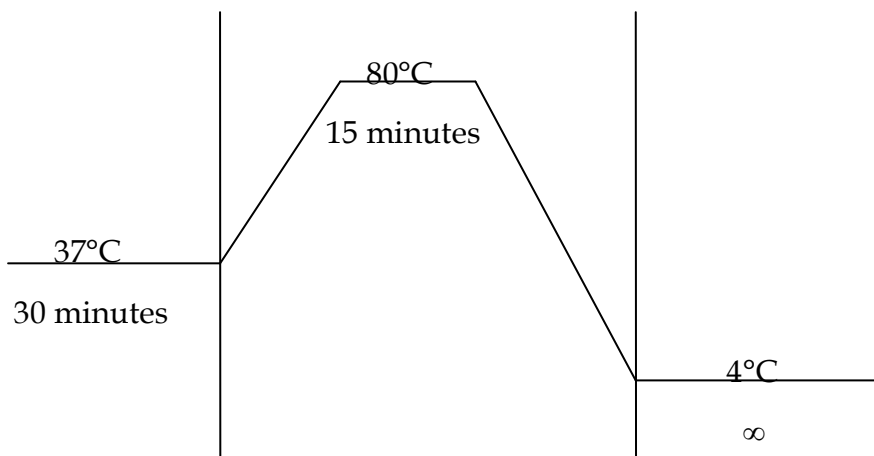


Fig.17. Thermo cycling conditions for EXO- SAP.

The purpose of this procedure was to chew up excess primers and removal of excess dNTPs from the PCR product, to make the DNA sequences readable.

3.13. Agarose Gel Electrophoresis

In order to visualize the PCR product to make sure that it worked and there were no multiple bands, the PCR product was electrophorased in 2% agarose gel. In order to prepare 2% agarose gel, 3g of agarose was added to 150mL of 1X TBE buffer. Mixture was boiled in microwave oven for 1.30 minutes at the interval of 30sec in order to avoid spillage. Solution was then cooled up to 45°C. After sufficient cooling Ethidium bromide was added in to it and solution was poured in to the gel cassette with the combs fitted into it to create wells. After solidification, combs were removed and 5 μ L of each PCR product was mixed with 2 μ L of loading dye. All the samples were loaded in the separate well. 2-Log DNA ladder (0.1 – 10.0kb) was loaded in the first well for comparison. Representation of Gel results is given in Fig.18.

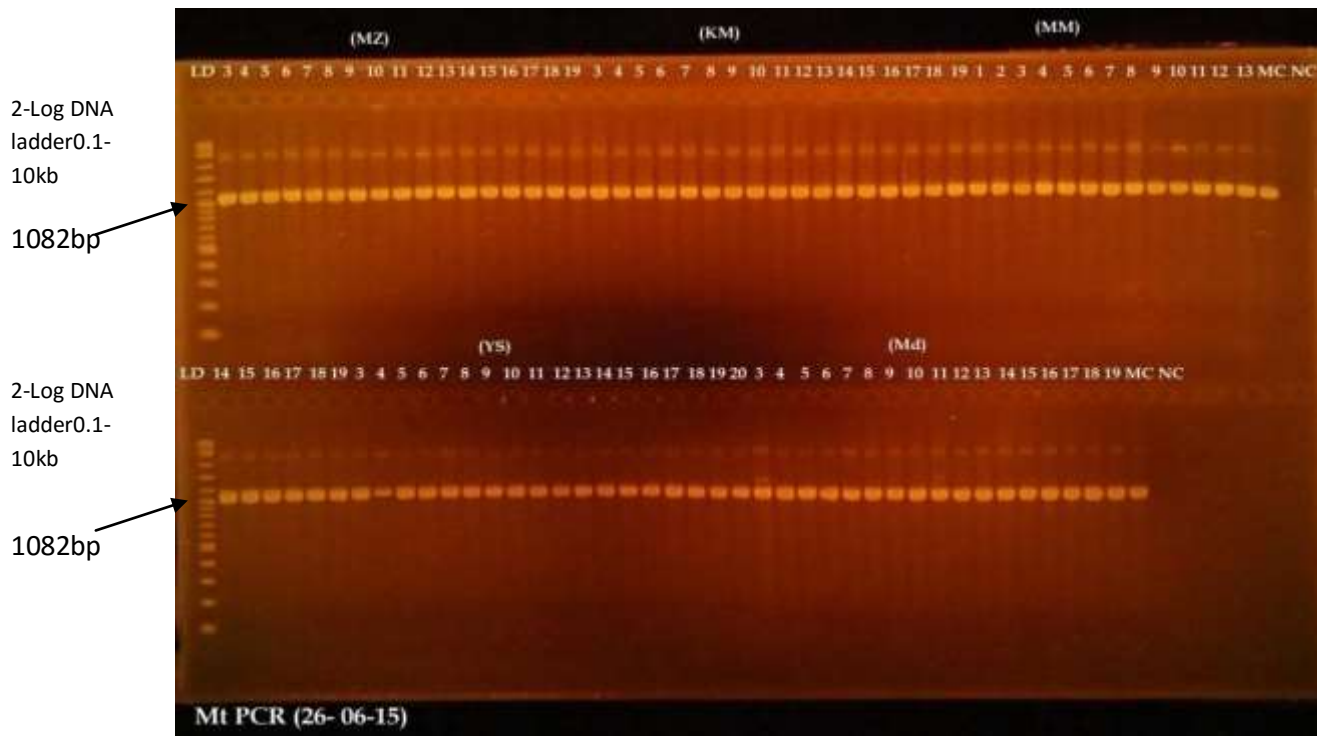


Fig.18. Agarose Gel Electrophoresis photograph of amplified mtDNA. MC: Male control, NC: Negative control

3.14 mtDNA Sequencing

For mitochondrial DNA Hyper variable region I (HVI) sequencing, the 2 μ L of good quality clean PCR product was mixed with 1 μ L of 5 μ M of 15971 F primer. Sequencing was carried out using Big Dye™ terminator cycle sequencing kit of Applied Biosystem on Genetic analyzer 3730.

3.14.1 Sequence reading

After retrieval, Sequences were read using the program Sequencher 4.7 and all sequence data was aligned with revised Cambridge reference sequence (rCRS) (rCRS; Andrews *et al.*, 1999) using bioedit program. Variable positions were determined from np 16024 – 16365 and haplogrup was assigned to each sample using phylotree build 16 (Van Oven M.2009) and mitomaster (Lott *et al.*, 2013).

To investigate the genetic diversity of five major Pathan tribes of Charsada and Mardan districts and to compare with the other Pathan populations in the neighboring area of Mohmand agency and Swat valley and populations from neighboring countries, their mtDNA HVI sequence data was exploited.

3.15 Comparative populations

For mtDNA analysis the comparative data from populations of India [Afridi, Tamil] (Metspalu *et al.*, 2004), Israel [Druze, Bedauin] (Hunley *et al.*, 2009), Greece [Greeks] (Irwin *et al.*, 2008), Russian Federation [Yakut] (Hunley *et al.*, 2009), Iran [Armenian, Azeris, Persian, Qashqis] (Derenko *et al.*, 2013), Iraq [Iraqi] (Abu *et al.*, 2008), China [Uighur] (Hunley *et al.*, 2009), Afghanistan, Turkey [Turkish] (Comas *et al.*, 1996), indigenous populations from Pakistan [Pathan, Kalash, Brusho, Baluch, Brahui] (Hunley *et al.*, 2009) was used.

3.16 Statistical analysis

Genetic diversity estimates were calculated for all the five Pathan populations of the two areas separately as well as collectively using a formula $[n(1-\sum p_i^2)/n-1]$, where n is the number of samples and p_i is the frequency of the i^{th} haplotype in the population (Nei, 1987). Genetic distances (Matrix of Slatkin linearized F_{ST}) were obtained by using ARLEQUIN Ver 3.5.2.2 software for our data and for other global populations. Of all pairwise F_{ST} values a distance matrix was constructed that was later used to construct a PCoA plot using GENALEX program (Peakall *et al.*, 2006). Median Joining (MJ) networks were constructed using NETWORK 4.6.1.3 (<http://www.fluxus->

engineering.com) by giving each locus a weight according to its estimated mutation rate (Keyser *et al.*, 2003). Mitomaster and phylotree were used to predict the mitochondrial haplogroups (<http://www.mitomap.org>) (Lott *et al.*, 2013) and Y-haplogroups were identified by using server haplogroup predictor (www.hprg.com). For STR analysis, the haplotypes were reduced to 17 STR loci (DYS19, DYS385a/b, DYS389I/II, DYS390, DYS391, DYS392, DYS393, DYS437, DYS438, DYS439, DYS448, DYS456, DYS458, DYS635, Y-GATA-H4) for comparison with published available global populations data sets.

3.17 Mantel test

At the end Mantel test were performed using GENALEX program (Peakall *et al.*, 2006) to measure the degree of correlation between mtDNA based Genetic distance among population and geographical distance, between Y-STR based Genetic distance and Geographic distance and between mtDNA based and Y-STR based genetic distances.

Chapter4 RESULTS

This chapter includes Y-STR and mtDNA analysis results profiles consisting of Genetic Diversity parameters for Y-STR and mtDNA HVI data sets, Haplogroup analysis, Network analysis, Analysis of Molecular Variance, Principal Coordinate analysis and Mantel Test.

4.1 The Y-STRs analysis

Analyses of the results obtained from Y-STR DNA samples obtained from 374 volunteers of the districts Mardan and Charsada segregated into 344 haplotypes wherein 374 samples were belonging only to 11 haplogroups (Annexure I), with an overall Discrimination Capacity (DC) of 92%.

4.2 Genetic Diversity

The genetic diversity (GD) obtained for all the five Pathan populations was 0.99 which is similar to the already studied Pathans from Pakistan (Lee *et al.*, 2014). Genetic diversity estimates and other population genetics parameters for different tribes are given in the table 4. Comparison of genetic diversity estimates among various indigenous Pakistani populations is given in table 5.

Table: 4. Population genetics parameters for 23 Y-STRs in five Pathan populations of Charsada and Mardan district.

Parameters	CHARSADA			MARDAN		All 5 populations
	(MZ)	(MM)	(KM)	(YS)	(MD)	
Number of samples(n)	75	75	75	75	74	374
No. of Haplotypes	72	71	61	73	67	344
Unique Haplotypes	70	67	51	71	62	320
Shared Haplotype	2	4	10	2	5	24
Random Match Probability (RMP)	0.0136	0.014	0.019	0.0133	0.0158	0.00305
Power of Discrimination (PD)	0.986	0.985	0.980	0.986	0.984	0.996
Genetic Diversity(GD)	0.999	0.998	0.994	0.999	0.997	0.998

Note: The abbreviations MZ, KM, YS, MM and MD denote the tribes Muhammadzai, Kakakhel Mian, Yousafzai and Mohmand respectively.

Table: 5. Comparison of Y-STR Genetic diversity among various ethnic groups of Pakistan

Parameters	Path (present study)	Mkr (Qamar <i>et al.</i> 2002)	Pth (Lee <i>et al.</i> 2014)	Bal (Qamar <i>et al.</i> 2002)	Brh (Qamar <i>et al.</i> 2002)	Haz (Qamar <i>et al.</i> 2002)	Bsh (Qamar <i>et al.</i> 2002)	Ksh (Qamar <i>et al.</i> 2002)	Par (Qamar <i>et al.</i> 2002)	Snd (Qamar <i>et al.</i> 2002)
No. of Sample	374	33	230	59	110	23	94	44	90	122
No. of haplotypes	344	30	211	48	85	11	63	26	60	101
Genetic Diversity	0.998	0.992	0.997	0.988	0.973	0.893	0.987	0.952	0.974	0.995

Note: The abbreviations Path, Mkr, Pth, Bal, Brh, Haz, Bsh, Ksh, Par, Snd are Pathan from Charsada and Mardan, Makrani, Pathan, Baluch, Brahui, Hazara, Barusho, Kalash, Parsi and Sindhi, respectively

4.3 Y-Haplogroups found

Analysis of haplogroups segregated from the DNA samples revealed that around 49.2% of the individuals belonged to one of three major Haplogroups i.e R1a, 17.9% from L haplogroup and 9.6% from G2a haplogroup. A relative incident of the haplogroups is presented in the table. 6.

Table: 6. Frequency of Y-haplogroups found in five populations under study and Pathan population from Southern Afghanistan.

	MZ n=75 (This study)	KM n=75 (This study)	MM n=75 (This study)	YS n=75 (This study)	MD n=74 (This study)	Total N=374 (This study)	Southern Afghanian (N=145) (Sangupta <i>et</i> <i>al.</i> 2007)
Haplogroup	Frequency of Haplogroups (%)						
R1a	50.7	41.3	54.6	54.6	44.6	49.2	66.8
L	17.3	18.7	10.7	17.3	25.7	17.9	5.6
G2a	10.7	4	14.7	10.7	8.1	9.6	8.2
Q	8	13.3	4	5.3	5.4	7.21	1.3
H	8	10.7	1.3	-	1.4	4.27	2.8
J1	-	6.7	-	1.3	8.1	3.20	2.0
R1b	-	1.3	6.7	4	4	3.20	0.6
J2b	1.3	2.7	4	-	1.3	1.87	-
E1b1a	-	1.3	-	5.3	-	1.33	0.6
E1b1b	2.7	-	1.3	-	1.4	1.06	0.6
I2a	1.3	-	2.7	1.3	-	1.06	-

(Note: The abbreviations MZ, KM, MM, YS, MD denotes the tribes Muhammadzai, Kakakhel Mian, Mohmand from Charsada, Yousafzai and Mohmand from Mardan, respectively).

Haplogroups Q, H, J2b, J1, J2a, R1b, E1b1a and E1b1b were also present comprising 7.2%, 4.2%, 1.87%, 3.2%, 1.33% and 1.06% respectively of the entire sample set investigated. Of the haplotypes generated, 7% of all were shared among populations and almost 93% of the haplotypes were unique to the individuals.

Muhammadzai (MZ), Kakakhel Mian (KM) and Mohmand (MM) samples came from Charsada area and Mohmand (MD) and Yousafzai (YS) from Mardan area. Total of 7% of the haplotypes were shared among all the five populations of the two areas.

4.4 Network analysis of Y Haplogroups

The network analysis of the haplogroups showed that the Haplotype „a“ is the central haplotype with overall frequency of 26.1% present in 3.8% of MZ haplogroup R1a haplotypes, 5.4% of MM Hg R1a haplotypes, 7.1% in KM R1a haplotypes, 6% in MD R1a haplotypes and 3.8% in YS R1a haplotypes. Similarly haplotype b is the second haplotype shared among all the five populations with the frequency of 6.5% in MM, 1.6% in MZ and MD and 0.5% in KM R1a haplotypes. Little haplotype sharing was observed in haplogroup H and G2a (Fig.19 a).

No common or shared haplotype was observed among the samples of either of the population in Haplogroup H while single haplotype was shared between the individuals of Mohmand from Charsada (MM), Kakakhel Mian (KM) and Muhammadzai (MZ) in Haplogroup G2a (Fig.19 b, c).

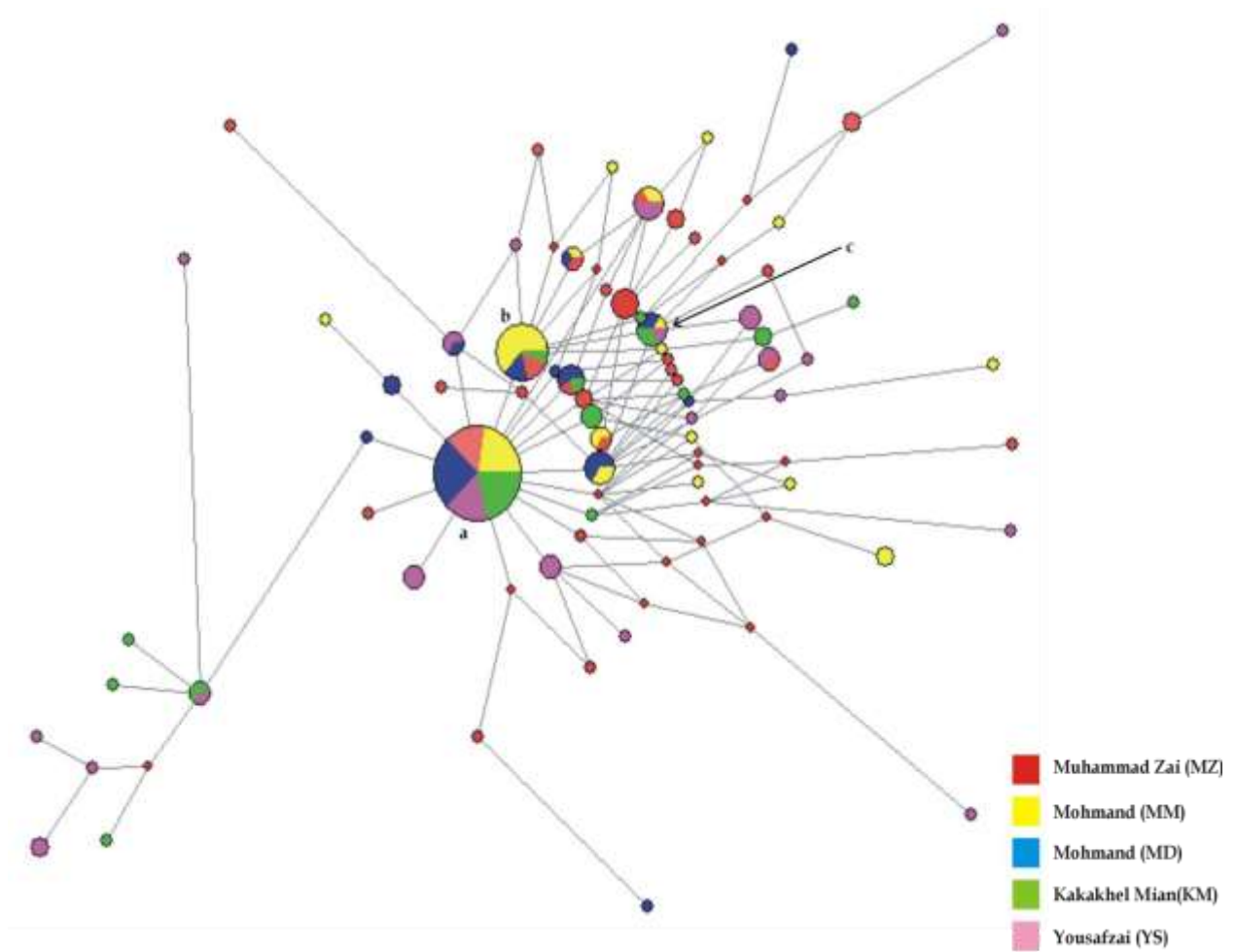


Fig.19.a. Median-Joining network of Y-haplogroup R1a in five populations of Charsada and Mardan. Areas of circles are proportional to the haplotype frequencies. Haplotype "a" and "b" are most frequent haplotypes shared among all the five populations.

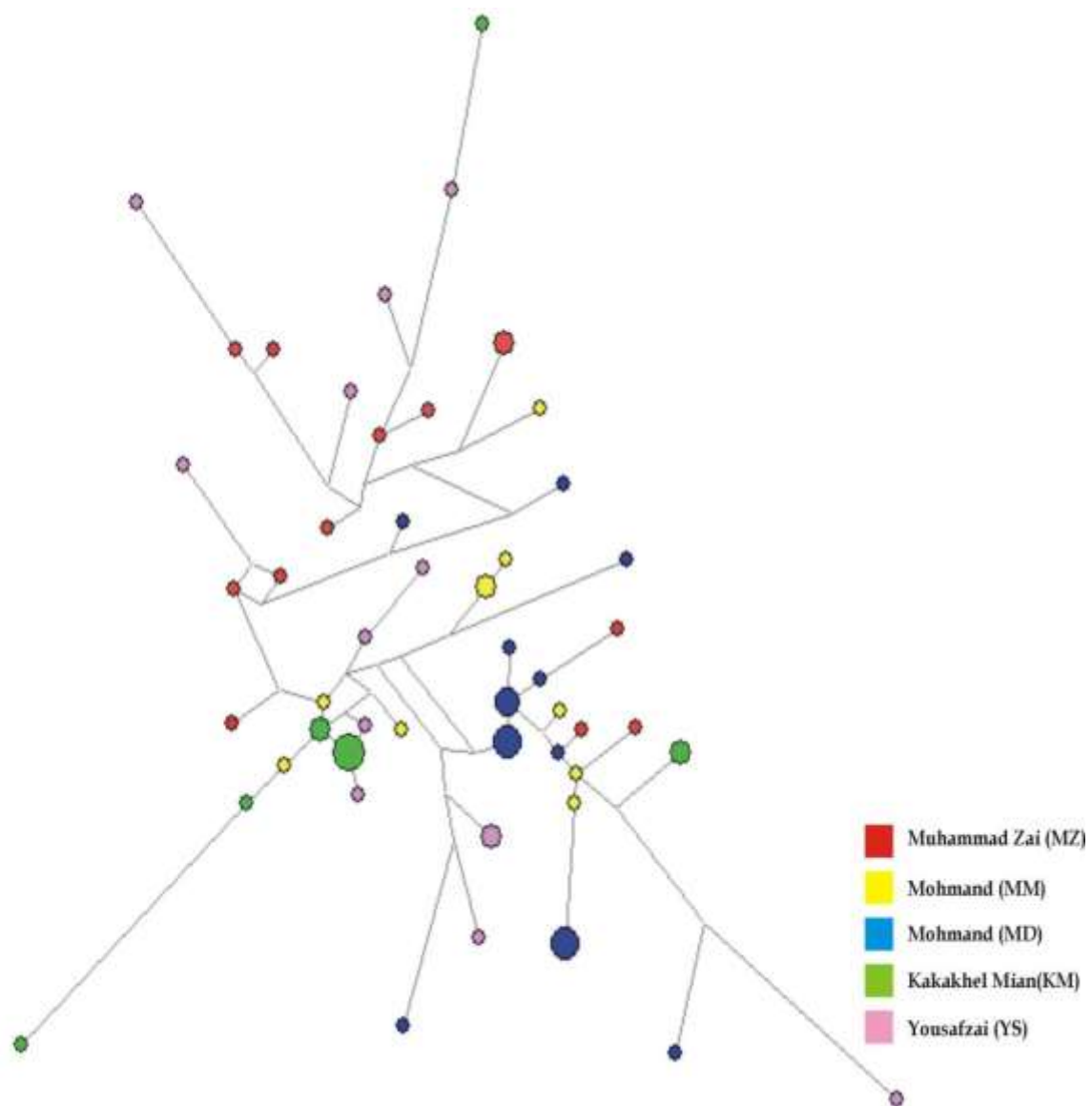


Fig.19.b. Median-Joining network of Y-haplogroup L in five populations of Charsada and Mardan. Areas of circles are proportional to the haplotype frequencies.

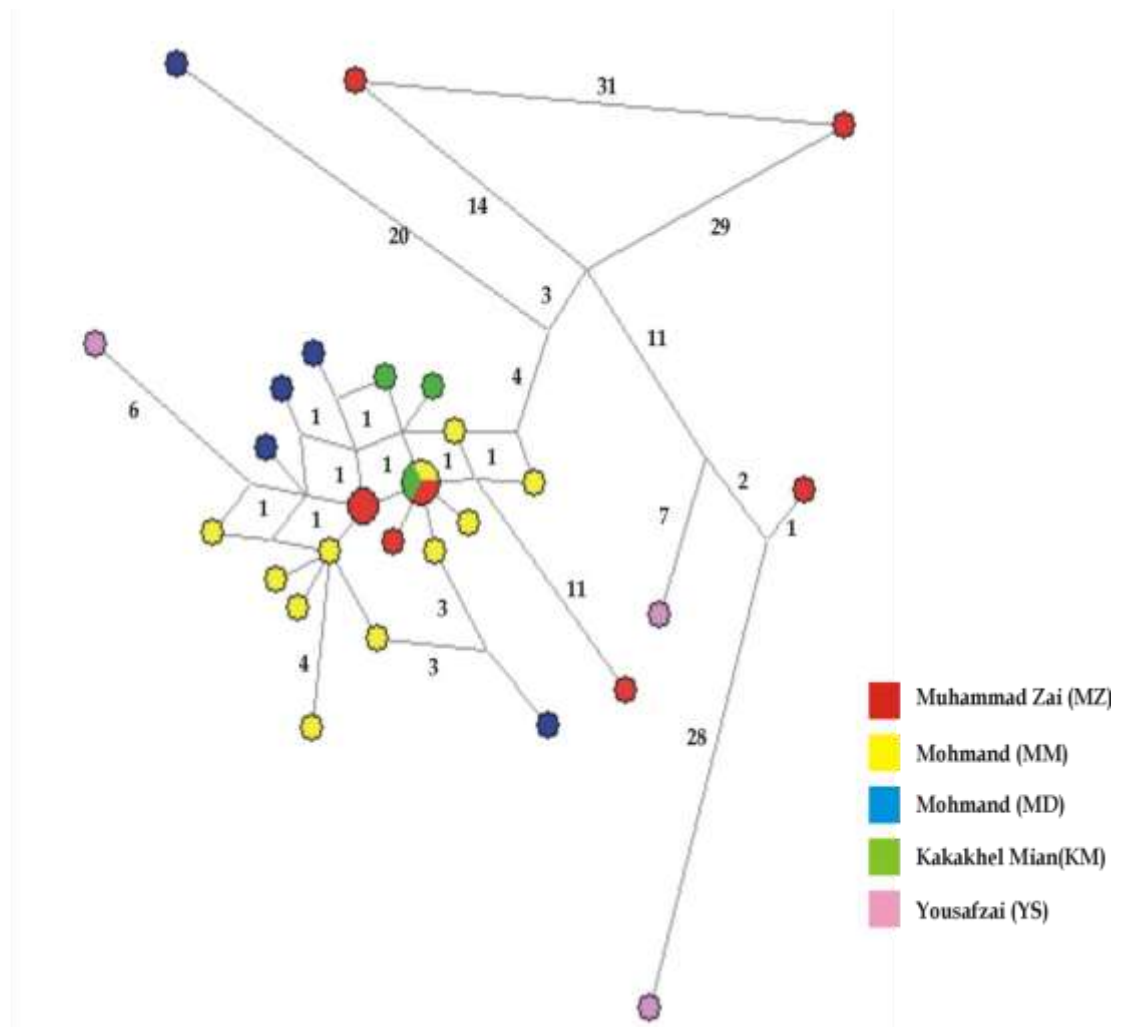
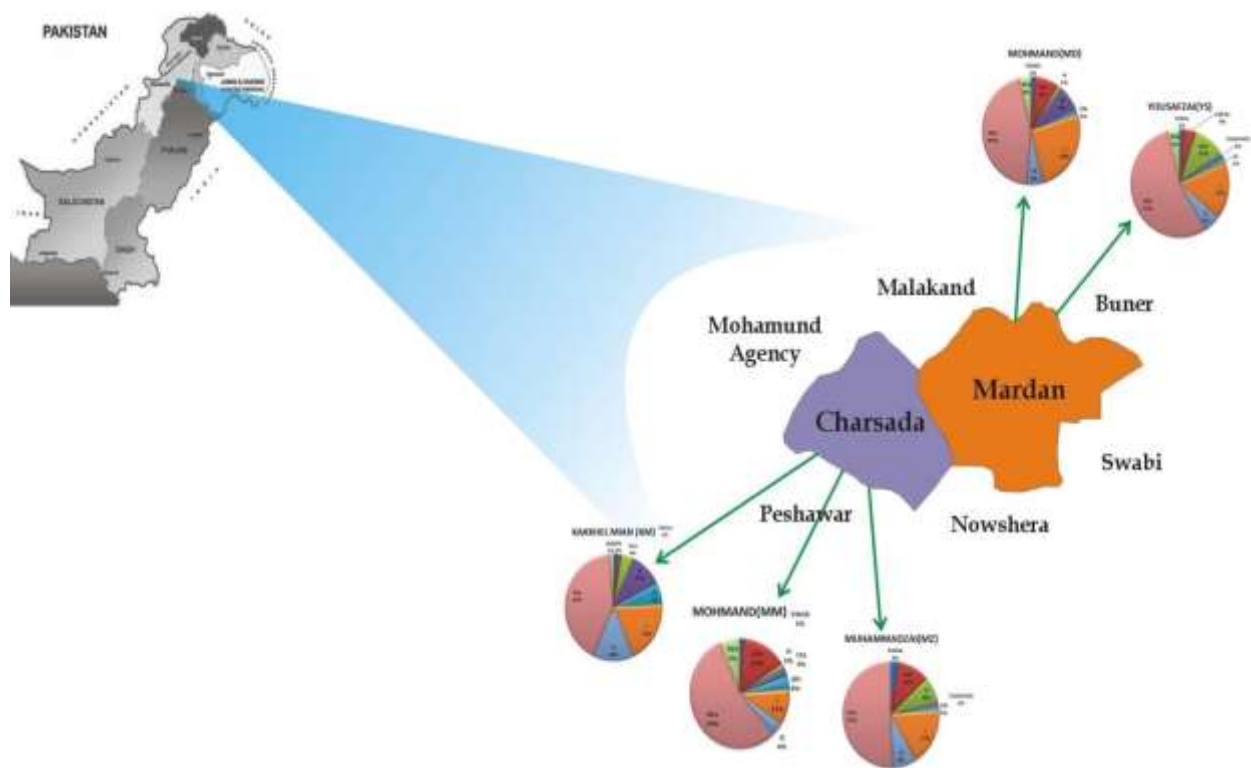


Fig.19.c. Median-Joining network of Y-haplogroup G2a in five populations of Charsada and Mardan. Areas of circles are proportional to the haplotype frequencies.

4.5 Haplogroup distribution among the population of two districts

Charsada populations had a few additional haplogroups not found in Mardan populations. More diverse haplogroups points out the different history of this location (Fig.20).



4.6 Genetic distance among five populations

No significant genetic distance (F_{ST}) was found between the pair of populations from both the districts. The highest genetic distance of 0.04 between Kakakhel Mian (KM) and Mohmand (MD) was observed followed by 0.03 between Kakakhel Mian(KM) and Muhammadzai (MZ), 0.02 between Mohmand (MD) from Mardan and Muhammadzai (MZ) while no distance was observed between Muhammadzai, Mohmand (MM) from Charsada and Yousafzai (YS) Fig.21.

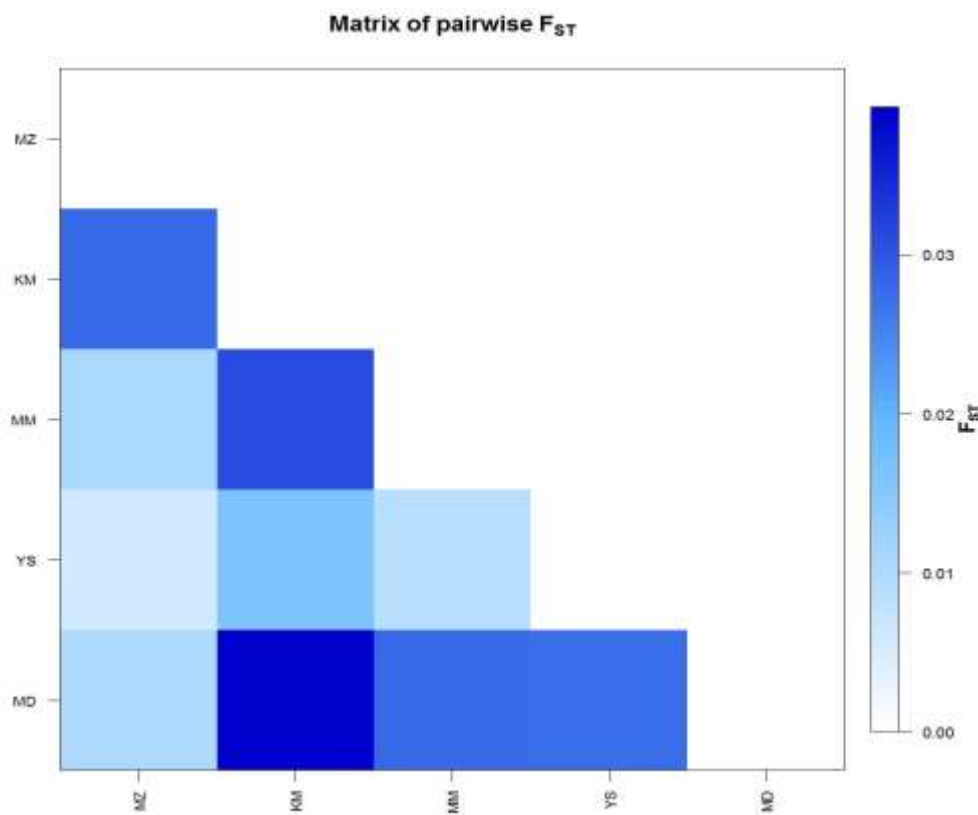


Fig.21. Y-STR based Pair wise Genetic distances (F_{ST}) between five Pathan population pairs of Charsada and Mardan district. (The abbreviations MZ, KM, MM, YS, MD denote the tribes Muhammadzai, Kakakhel Mian, Mohmand from Charsada, Yousafzai and Mohmand from Mardan, respectively).

4.7 Analysis of Molecular Variance among population of two districts

AMOVA analysis for Y-STR haplotypes was used to examine the partitioning of genetic variation for these five populations of two districts divided into two groups. In this case 98.20% of variation found “within the populations” while -0.6% of variation was found “among the groups” and 2.46% “among population within the group” (table: 7).

Negative values indicate zero value or no variation between the groups.

Table: 7. AMOVA design and results (average over 23 Y-STR loci) in five populations of two districts (Groups).

Source of Variation	Sum of Squares	Variance Components	Percentage Variation
Among groups	13.379	-0.04132	-0.56909
Among populations within groups	62.417	0.18488	2.54624
Within populations	2601.665	7.11737	98.02285
Total	2677.460	7.26093	

4.8 Allele Frequencies in all the five Populations

Number of alleles at 23 different loci in all the five populations is graphically represented in fig.22.

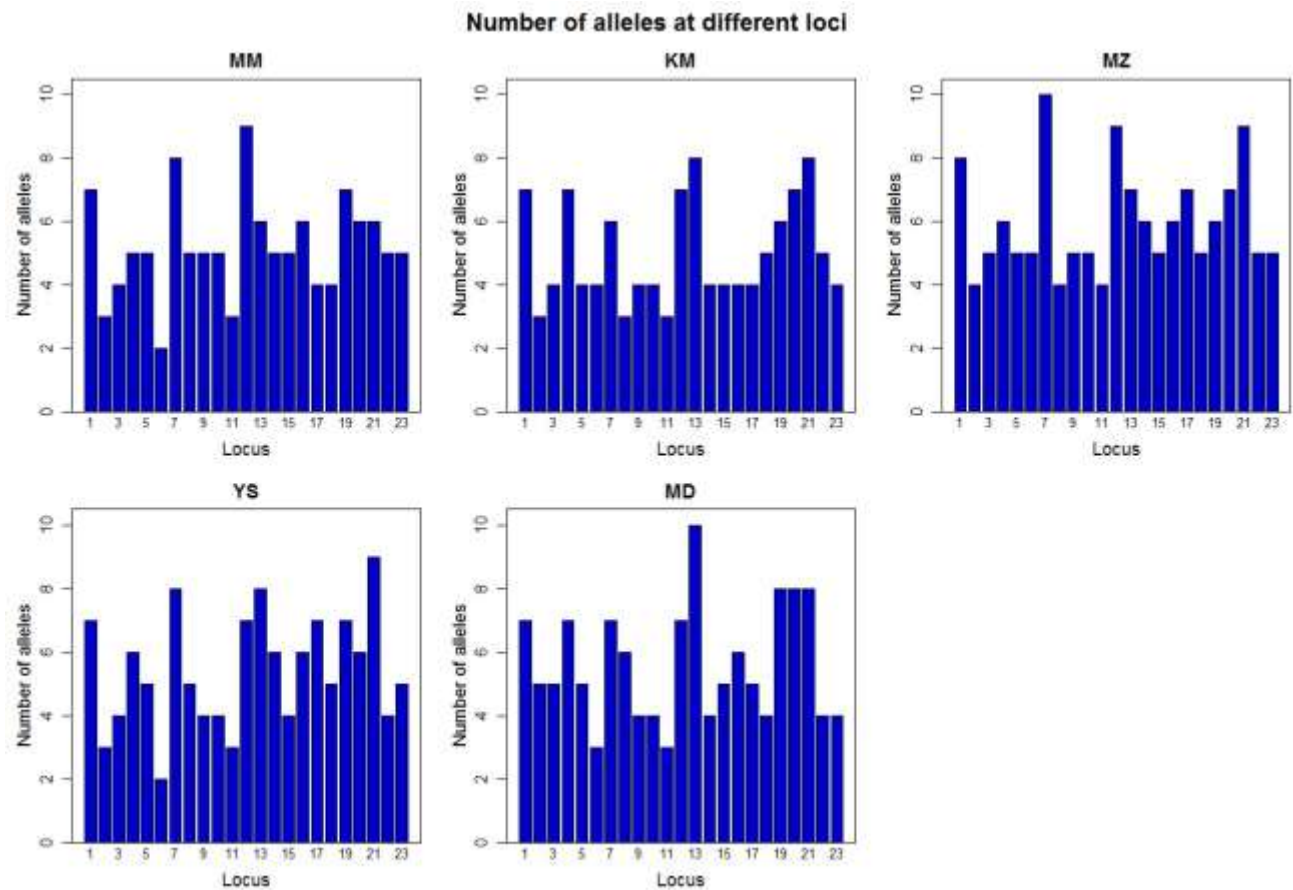


Fig.22. Graphical representation of allele frequencies of 23 loci in five populations of Charsada and Mardan district.

(The abbreviations MZ, KM, MM, YS and MD denote the tribes Muhammadzai, Kakakhel Mian, Mohmand from Charsada, Yousafzai and Mohmand from Mardan, respectively).

4.8

.1 Yousafzai: YS (Mardan)

Allele frequencies of 23 Y-STR loci in Yousafzai population of Mardan district are given in table.8. Where the most common allele found was DYS456 (15) with the frequency of 0.720 while the least occurring alleles found in this population were DYS576 (10, 15), DYS19 (13), DYS389II (33), DYS481 (21, 27), DYS549 (15), DYS635 (20), DYS390 (20), DYS643 (15), DYS393 (15), DYS385b (20, 21) and GATAH4 (10, 14) with the frequency of 0.0133.

The most polymorphic locus in this population was DYS385b with total of 9 alleles found and the least polymorphic was DYS19 with two alleles found at this locus.

Table: 8. Allele Frequencies of 23 Y-STRs in Yousafzai (Ys) population of Mardan district.

Alleles	DYS 576	DYS 389I	DYS 448	DYS 389II	DYS 19	DYS 391	DYS 481	DYS 549	DYS 533	DYS 438	DYS 437	DYS 570	DYS 635	DYS 390	DYS 439	DYS 392	DYS 643	DYS 393	DYS 458	DYS 385a	DYS 385b	DYS 456	YGATA H4
8																	0.0400						
9										0.04							0.0267			0.04			
10	0.0133					0.56			0.0933	0.28					0.48	0.0667	0.6133						0.0133
11						0.44		0.0933	0.2000	0.64					0.28	0.6933	0.1867	0.0667		0.5867			0.1067
12		0.12						0.0133	0.6667	0.04					0.16	0.0400	0.0667	0.0533		0.0400			0.6133
13		0.64			0.0133			0.3600	0.0400						0.08	0.0267	0.0400	0.7067		0.2133	0.0400		0.2267
14		0.24			0.2400			0.0267			0.6800	0.0533				0.1200		0.1600	0.0400	0.0933	0.5067	0.0533	0.0133
15	0.0133				0.3333			0.0133			0.1467	0.0933				0.0267	0.0133	0.0133	0.4133	0.0267	0.0933	0.7200	
16	0.1467				0.2533						0.1733	0.0533							0.1867		0.2000	0.2000	
17	0.2133				0.0533							0.1600							0.2133		0.0800	0.0267	
18	0.3733		0.0400									0.2400	0.04						0.0400		0.0267		
19	0.1600		0.1867				0.0667					0.2933							0.0800		0.0267		

20	0.0800		0.5867									0.1067	0.0133	0.0133					0.0267		0.0133		
21			0.1467				0.0133						0.0667	0.0267							0.0133		
22							0.0800						0.0267	0.0933									
23							0.4400						0.5200	0.2400									
24							0.1867						0.2133	0.4400									
25							0.1333						0.0800	0.1867									
26							0.0667						0.0400										
27							0.0133																
28				0.1067																			
29				0.1733																			
30				0.4533																			
31				0.1467																			
32				0.1067																			
33																							

				0.0133																			
GD	0.7715	0.5254	0.5716	0.7297	0.7151	0.4995	0.7481	0.6123	0.5121	0.5157	0.4926	0.8159	0.6782	0.7139	0.6681	0.4760	0.5768	0.4739	0.7485	0.6058	0.6941	0.4440	0.5438

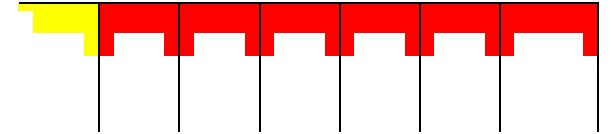
4.8

.2 Mohmand: MD (Mardan)

In Mohmand(MD) population of Mardan District, the most frequently occurring allele was DYS533 (12) with the highest frequency of 0.6301 while the least occurring alleles were DYS576 (21), DYS389II (27, 32), DYS19 (13), DYS549 (15), DYS635 (19, 20), DYS392 (15), DYS643 (14), DYS458 (20, 22), DYS385b (20, 22), GATAH4 (10) with the frequency of 0.0137. The most polymorphic locus among all 23 loci in this population was found DYS635 with total 9 alleles(Table: 9).

Table: 9. Allele Frequencies of 23 Y-STRs in Mohmand (MD) population of Mardan district.

Alleles	DYS 576	DYS 389I	DYS 448	DYS 389II	DYS 19	DYS 391	DYS 461	DYS 549	DYS 533	DYS 438	DYS 437	DYS 570	DYS 635	DYS 390	DYS 439	DYS 392	DYS 643	DYS 393	DYS 498	DYS 385a	DYS 385b	DYS 498	YGATA 134
7																							
8															0.0822								
9										0.0548							0.0685			0.1507			
10						0.5616		0.0959	0.0822	0.3973					0.4384	0.0685	0.5205			0.0274			0.0137
11		0.0822				0.3973		0.0959	0.2603	0.5068					0.1918	0.6438	0.1096	0.0137		0.4247			0.3425
12		0.0548				0.0411		0.4795	0.6301	0.0411					0.2329	0.0411	0.2877	0.3562		0.1644			0.5616
13		0.7123			0.0137			0.2192	0.0274						0.0548	0.0274		0.5616		0.1233	0.0411	0.0685	0.0822
14		0.1370			0.2055			0.0959			0.5616					0.2055	0.0137	0.0685	0.0274	0.0274	0.4384	0.0548	
15	0.1233	0.0137			0.3699			0.0137			0.2055	0.0411				0.0137			0.3425	0.0274	0.0548	0.5753	
16	0.1918				0.3562						0.2329	0.2466							0.1644		0.1781	0.3014	
17	0.0548		0.0822		0.0411							0.1096							0.2055		0.2055		
18	0.1233		0.0411									0.1096							0.0959		0.0548		
19	0.4110		0.3836				0.0685					0.3699	0.0137						0.1370				
20	0.0822		0.3836									0.0548	0.0137						0.0137		0.0137		
21	0.0137		0.1096									0.0685	0.1918										
22							0.0548						0.1507	0.2466					0.0137		0.0137		
23							0.1644						0.4247	0.2329									
24							0.2466						0.1096	0.4521									
25							0.3288						0.0274	0.0685									
26				0.0822			0.0548						0.0274										
27				0.0137			0.0822						0.0411										
28				0.0274																			
29				0.2466																			
30				0.5205																			
31				0.0959																			
32				0.0137																			
GD	0.7645	0.4703	0.6948	0.6602	0.6933	0.5323	0.7976	0.7040	0.5350	0.5887	0.5963	0.7797	0.7664	0.6853	0.7169	0.5434	0.6381	0.5605	0.7953	0.7599	0.7359	0.5784	0.5681



4.8

.3 Muhammadzai: MZ (Charsada)

Allele frequencies of all 23 Y-STRs in Muhammadzai (MZ) population are given in table: 10.

Most common alleles identified in this population were DYS392 (11), DYS456 (15) with the frequency of 0.7067. The lowest frequency of alleles found among all the 23 alleles was 0.0133 for DYS389I (10), DYS448 (18, 22), DYS391 (12), DYS481 (18, 27, 28), DYS437 (17), DYS570 (11, 14, 22), DYS635 (19), DYS390 (26), DYS439 (14), DYS392 (15), DYS643 (8, 9, 13), DYS393 (11), DYS385b (13, 21), DYS456 (13), GATAH4 (10, 14). The most polymorphic among all the loci was DYS481 with 10 alleles ranging from allele number 18 to allele number 28.

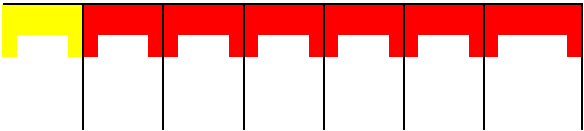
Table:

10. Allele Frequencies of 23 Y STRs in Muhammadzai (MZ) population of Charsada district.

Alleles	DYS 376	DYS 389I	DYS 448	DYS 389II	DYS 19	DYS 391	DYS 461	DYS 549	DYS 531	DYS 438	DYS 437	DYS 570	DYS 635	DYS 390	DYS 439	DYS 392	DYS 643	DYS 381	DYS 436	DYS 385a	DYS 385b	DYS 434	YGATA 114
7						0.04																	
8										0.0267							0.0133						
9						0.0267			0.0267	0.0800							0.3333			0.0400			
10		0.0133				0.4933			0.0800	0.2533					0.4800	0.1200	0.6267						0.0133
11						0.4267		0.0267	0.2133	0.6400		0.0133			0.3333	0.7067	0.1067	0.0133		0.4933			0.4000
12		0.0800				0.0133		0.6400	0.6533						0.1333	0.0667	0.1067	0.2000		0.0800			0.4133
13		0.7600			0.0533			0.2800	0.0267						0.0400	0.0267	0.0133	0.6800		0.2000	0.0133	0.0133	0.1600
14	0.0533	0.1467			0.0533			0.0533			0.6533	0.0133			0.0133	0.0400		0.1067	0.0267	0.1067	0.4667	0.0400	0.0133
15	0.0400				0.4133						0.1067	0.0533				0.0133			0.4000	0.0400	0.0800	0.7067	
16	0.1733				0.4133						0.2267	0.0667							0.1867	0.0400	0.1067	0.2133	
17	0.0667				0.0533						0.0133	0.2000							0.2000		0.0933	0.0267	
18	0.1867		0.0133				0.0133					0.1733							0.1067		0.0667		
19	0.3067		0.2933				0.0667					0.4133	0.0133						0.0800		0.0933		
20	0.1467		0.5733									0.0533	0.0800								0.0667		
21	0.0267		0.1067				0.0533					0.0133	0.1467	0.0533							0.0133		
22			0.0133				0.0800						0.0400	0.1467									
23							0.3067						0.4533	0.2533									
24							0.1200						0.2000	0.3733									
25							0.2933						0.0667	0.1600									
26							0.0400							0.0133									
27							0.0133																
28				0.0933			0.0133																
29				0.2267																			
30				0.5067																			
31				0.1200																			
32				0.0133																			
33																							
GD	0.8209	0.3996	0.5813	0.6760	0.6490	0.5798	0.8004	0.5153	0.5269	0.5279	0.5171	0.7586	0.7301	0.7564	0.6476	0.4566	0.5798	0.5106	0.7568	0.7034	0.7478	0.4587	0.6519

Table:

-



4.8

.4 Kakakhel Mian

In table: 11, the allele frequencies of all the loci in Kakakhel Mian (KM) population are given. The highest allele frequency was found in this population was 0.7467 for DYS533 while the lowest allele frequency recorded in this population was 0.0133 for DYS576 (14, 15), DYS389II (20, 24, 26), DYS19 (17), DYS391 (9, 12), DYS481 (22), DYS533 (9), DYS438 (12), DYS635 (18, 20, 25, 26), DYS385a (10, 15), DYS385b (13), DYS456 (17) and GATAH4 (10). DYS635 and DYS 385b were considered to be the most polymorphic loci among all 23 loci typed in this population with 8 alleles.

Table:

11. Allele Frequencies of 23 Y STRs in Kakakhel Mian (KM) population of Charsada district.

Alleles	DYS 576	DYS 389I	DYS 448	DYS 389II	DYS 19	DYS 390	DYS 481	DYS 549	DYS 533	DYS 438	DYS 437	DYS 570	DYS 635	DYS 390	DYS 439	DYS 392	DYS 643	DYS 380	DYS 458	DYS 385A	DYS 385B	DYS 456	YGATA H4
9						0.1333			0.0133	0.2133							0.1467					0.0267	
10						0.5467			0.0400	0.2267				0.4533	0.1200	0.6533			0.0133				0.0133
11						0.3067		0.0800	0.2000	0.5467				0.1867	0.7067	0.1467	0.1333		0.3867				0.2400
12		0.26667				0.0133		0.4000	0.7467	0.0133				0.1600		0.0533	0.1333		0.0400				0.6933
13		0.65333						0.5200						0.2000				0.5867		0.2800	0.0133	0.0267	0.0533
14	0.0133	0.08000			0.2800						0.5600					0.1467		0.0533	0.0267	0.2400	0.4267	0.0267	
15	0.0133				0.0800						0.2533	0.2667						0.0933	0.5333	0.0133	0.0400	0.6400	
16	0.2267				0.5067						0.1867	0.0933				0.0267			0.1467		0.0667	0.2933	
17	0.1600				0.0133							0.1600							0.2133		0.0267	0.0133	
18	0.4667		0.1200									0.0533	0.0133						0.0267		0.0400		
19	0.0933		0.2533				0.0400					0.2133							0.0400		0.1867		
20	0.0267		0.5333	0.0133								0.1467	0.0133								0.2000		
21			0.0933									0.0533	0.2000										
22							0.0133						0.0400	0.1867									
23							0.4800						0.4933	0.2400									
24				0.0133			0.2933						0.2133	0.5200									
25							0.0267						0.0133	0.0533									
26				0.0133			0.1200						0.0133										
27																							
28				0.2400																			
29				0.1200																			
30				0.5200																			
31				0.0800																			
GD	0.7049	0.50234	0.6368	0.6595	0.5674	0.5971	0.6572	0.5708	0.4061	0.6123	0.5953	0.8282	0.6778	0.6429	0.7034	0.4703	0.5344	0.6169	0.6446	0.7214	0.7445	0.5096	0.4649

4.8

.5 MohmandCharsada

Allele frequencies for all alleles are given in table: 12. The highest allele frequency was found to be 0.8243 for DYS393 in this population while the lowest allele frequency recorded was 0.0135 for DYS576(21), DYS448(18), DYS19(17), DYS549(14), DYS355(9), DYS481(18), DYS438(8), DYS570(14), DYS635(26), DYS392(10), DYS385a(16), DYS385b(20), DYS456(17) and GATAH4(14). The most polymorphic among all the loci in this population was DYS570 with 10 alleles ranging from allele number 13 to allele 21.

Table:

12. Allele Frequencies of 23 Y STRs in Mohmand (MM) population of Charsada district.

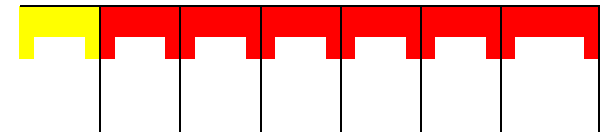
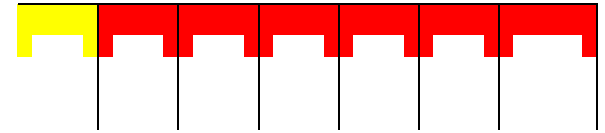


Table:

Alleles	DYS 576	DYS 389I	DYS 448	DYS 389II	DYS 19	DYS 391	DYS 481	DYS 549	DYS 533	DYS 438	DYS 437	DYS 570	DYS 635	DYS 390	DYS 439	DYS 392	DYS 643	DYS 385	DYS 458	DYS 385c	DYS 383c	DYS 456	YGATA H4
6																							
7																							
8										0.0135													
9									0.0135	0.0946							0.0405			0.0405			
10						0.3784			0.1622	0.2297					0.5270	0.0135	0.7297						0.0270
11						0.6216		0.1216	0.0676	0.5946					0.2568	0.7973	0.0135	0.0541		0.6216			0.1892
12		0.1216						0.4459	0.7027	0.0676					0.1081		0.2162	0.0811		0.0405			0.5541
13		0.7973			0.0270			0.4189	0.0541			0.0270			0.0946	0.0405		0.8243		0.2162		0.0405	0.2162
14		0.0811			0.1757			0.0135			0.5811	0.0135			0.0135	0.0811		0.0405		0.0676	0.5811	0.0270	0.0135
15	0.0676				0.2568						0.2162	0.0270				0.0270			0.4189		0.0811	0.5946	
16	0.2162				0.5270						0.2027	0.1081				0.0405			0.2432	0.0135	0.2297	0.3243	
17	0.1486				0.0135							0.2432							0.1351		0.0405	0.0135	
18	0.1892		0.0135				0.0135					0.1081							0.0811		0.0541		
19	0.2838		0.2432				0.1351					0.3514							0.0405				
20	0.0811		0.5541									0.0676							0.0541		0.0135		
21	0.0135		0.1892				0.0405					0.0541	0.0676	0.0405									
22							0.0405						0.0405	0.0811					0.0270				
23							0.3919						0.4595	0.2027									
24							0.1216						0.4054	0.5405									
25							0.2162						0.0135	0.1351									
26							0.0405						0.0135										
27																							
28				0.081																			
29				0.243																			
30				0.527																			
31				0.135																			
32				0.014																			
GD	0.8145	0.3477	0.6061	0.647	0.6331	0.4769	0.7719	0.6220	0.4787	0.5879	0.5812	0.7956	0.6264	0.6490	0.6442	0.3584	0.4247	0.3136	0.7453	0.5665	0.6064	0.5461	0.6179

Table:

-



4.9 Principle Coordinate Analysis

A Principle Coordinate (PCoA) plots was constructed using data from the five populations from the study area and other neighboring populations available on YHRD with accession number (Afghanistan (Pathan): YC000226; Xinjiang (Uighur): YA004122; Tibet China (Tibetan): YA004005; Xingjiang (Kazakh): YA003848; Greece (Greeks): YC000125; India (Tamil): YC000123; Israel: YC000064; Iran: YA003782; Iraq (Iraqis): YC000007; Russia: YC000079; Turkey: YC000296, YC000173, YA003467; Pakistan (Yousafzai): YC000226; Pakistani (Pathan): YC000147). The clustering pattern revealed that all the Pathan populations (including previously studied Pathan populations) from Pakistan and Afghanistan are clustered together which shows great homogeneity with each other and little genetic sharing with Russian populations as well (Fig.23).

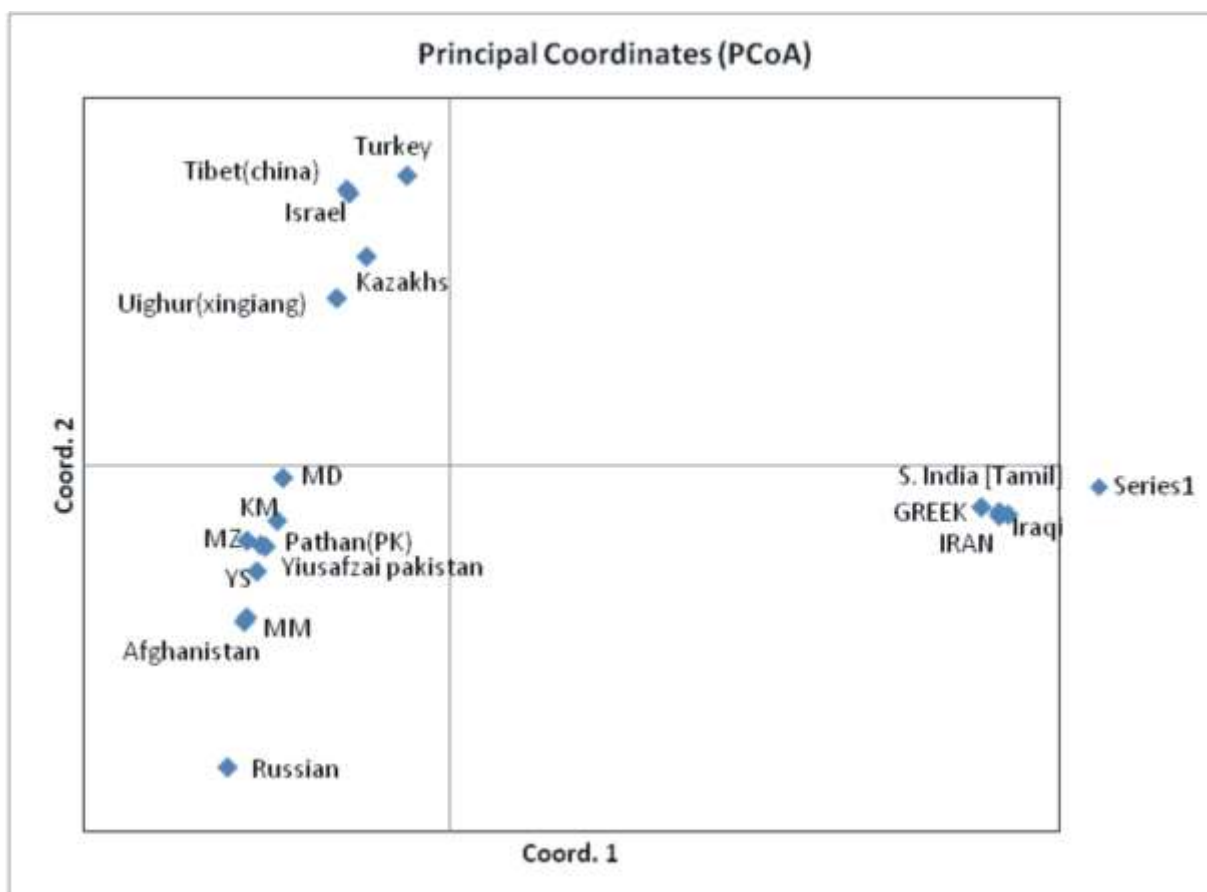


Fig.23. Y-F_{ST} based PCoA plot of five populations of Charsada and Mardan and comparative global populations (*The abbreviations MZ, KM, MM, YS, MD denote the tribes Muhammadzai, Kakakhel Mian, Mohmand from Charsada, Yousafzai and Mohmand from Mardan, respectively*).

4.10 mtDNA HVI analysis

A total of 93 haplotypes were identified in 165 samples (table: 15). 38.7% of the haplotypes were found to be shared among all the populations and 57 haplotypes (61.3%) were singletons.

4.10.1 Genetic Diversity

Genetic diversity in all the samples amplified for HVI control region was 0.993 and

Match probability was 0.987 (table. 13).

Table 13. Comparison of *mtDNA* Genetic diversity among various ethnic groups of Pakistan.

Parameters	Path (this study)	Mkr (Siddiqi <i>et al.</i> 2015)	Pth (Rakha <i>et al.</i> 2011)	Bal (Murci <i>et al.</i> 2004)	Brh (Murci <i>et al.</i> 2004)	Haz (Murci <i>et al.</i> 2004)	Bsh (Murci <i>et al.</i> 2004)	Ksh (Murci <i>et al.</i> 2004)	Par (Murci <i>et al.</i> 2004)	Snd (Murci <i>et al.</i> 2004)	Sar (Hayat <i>et al.</i> 2015)
No. of Sample	165	100	230	39	38	23	44	44	44	23	85
No. of haplotypes	93	70	157	26	22	21	32	12	22	21	63
No of unique haplotypes	57	54	128	18	15	19	25	5	12	19	58
Genetic Diversity	0.993	0.968	0.993	0.974	0.952	0.992	0.980	0.851	0.950	0.992	0.957

Note: The abbreviations Path, Mkr, Pth, Bal, Brh, Haz, Bsh, Ksh, Par, Snd and Sar stands for Pathan, Makrani, Pathan, Baluch, Brahui, Hazara, Burusho, Kalash, Parsi, Sindhi and Saraiki respectively

4.10.2 mtDNA statistics

Types and frequency of variations observed in the mitochondrial HVI sequences of all the five populations is given in the table: 14.

Table: 14. Statistic parameters of five populations of Charsada and Mardan.

Population	MZ(n=28)	KM(n=35)	MM(n=31)	YS(n=36)	MD(35)	Mean	S.d
No. of Transitions	22	18	35	15	20	22.0	7.714
No. of Transversion	4	6	3	2	4	3.8	1.483
No. of Substitutions	26	24	38	17	24	25.8	7.629
No. of Indels	0	2	4	1	2	1.8	1.483

Note: The abbreviations MZ, KM, MM, YS, MD and S.d stands for Muhammadzai, Kakakhel Mian, Mohmand from Charsada, Yousafzai, Mohmand from Mardan and Standard deviation respectively.

Table: 15. List of SNPs defining mtDNA HVI Haplogroups in five Pathan populations of Charsada and Mardan district.

Samples	Macro Haplogroup	Haplogroup	Frequency	Origin	No. of variants	SNPs
KM01	R	U2e	4	S.Asian	7	A16051G, G16129C, A16182C, A16183C, T16189C, A16194C, C16197G

KM02	R	H2(T152C T16311C)	SW.Asian/Middle Eastern	5 1	T16311C
KM04	M	M3	S.Asian	7 4	T16126C, C16223T, T16311C, T16519C
KM06	R	J1b1a1	Middle eastern	5 7	C16069T, T16126C, G16145A, T16172C, C16222T, C16261T,
G16474T			S.Asian		
KM08	R	U5a2	S.Asian	1 5	C16192T, C16256T, C16270T, C16278T, G16526A
KM10	R	U2c	SW.Asian/Middle Eastern	2 2	A16051G, C16179T
KM11	R	I1	SW.Asian/Middle Eastern	1 8	G16129A, A16180G, C16223T, A16254G,
A16293C, T16311C, G16391A, T16519C			Middle eastern		
KM12	R	R6(G16129A)	S.Asian	3 4	G16129A, G16213A, T16362C, T16519C
KM13	R	J1c	S.Asian	1 6	C16069T, T16126C, C16168T, C16266T, C16278T, T16311C
KM14	R	U6c	SW.Asian/Middle Eastern	3 5	G16129A, C16169T, A16183C, T16189C, A16194C
KM15	M	M5a1	Middle eastern	1 5	G16129A, C16223T, C16234T, C16291T, T16519C
KM16	R	H1	S.Asian	1 2	A16051G, C16239T
KM17	R	J1b5b	S.Asian	2 7	C16069T, T16126C, G16145A, C16261T, T16263C, C16290T,
T16519C			S.Asian		
KM18	R	U2e1f	SW.Asian/Middle Eastern	A16051G, G16129C, A16182C, A16183C, T16189C, A16194T, C16221T, T16311C, T16325C, T16362C,	
KM20	M	M33	S.Asian	1 11	C16447T
KM25	M	M6a1b	SW.Asian/Middle Eastern	1 4	C16223T, T16324C, T16357C, C16527T
KM27	R	R8a1a3	C.Asian	2 5	C16188T, C16223T, T16231C, T16362C, T16519C
KM28	M	M4	S.Asian	2 4	C16259T, C16292T, A16497G, T16519C
T16519C			C.Asian	3 7	T16086C, C16111T, G16145A, C16223T, C16261T, T16311C,
KM30	R	H6	S.Asian	1 2	T16362C, A16482G
KM31	N	N7	SW.Asian/Middle Eastern	2 3	G16129A, C16223T, T16519C
KM32	R	U2e1h	SW.Asian/Middle Eastern	2 6	A16051G, G16129C, A16183C, C16193CC, T16362C, T16519C
KM34	M	C4a	Middle eastern	1 5	T16093C, G16129A, T16298C, C16327T, T16519C
KM35	R	R2	S.Asian	5 2	C16071T, T16519C
KM38	R	H10((T16093C))	SW.Asian/Middle Eastern	3 4	T16093C, C16278T, G16319A, T16519C
MD02	R	H2a2a1d	2 3		T16172C, A16182AC, A16183C
MD03	N	N1a1a1a	1 7		C16147A, T16172C, C16223T, C16248T, C16320T, C16355T, T16519C
MD04	R	U2a	1 3		A16051G, A16206C, T16311C
MD05	R	H2a3	1 5		T16093C, C16223T, C16234T, G16274A, T16519C
			SW.Asian/Middle Eastern		
			E.Asian		
MD07	N	W4	2	IndoEuropean	5 T16086C, C16223T, C16286T, C16292T, T16519C
MD08	N	W	2	IndoEuropean	5 C16192T, C16223T, A16284G, C16292T, T16519C

MD09 R H2a2a1g 6 4 T16172C, A16182AC, A16183C, T16189C MD10 M G2a1d2 1 4 C16223T, A16227G, C16278T, T16362C MD12			R	U4a3	1	3
T16356C, T16362C, T16519C						
MD13	R	U7	S.Asian	4 5	A16309G, A16318T, A16343G, T16362C, T16519C	
MD18	R	U5a1b(T16362C)		2 6	C16192T, C16256T, C16270T, T16362C, A16399G, T16519C	
MD20	M	M5		2 3	G16129A, C16223T, T16519C	
MD23	M	M65a(C16311T)		2 5	T16126C, C16223T, A16289G, A16399G, T16519C	
MD25	M	M5a2a1a		1 4	G16129A, C16223T, A16265C, T16519C	
MD26	R	H94		1 2	C16339T, C16355T	
MD29	N	N	SW.Asian/Middle Eastern	6 3	C16223T, C16292T, T16519C	
MD31	R	H5	C.Asian	4 3	C16266T, T16304C, T16519C	
MD34	M	D4q	SW.Asian/Middle Eastern	1 6	C16223T, C16256T, T16311C, G16319A, T16362C, T16519C	
MD36	M	M52a	E.Asian	1 5	C16223T, A16275G, G16303A, G16390A, T16519C	
MD38	M	M71	S.Asian	1 2	C16223T, T16271C	
MD39	R	T1a	S.Asian	1 6	T16126C, A16163G, C16186T, T16189C, C16294T, T16519C	
MM01	R	P6	W.Eurasian	1 6	C16221T, T16311C, T16325C, T16362C, C16447T, T16519C	
MM02	M	M2a1	SE.Asian	1 7	T16075C, C16223T, C16270T, G16274A, G16319A, T16352C,	
T16519C			S.Asian			
MM03	R	H15a	SW.Asian/Middle Eastern	2 1	C16184T	
MM07	R	T1a1'3	W.Eurasian	2 6	T16126C, A16163G, C16186T, T16189d, C16294T, T16519C	
MM11	M	M49	S.Asian	2 3	C16223T, C16234T, T16519C	
MM13	R	U2e3	S.Asian	1 4	C16168T, C16234T, C16260T, T16362C	
MM15	R	H13a1d	SW.Asian/Middle Eastern	1 1	C16234T	
MM20	N	Y2	C.Asian	1 5	T16126C, C16223T, C16266T, T16311C, T16519C	
MM22	R	H2a2b	SW.Asian/Middle Eastern	1 2	C16291T, T16311C	
MM23	R	H1e	S.Asian	2 3	G16129A, A16182AC, A16183C	
MM26	R	R0a	SW.Asian/Middle Eastern	2 3	T16126C, T16362C, T16519C	
MM27	R	H4a	S.Asian	1 1	C16287T	
MM29	M	M18b	SE.Asian	1 5	A16160G, C16223T, A16318T, T16325C, T16519C	
MM30	R	P4a	SW.Asian/Middle Eastern	1 3	A16037G, C16111T, G16319A	
MM35	R	H14b1	C.Asian	1 2	T16126C, T16519C	
MM36	M	C4a1(G16129A)	S.Asian	4 5	G16129A, C16223T, T16298C, C16327T, T16519C	
MM38	M	M2b	SW.Asian/Middle Eastern	1 1	C16169CC	
Mz04	R	H1bt	SE.Asian	1 6	C16292T, C16355T, T16406d, A16497G, T16519C, C16527T	
MZ05	R	F1c1a		1 3	C16111T, G16129A, T16304C	
MZ06	N	W6		6	C16192T, C16223T, C16292T, T16325C, C16465T, T16519C	
			2	IndoEuropean		

MZ11	R	H13a1a1d	SW.Asian/Middle Eastern	1 6	T16086C, G16145A, C16173T, C16261T, T16311C, T16519C
MZ16	R	I6a	SW.Asian/Middle Eastern	1 8	G16129A, C16223T, A16293C, T16311C, T16362C,
G16391A, C16447T, T16519C			SW.Asian/Middle Eastern		
MZ22	R	H3p	Middle eastern	1 3	C16069G, C16222T, A16269G
MZ24	R	J1b3	S.Asian	1 6	C16069T, T16126C, G16145A, C16222T, A16235G, C16261T
MZ27	M	M39a	W.Eurasian	1 1	C16353T
MZ28	R	T1	SW.Asian/Middle Eastern	1 7	T16093C, T16126C, A16163G, C16186T, T16189C, C16294T,
T16519C			W.Eurasian		
MZ33	R	H14a	S.Asian	1 3	C16256T, C16270T, T16352C
MZ34	R	T2e	S.Asian	1 5	T16126C, G16153A, C16294T, C16296T, T16519C
MZ36	R	U2b2	SW.Asian/Middle Eastern	1 2	A16051G, C16239T
YS01	M	M2a1a	Middle eastern	2 6	C16223T, C16270T, G16319A, T16352C, C16449T, C16451T
YS03	R	H2a2a1c	W.Eurasian	4 8	A16051G, T16086C, C16259A, C16267T, C16291T,
A16300G, A16326G, C16353T			S.Asian		
YS04	R	JT	Middle eastern	1 3	T16086C, T16126C, T16519C
YS06	R	HV2	SW.Asian/Middle Eastern	1 2	T16217C, C16446T
YS07	R	U1a2	C.Asian	1 4	G16129A, T16189C, C16192T, A16202C
YS09	R	J1b4a	S.Asian	1 6	C16069T, T16126C, G16145A, C16218T, C16261T, C16287T
YS11	R	H17a1	W.Eurasian	1 4	G16129A, C16223T, C16291T, T16519C
YS12	M	C4a2'3'4	S.Asian	1 6	C16223T, T16297C, T16298C, C16327T, T16357C, T16519C
YS13	R	U2a1a	C.Asian	1 6	A16051G, T16154C, A16206C, A16230G, T16311C, T16519C
YS19	R	T2b2b	SW.Asian/Middle Eastern	1 5	C16111A, T16126C, C16294T, C16296T, T16519C
YS20	M	M3c2	Middle eastern	1 4	T16126C, T16154C, C16223T, T16519C
YS21	M	C4a4b	S.Asian	1 8	T16086C, G16129A, C16150T, C16223T, T16298C,
C16327T, T16357C, T16519C			C.Asian		
YS23	R	H1k		1 4	A16051G, T16189C, C16290T, C16292T
YS24	R	J1b		2 5	C16069T, T16126C, G16145A, C16261T, T16519C
YS30	R	R		1 3	C16292T, A16497G, T16519C
YS31	N	N1b1		1 7	G16145A, C16176G, C16223T, C16256T, A16309G, G16390A, T16519C
YS36	L3	L3	1	5	T16093C, G16129A, C16223T, A16305T, T16519C
YS37	M	M38d	S.Asian	1 5	G16129A, C16223T, C16266T, T16311C, T16519C
YS38	R	R6b	E.Asian	2 7	C16179T, A16227G, C16245T, C16266T, G16274A, C16278T,
T16362C					

4.10.3 Mitochondrial DNA HVI Haplogroups found

To characterize the maternal genetic variation among the populations, haplogroup frequencies were calculated. Among the 93 haplotypes, 63.4% of the samples belong to the haplogroup R while haplogroup M, N and L were found with the frequency of 26.8%, 8.6% and 1.1% respectively (Fig.24).

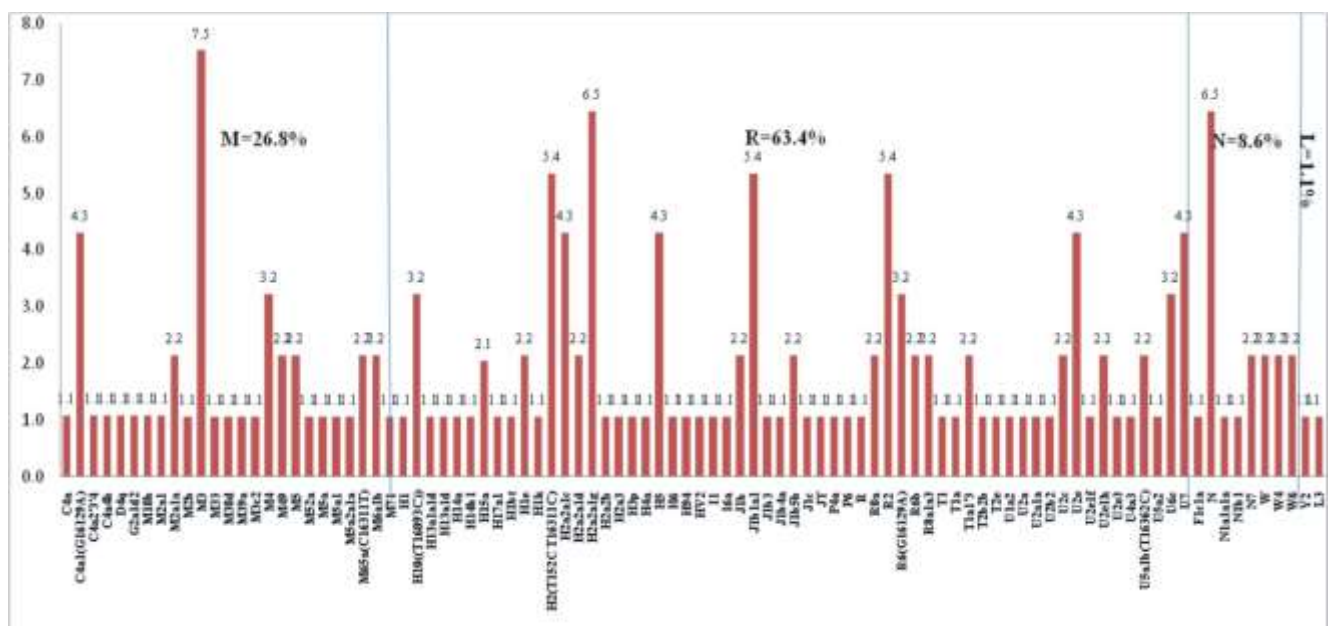


Fig.24. mtDNA HVI Haplogroup frequencies among five Pathan populations of Charsada and Mardan.

4.10.4 Network Analysis

To obtain insight into the genetic structure of the studied populations Median joining Networks were constructed for all the major haplogroups based on the frequency of haplotypes in these five populations using program NETWORK 4.6.1.3.

3.10.5 Macro Haplogroup R

Little sharing of haplotypes was observed among populations in haplogroup R and a “star haplotype” with total frequency of 47.8% in MM, 17.4% in MD, 13.04% in MZ and KM and 8.7% in YS was found in this haplogroup sequences (Fig.25a).

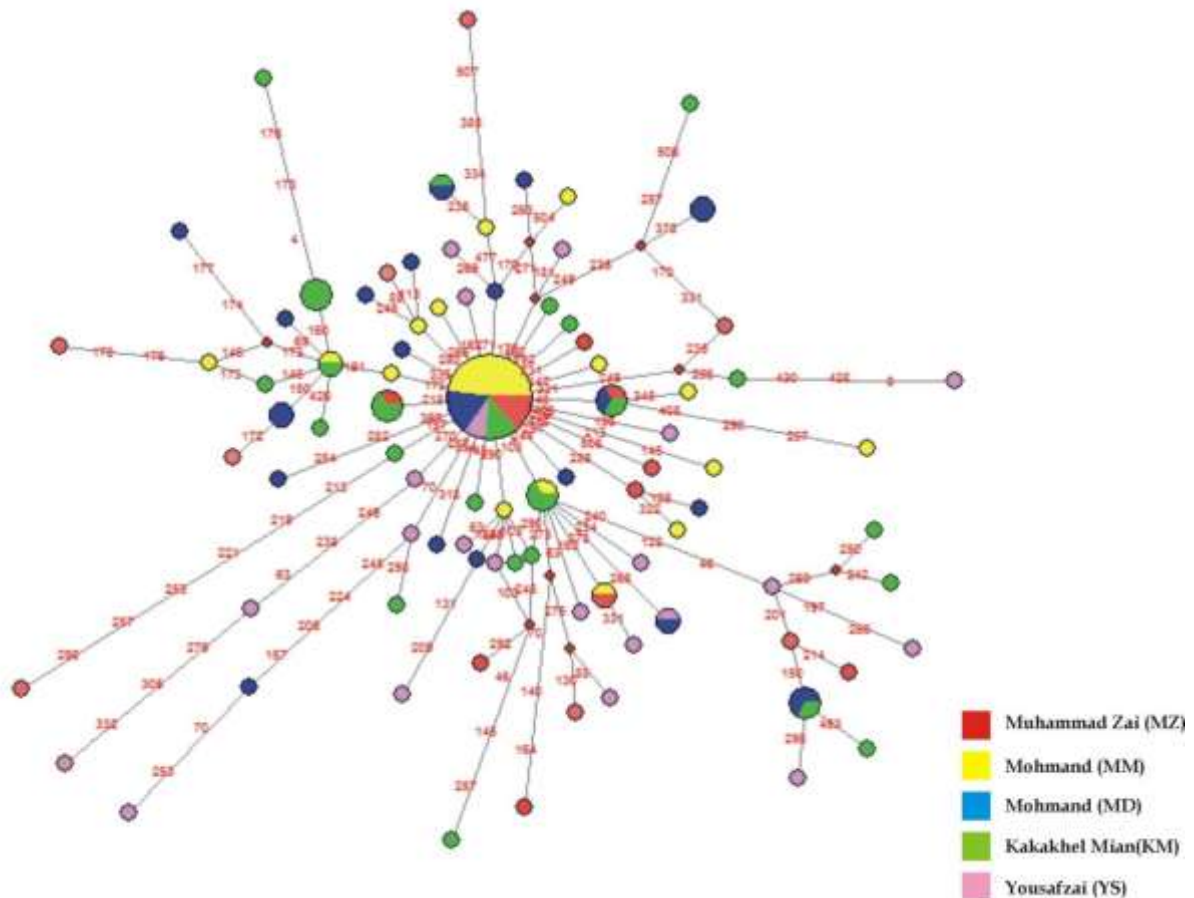


Fig.25.a. Median-Joining network of mtDNA haplogroup R in five populations of Charsada and Mardan. Areas of circles are proportional to the haplotype frequencies.

4.10.6 Macro Haplogroup M

Very little sharing of haplotypes was observed in Hg M. and (Fig.25 b).

There was no haplotype found shared among all the five populations. Two haplotypes were shared between Mohmand from Charsada (MM) and Yousafzai (Ys) from Mardan. Similarly single haplotype was shared between Kakakhel Mian (KM) and Muhammadzai (MZ).

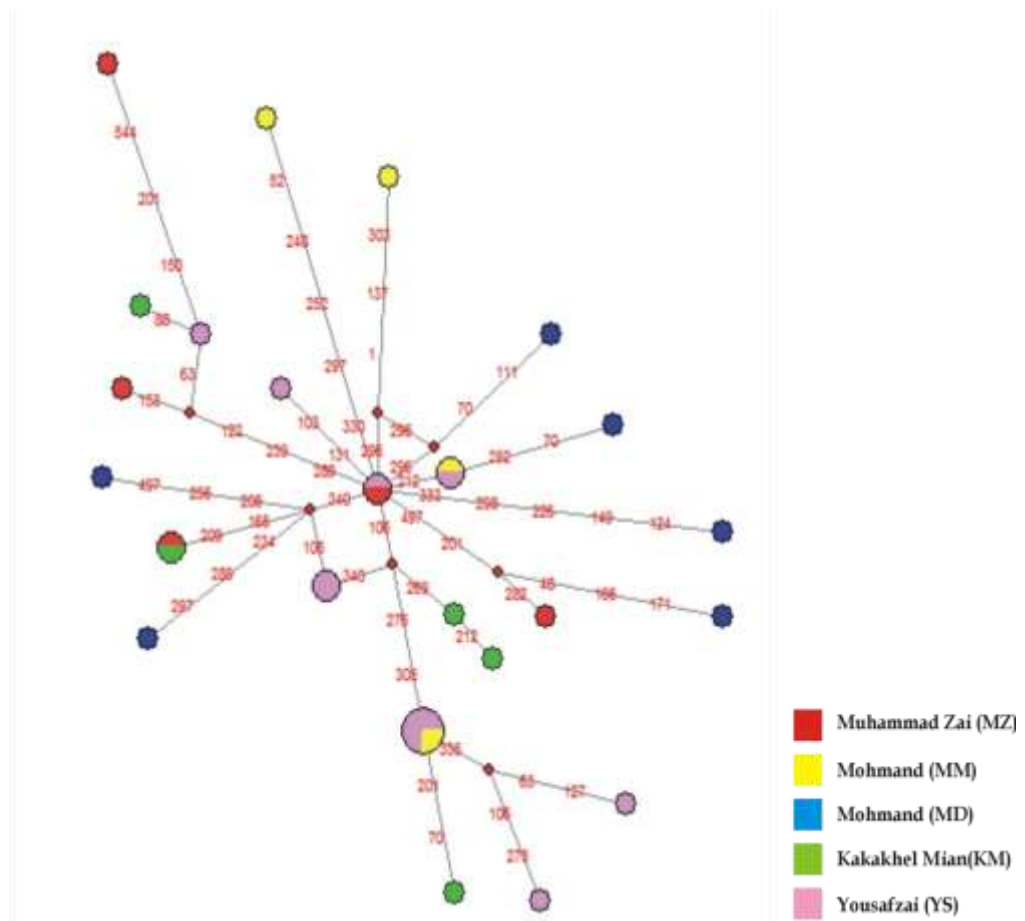


Fig.25.b. Median-Joining network of mtDNA haplogroup M in five populations of Charsada and Mardan. Areas of circles are proportional to the haplotype frequencies.

4.10.7 Macro Haplogroup N

This haplogroup is present in 8.6% of the samples and no haplotype sharing was recorded in Hg N (Fig. 25c).

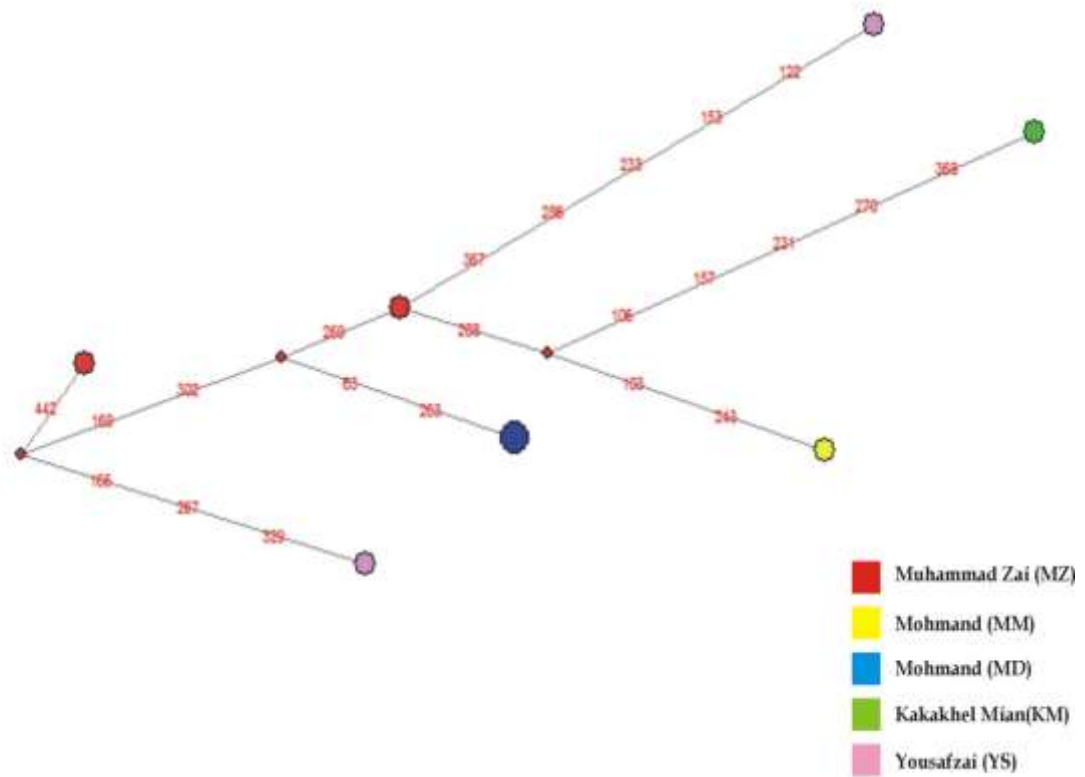


Fig.25.c. Median-Joining network of mtDNA haplogroup N in five populations of Charsada and Mardan. Areas of circles are proportional to the haplotype frequencies. mtDNA HVI F_{ST} among all the five populations is represented in the fig.26.

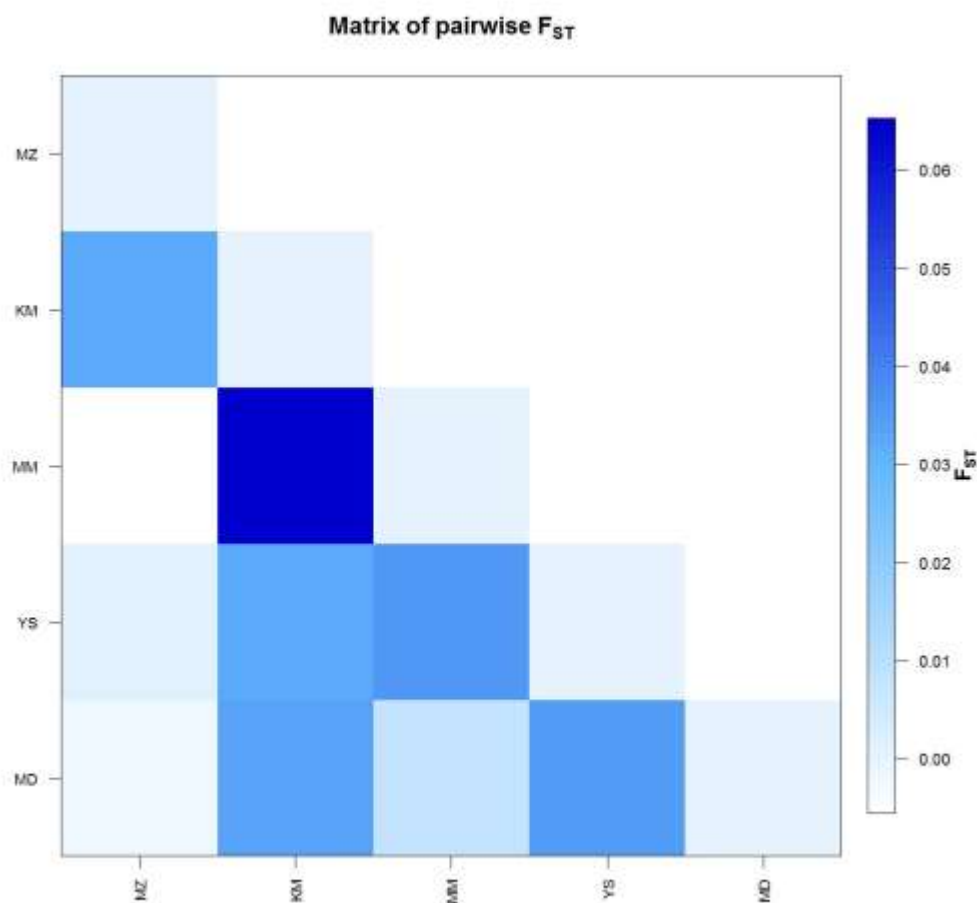


Fig.26. mtDNA HVI based Pairwise genetic distance (F_{ST}) among five population pairs. (The abbreviations MZ, KM, MM, YS, MD denote the tribes Muhammadzai, Kakakhel Mian, Mohmand from Charsada, Yousafzai and Mohmand from Mardan, respectively).

4.10.8 Analysis of Molecular variance (AMOVA)

The proportion of genetic variation distributed within and between the populations of two districts was assessed by analysis of molecular variance (AMOVA). Percentage of variation among groups was -1.57% and variation among populations within the group was 3.63% while percentage of variation within populations was found to be 97.94% (table: 16).

Table: 16. AMOVA Analysis (mtDNA HVI Haplotypes) of five populations from two districts (groups).

Source of Variation	Degree of freedom	Sum of Squares	Variance Components	Percentage of Variation
Among Groups	1	1.270	-0.02156 Va	-1.57
Among population within Groups	3	9.600	0.04977 Vb	3.63
Within populations	182	244.445	1.34311 Vc	97.94
Total	186	255.316	1.37131	

4.10.9 Principle Coordinate Analysis

F_{ST} values were calculated for all pair of populations in Arlequin V.3.5.2.2 (Excoffier *et al.*, 2010). No significant genetic distance (F_{ST}) was observed between population pairs except for the Kakakhel Mian and Mohmand. The PCoA plot provided information regarding the degree of association between different indigenous and global populations using comparative sequence data from Greece, Russia, Iraq, Iran, Israel, Afghanistan, Xinjiang china, Turkey, South India, North Indian Afridi, Balouchi, Brahui, Brusho, Kalash and Pakistani Pathan population. Clustering pattern revealed that Mohmand (MD) and Yousafzai from Mardan are clustered together with Balouchi and Pathans from Pakistan and Afridi pathans from India while Muhammadzai and Kakakhel Mian from Charsada are clustered together with Brahui and Brusho from Pakistan. On the other hand Mohmand (MM) from Charsada clusters with Turkish population, whereas Tamil from

South India and Kalash from Pakistan clustered separately (Fig.27).

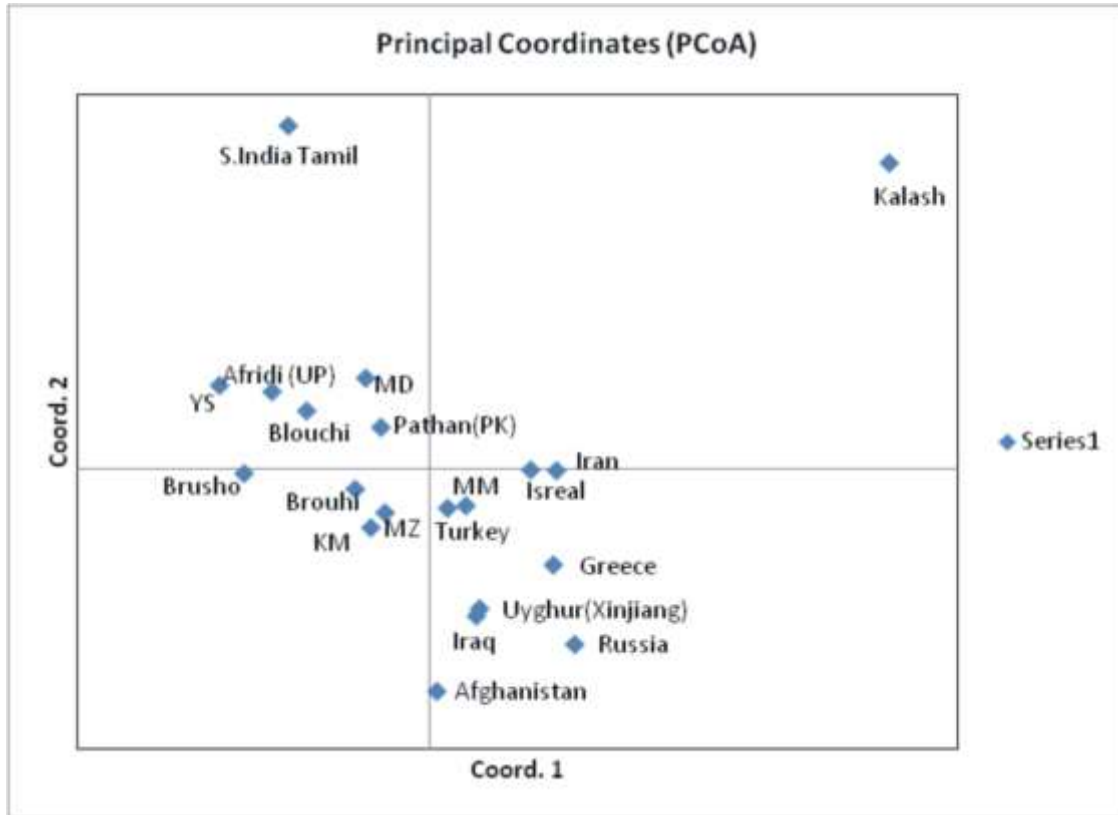


Fig.27. mtDNA- F_{ST} based PCoA plot of five populations of Charsada and Mardan and comparative indigenous Pakistani and global populations. (The abbreviations MZ, KM, MM, YS, MD, denote the tribes Muhammadzai, Kakakhel Mian, Mohmand from Charsada, Yousafzai and Mohmand from Mardan, respectively).

4.11 Spatial correlation of data

Result of the Mantel Test revealed that no significant correlation existed between the genetic distance obtained from Y-Fst or mtDNA-Fst and geographic distance. A very little positive correlation (Table: 17) was recorded between Y-Fst and mtDNA-Fst values.

Table17. Correlation between genetic distance based on Y-Fst or mtDNA-Fst and geographic distance

Y Chromosome, mtDNA and Geography		Coefficient	P value
Correlation Coefficient	Y-chromosome / mtDNA	0.21	0.13

Y-chromosome / Geography	-0.45	0.04
mtDNA / Geography	-0.07	0.46

Chapter-5

DISCUSSION

Several Pathan tribes are residing in Pakistan, whose origin and affinities are still question mark. A research endeavor was carried out to elaborate Pathan population of Mardan and Charsada using 17 STR loci with high Discrimination Capacity reported by Lee *et al.* (2014). It reported that the 17 loci in Y-filer had low haplotype diversity in Pathan populations of Pakistan, but diversity was considerably increased with additional useful STR loci. In the present study, increased haplotype diversity and considerably increased discrimination capacity was observed in different Pathan populations of Charsada and Mardan with the use of 23 Y-STR loci which signals that the additional six loci in Promega PowerPlex® Y23 System are very useful for genetic investigations of these populations. Pathans generally have been reported to have low haplotype diversity due to patrilocality (Vermeulen, M. *et al.*, 2009; Mohyuddin, 2001; Ghosh, 2011) but here the increased haplotype diversity has been observed among these tribes using 23 Y-STR loci. The most polymorphic locus among them was DYS570 and being rapidly mutating (RM) locus such markers are good candidates for population differentiation (Ballantyne *et al.*, 2010; Ballantyne *et al.*, 2012).

Genetic finding of Haber *et al.*, (2012) shows that the Pathans of Afghanistan exhibit genetic affinity with the North and west India and they split from rest of the afghans

4.7kya during the rise of area" s first civilizations at Indus valley and Bactria-Margiana.

The main lineage in Pathans of Afghanistan has the oldest coalescent time of 14kya in Indus valley.

The Y-haplogroup frequencies among the Pathan populations of Mardan and Charsada Districts show that they exhibit a blend of haplogroups of different origins, with contribution of 71.3% of South Asian, 15.73% from Middle Eastern, Caucasian or GrecoAnatolian and 10.4% of Central Asian origin besides little or negligible percentage of haplogroups of African origin, e.g. haplogroup E1b1a and E1b1b which are sub Saharan African and Arabian specific haplogroups. The E1b1b1-M35 lineages in some Pakistani Pathan were previously traced to a Greek origin brought by Alexander" s invasions (Firasat *et al.*, 2007). However, RM network analysis of E1b1b1-M35 revealed that Afghanistan" s lineages were correlated with Middle Easterners and Iranians but not with populations from the Balkans. The Islamic invasion in the 7th century CE left an immense cultural impact on the region, with reports of Arabs settling in Afghanistan and mixing with the local population (Emadi, 2005). However the genetic signal of this expansion is not clearly evident as some Middle Eastern lineages such as E1b1b1-M35 were also found in Afghanistan.

The high frequency of Y haplogroup R1a in all of the five populations has been noticed. Almost equal frequency of this haplogroup in Pathans from Afghanistan has been

observed that indicates the genetic similarity between the two (Lacau *et al.*, 2012) and similarly equal frequency of this haplogroup has been recorded in Pakistani Pathans also (Lee *et al.*, 2014). South Central Asia i.e. Pakistan and India (Underhill *et al.*, 2010; Mirbal *et al.*, 2009) are proposed as the most likely places of origin because in both regions R1a1a-M198 has been observed at frequencies 50% or above. According to Underhill *et al.*, 2010, the highest Y-STR haplotype diversity for the R1a1a lineage is observed in South Central Asia with a coalescent time of 14kya, suggesting that this region is the likely source of the dispersal for the M198 mutation. Alternatively, Klyosov claims that haplogroup R1a1a-M198 originated in South Siberia about 20kya. But later the South Central Asian origin of this haplogroup was well supported. Our results revealed the frequency of this haplogroup ranging from 41.3% to 54.6% among all the five populations of this region providing a clear evidence for the local origin of this haplogroup.

Similarly, Y haplogroup L being second most prevalent haplogroup comprising 17.9% of the total haplotypes, has already been reported in Pakistani Pathans with frequency of approximately 7% and with moderate frequency of approximately 2% among other Pakistani populations, whereas its high frequency approximately 23% was reported in Pathans of Afghanistan (Firasat *et al.*, 2007). The Y chromosomal DNA variation in Pakistan by Qamar *et al.*, (2002) reported 14% frequency of Haplogroup L where it was called haplogroup 28, in overall Pakistani population (Qamar *et al.*, 2002), while 5.9% of this haplogroup had been reported in Pakistani Pathans in 2014 (Lee *et al.*, 2014), which is said to be related with the arrival of agriculture in this area of Indian-subcontinent (Qamar *et*

al., 2002). Haplogroup L-M20, was hypothesized to have originated in India or the Middle East (Wells, 2007) approximately 30kya. This marker, which is found at 25% in north Afghanistan and 4.8% in the south, has also been previously reported at high frequencies (48%) in the Kallar community of South India (Wells *et al.*, 2001) as well as in the Druze (35%) population from Israel. (Shen *et al.*, 2004) Time estimates generated based on seven Y-STR loci within L-M20 lineages for north (14.6 ± 7.3 kya) and south (17.8 ± 8.4 kya) Afghanistan populations are intermediate to those of Pakistan (26.3 ± 5.3 kya) and India (7.5 ± 1.7 kya) (Lacau *et al.*, 2012). Furthermore, Pakistan displayed higher haplotype variance (0.548) than India (0.118), suggesting that L-M20 most likely originated in what is today Pakistan rather than in India (Lacau *et al.*, 2012) and findings of present investigation confirms this argument and also support the dispersal of this haplogroup L from its place of origin to Southern Afghanistan.

Likewise, the highest frequencies of Y-chromosome Haplogroup H-M69 are in India and Pakistan especially in 4.1% Burusho, 20.5% Kalash, 4.2% Pathan and 6.3% in other Pakistani populations (Thanseem *et al.*, 2006; Qamar *et al.*, 2002). It is also present in high frequency of 25 – 35% in tribal population and in low frequency of 10% in Western India and Pakistan (Sahoo *et al.*, 2006; Cordaux *et al.*, 2004). The low frequency of Ychromosome Hg R1b, E1b1b and J2 of predominantly Greek lineage in these populations and in the Pathan population of Afghanistan was observed, which strengthen the idea of Firasat *et al.*, (2007) about the limited Greek contribution to the Y chromosome structure of Pathans. The E1b1b1-M35 lineages in some Pakistani Pathan were previously traced to a Greek origin brought by Alexander" s invasions (Firasat *et al.*, 2007).

For mtDNA analysis, it has been noticed that profound genetic variations were accounted within populations (97.97%) rather than between the populations (3.63%). while variance distribution was greater within populations (1.3%) than among the populations (0.04%). This relatively high heterogeneity among these Pathan populations further modeling the mtDNA landscape that might be the result of unequal genetic influence obtained during different settlement events from geographically adjacent neighbors. On account of that, some haplotypes are shared across the tribal boundaries which may be due to maledriven intermarriages.

The mtDNA Macro haplogroups recovered from Pathans of Mardan and Charsada are R, M, N and L. The first three haplogroups are thought to be originated around 60000-75000 years ago in South Asia (Kivisild, 2003). Therefore, the presence of these haplogroups suggests a South Asian maternal origin of these populations. The macro-haplogroup L (Sub-Saharan Africa) was detected in just 0.5% i.e. only on one subject, while East African haplogroups (L4, L5, L6 and L7) are also not detected in Pathans as yet; while all the Arabian Peninsula countries are more likely to have a comparatively high frequency, such as Yemen (38%), Oman (16%) and Saudi Arabia (10%) (Abu-Amro *et al.*, 2007). The most common South Asian haplogroups in Pathans comprises of R (67.9%) and M (22.3%). In the present study, the mitochondrial haplogroups of South Asian origin have highest proportion of 37.6%, comprising M3 (7.5%), R2(5.3%), U7, U2e (4.3%), M4, U6c (3.2%), U2c, M65a, M6a1b, U5a1b, M49, M5, M2a1a, M6a1b, R0a, U2e1h (2.2%) and U5a2, U2e3, U4a3, U2e1f, U2b2, U2a1a, U2a, M52a, M5a1, M33, M38d, M39a, M2b, M18b,

M71(1.1%). The second major haplogroups identified were of Southwest Asian or Middle Eastern origin (36.5%) including H2a2a1g (6.4%), J1b1a1, H2 (5.2%), H2a2a1c, H5 (4.3), H10, R6 (3.2), H15a, H1e, H2a2a1d, R8a1a3, J1b, J1b5b (2.2%). The third most prevalent mitochondrial haplogroups are of Central Asian origin (8.6%) followed by West Eurasian (6.5%), east Asian and Indo-Europeans (3.2%) and the least frequent haplogroup was found L (1.1%) of East African origin. The high frequency of macro haplogroup M had been recorded in Asia, particularly in India, Bangladesh, Nepal and Tibet reaching up to 60 to 80% (Rajkumar *et al.*, 2005). Mitochondrial haplogroup M3 being the sub-clad of HgM, has been found in South Asia, with highest frequency in West India and Pakistan (Metspalu *et al.*, 2004). It has been reported in Pakistani Pathan population with frequency of 7.8% (Rakha *et al.*, 2011) and this study reported the 7.5% occurrence of this haplogroup. This concurrence of mtDNA markers could evident significant female enigmatic contribution towards the genetic structuring and socio-cultural expansion of multiethnic Pathan population (Rakha *et al.*, 2011). Moreover this uniparental genetic landscape of mtDNA HVI data proposed that the Pathans revealed a profound blend of genetic components from South Asia, the Caucasus, Europe, Arabian Peninsula and Central Asia (Quintana-Murci *et al.*, 2004). It is obvious that the majority of Pathan autochthonous M lineage huddled into M5 and M3 sub-haplogroups which have characteristic South Asian origin.

Phylogeographic analysis has revealed that M5a1 haplogroup is most abundant in all Roma Gypsies population, including Hungarian, Bulgarian, Iberian and Balkan Roma (ranging from 6 to 29%), constituted a founder population that distributed across West

Asia and whose origin might be traced back to the Indian Subcontinent, particularly Pakistan (Mendizabal *et al.*, 2011). In these five populations it was found in just 1.1%. Another mitochondrial DNA lineage, the West Eurasian macrohaplogroup N has been proved to be the ancestor of numerous haplogroups prevailing in Europe, Middle East, Asia and the Americas (Kivisild *et al.*, 1999; Nasidze *et al.*, 2008). It is present though less common in Pathans, with the overall frequency of 8.6% in their genetic pool, which is in agreement with previous studies (Quintana-Murci *et al.*, 2004; Rakha *et al.*, 2011). The haplogroup R (sub-clad of N) cuddled the majority of the West Asian and European haplogroups H, HV, J, T and U, which have varying frequencies in five Pathan populations (Malmstrom *et al.*, 2015). The incidence of overwhelming immensity of R lineage (63.4%) in Pathan has a clear Europe and West Asia attribution due to Paleolithic and Neolithic expansions of Caucasian that reached in South Asia via Iranian plateau and might be Arabian Sea nautical routes, could be the possible argument for this genetic influx (Stoljarova *et al.*, 2016; Haber *et al.*, 2016). Furthermore, haplogroup R0 comprising HV0, HV1 and HV2 was scarcely found in these five Pathan populations (1.15%) as compared to other Pakistani population, having highest frequency in Balochi (10.3%), Pathans (9.1%) and absent in Burusho and Hazara (0%) (Kivisild *et al.*, 2002; AlvarezIglesias *et al.*, 2009). However, neighboring countries exhibited low frequency such as Uzbek (2.4%), Turkmen (2.4%) and in Turkish its frequency dropped to 0% (Rakha *et al.*, 2011; Siddiqi *et al.*, 2015). The haplogroup H was 9.8% prevalent in overall five Pathan populations but different from West Asia (45%) and Near East (25%) (Palanichamy *et al.*, 2015). Whereas neighboring population exhibit relatively high frequency of this haplogroup, such as Iranian (14.3%),

Uzbek (21.4%), Turkmen (22%) and Tajik (29.5%) (Whale, 2012; Derenko *et al.*, 2013). Similarly other Pakistani populations exhibited very high frequency of haplogroup H i.e. Sindhi (28%), Brahui (26.3%), Balochi (20.5%) but moderate in Burusho (12.3%) and Hazara (13%) population (Quintana-Murci *et al.*, 2004; Bhatti *et al.*, 2016). Notably, under these assumptions it is hard to discern sequential gene flow or expansions at the population level because the most recent migration i.e. British colonization in the 18th century could hold both early and derivative lineages, which contributed significantly in the genetic pool of the Pathan (Jons, 2016). Besides aforementioned haplogroups, analysis of specific haplogroups J revealed some interesting geological and chronological distribution in Pathan. It was found with the frequency of 9.6% reflected a deep Jewish and European ancestry conglomeration. It is noteworthy that expansion of the Jewish community particularly Ashkenazi, across the Roman Empire and Iranian Plateau, was going to be around 1000 BC (Mosk, 2013). However, on account of expulsion in the Western Europe during the 15th century, they subsequently dispersed into Eastern Europe and moved up to Afghanistan, North West Pakistan and India (Behar *et al.*, 2004; Shlush *et al.*, 2008; Costa *et al.*, 2013). Yet, Pakistani and Indian Jewish communities are an early branch of the Jewish Diaspora with their several exclusive socio-cultural features, intricate history and minor Middle East specific ancestry components (Oddie, 1991; Ostrer, 2001; Chaubey *et al.*, 2016). Another distinguished mitochondrial super-haplogroup U that was most abundant in Pathan (21.8%), which includes haplogroups U2, U5 and U7.

The haplogroup U was widely distributed in the Europe and Near East with the coalescence age of about 51,000–67,000 years before present (Roostalu *et al.*, 2007).

However, this widespread distribution, when incorporated with the presence of a profound associated haplogroup U2 found in native South Asians, suggests that haplogroup U2 is very much entrenched, divergent and was presumably a former lineage of the super-haplogroup U in South Asia (Barcaccia *et al.*, 2015). This statement was further strengthened by the incidence of U2 haplogroup in excavated human remains in the Southern Russia at around 30 000 years before present i.e. during Pleistocene expansion. Due to frequent distribution of U2 in South Asian and European huntergatherer lineages, it has practically predicted some association among „Paleolithic Hunters“ and present day native inhabitants (Krause *et al.*, 2010). Another aforementioned mitochondrial lineage, U7 has originated in the „Black Sea“ zone between Southeastern Europe and Western Asia dated back to 51 000–67 000 years before present. It is markedly found among India (20%) and Near East (10%) (Roostalu *et al.*, 2007) and found 4.3% in these Pathan, 23% in North Indians, 10% in South Indians, 7.2% in Iran while absent in the Western Asian or Anatolian and Eastern European or Balkans population (Palanichamy *et al.*, 2015; Bahmanimehr *et al.*, 2015; Chaitanya *et al.*, 2015). This implied some degree of complex overlapping of haplogroups, across the entire Persia, Pakistan (KPK) and India during various migratory proceedings that was an outcome of succeeding historical and geographic gene flow (Bahmanimehr *et al.*, 2015). Despite primary sub-haplogroups of U, one European-specific haplogroup U5 exhibited relative frequency of 3.3% in Pathan. Earlier investigations of ancient mtDNA proposed that U5 haplogroup was most frequent in Mesolithic and Neolithic Europeans. For example, a high frequency (65%) of U5

haplogroups has been assessed in the European hunter-gatherers individuals (Malmstrom *et al.*, 2009; Sanchez-Quinto *et al.*, 2012; Cassidy

et al., 2016).

The frequency of Y-haplogroups and mtDNA haplogroups on the basis of their origin in these Pathans is almost similar with a little difference. The plausible explanation for this difference may be the limited sample size typed for mtDNA analysis and a different picture could be expected by increasing the sample size for populations.

The frequency of these mtDNA macro haplogroups found in the populations we genotyped in present study is in close agreement with the one already reported in Pakistani Pathans with a little variation in frequency of sub-clades. Mitochondrial haplogroups HV9, M65, H1k, U2a1, M71, M52 and U5a1b were confined to the populations of Mardan (Mohmand: MM, Muhammadzai and Kakakhel Mian) only while haplogroups U7, M18 and P6 were restricted to Charsada populations (Yousafzai and Mohmand: MD) only and absent in Mardan. The high frequencies of local lineages in these populations confirm their descent as a consequence of Paleolithic population expansion (McElreavey *et al.*, 2005).

It is indicative that the pattern of Y-haplogroup distribution in Afghanistan and already studied general Pathan population from Pakistan investigated by Sangupta *et al.*, 2006, show great similarity. In the light of our results and their homology with the previously documented genetic information, it can be inferred that these populations have been descended from Durrani Pathans in South and Southwest Afghanistan which is in agreement with the oral historical tradition that the potential paternal ancestors of these

Pathan populations were Durrani which were descendants of haphthalites rooted from Qais (Allah *et al.*, 2013). On mtDNA- F_{ST} based PCoA scatter plot, Mohmand from Charsada (MM) showed great genetic affinity with Turkey, Israel and Iran positioned somewhere at the middle of these three, depicting their admixed maternal lineage from Turkey, Israel and Iran rather than Afghanistan. Similarly Yousafzai and Mohmand (MD) from Mardan show relatedness with Afridi Pathans from West India. The logical explanation for different clustering on mtDNA variation may be the different origin of matriline and patriline of these populations. But the difference between paternal and maternal origin is not confirmed by although very little positive correlation between distance matrix of mtDNA and Y-STR. In the light of great genetic similarity revealed among the populations studied, results validate the origin of the three of the four Pathan tribes from the single patriline as documented in the legend, i.e. from “Sarban confederacy” While lack of Arabian Y clusters and mtDNA haplogroups in Kakakhel Mian population refute the Arabian descent of Kakakhel Mian. Another logical explanation for different clustering pattern may be the more frequent male movement among the two areas as compared to the females which contribute to the more gene flow in paternal gene pool reducing the differentiation among the males of these populations while strict practice of endogamy within the populations contribute to retain their genetic differentiation on matriline.

CONCLUSION

On the basis of our findings compared with already available scientific information on

Pathans we conclude that vast majority of paternal gene pool of the Pathans from Mardan and Charsada populations exhibit genetic uniformity with other Pathans from Pakistan and Afghanistan, displaying a mosaic gene pool with haplogroup sharing of the Middle Eastern, Caucasian, South Asian and Central Asian gene pools. It can be inferred from our results that these populations are donors of the Pathan population of areas of Afghanistan from where these populations migrated to their present settlements. Furthermore the Pathans have maintained the Genetic diversity irrespective of the geographical location and sub-population or caste. The possible reason for this pattern is the common practice of endogamy restricting their intermixing with other ethnic groups. Moreover our results are congruent with the above stated hypothesis about the legendary Afghan ancestor, where these populations exhibit considerable homogeneity contributed by being a descendant of a single Y chromosome i.e. Sarban. The study presented in this dissertation reports the first hand information with respect to Pathans of KP province which can be extended to other areas of the region with maximum analytical material and tools for establishing intra and inter population affinities of the tribes and clans. This information can be used as a useful database for establishing a forensic baseline for the Pathan population of the region. For complete elaboration of the phylogenetic position of Pathan lineage, complete Mitogenome sequence analysis and Y chromosomal SNP analysis is required.

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APPENDIX I



**ETHNOGENETIC PROJECT DEPARTMENT OF GENETICS HAZARA
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NOTE: This Performa is the property of the **ETHNO GENETIC PROJECT (HEC)** Department of Genetics Hazara University Mansehra Pakistan. This information will be used for research purpose only and will be kept secret.

Ref. #: _____

Lab code: _____

Date: _____

DETAILS OF PARTICIPANT

Name: _____ **Father's Name:** _____

Age: _____ **Sex:** _____ **Native Language:** _____

Ethnic Group: _____ **Caste** _____

Collection point: _____

Home Address: _____

Biological Samples Collected

☐ Dental impressions ☐ Saliva

Consent of Participant

~ The researchers have informed me about the purpose of sample collection and their use in human genetics research. I have provided samples voluntarily to improve science in

Pakistan. **Signature of Participant:** _____ **APPENDIX II**

STOCK REAGENTS

Phenol:Chloroform Mixture (1:1)

For each sample 200uL of phenol and 200uL of chloroform were used.

Lysis Buffer

500mM Tris-base

250mM EDTA

5% SDS

Proteinase K 75ug/ mL of lysis solution

B-mercaptoethanol (14.4M), 1uL/mL of lysis solution

50X TAE buffer

M Tris-HCl PH8

0.5 M EDTA

Make up to 1 L with dH₂O and autoclave

Bromophenol blue dye

0.1 g bromophenol blue

Dissolve

Adjust volume to 100 ml with dH₂O, stir overnight

PH to 8.0

Filter through Whatmann filter paper

Store at room temperature

10 mg/ml Ethidium bromide (EtBr)

Add 1 g of Ethidium bromide to

100 ml of ddH₂O

Stir for several hours until completely dissolved Store wrapped in aluminum foil at

4× 1M Tris

HCl

For 1L:

121.1g Tris

800ml m H₂O

~42ml HCL

Autoclave for 20 minutes on liquid setting.

SDS 20%

Dissolve 20 g SDS (sodium dodecyl sulfate or sodium lauryl sulfate) in H₂O to 100ml total with stirring (it may be necessary to heat the solution slightly to fully dissolve the powder).

Filters sterilize using a 0.45-µm filter.

Keep 10ml of 20% solution in a Falcon tube with liquids.

dNTP

Master stock: 100mM.

Create a stock solution that is 2.5mM for each dNTP.

Add 25ul of each dNTP to 900ul of H₂O for a total stock solution size of 1ml. The final recommended final concentration for each dNTP in the PCR reaction is usually 200uM. Therefore, for a 25ul PCR reaction, add 2ul of the above 2.5mM stock solution.

For Electrophoresis

6X Loading dye Solution (from Fermentas)

10mM tris-HCl (pH7.6)

10mM EDTA

0.005% bromophenol blue

0.005% xylene cyanol FF and

10% glycerol

TAE buffer, 50x

242 g Tris base

57.1 ml glacial acetic acid

37.2 g Na₂EDTA • 2H₂O (2mM)

H₂O to 1 liter

This solution does not normally needs to be sterilized. The Tris base and acetic acid correspond to 40 mM Trisacetate

TBE buffer, 10x

108 g Tris base (890mM)

55g boric acid (890mM)

40 ml 0.5 M EDTA, pH 8.0 (20mM)

H₂O to 1 liter

10x and 5x TBE tend to precipitate over time. If convenient, dilute to 2x or 1x immediately, or stir continuously. This solution does not normally need to be sterilized.

SB buffer, 50X

0.5M Sodium Hydroxide (NaOH), pH adjusted to 8.5 with boric acid (H₃BO₃).

OR

250mM Disodium borate decahydrate (Na₂B₄O₇ · 10H₂O)

DNA Ladder (Fermetas)

For:

GeneRuler DNA Ladder Mix**GeneRuler 1kb DNA Ladder Plus**

Add 20 ul of DNA Ladder, 20

ul of Loading Dye Solution

80 ul of water.

Primer Dilutions for PCR and Sequencing

Dilute primers to 5ul/ml for PCR and 10 ul/ml for Sequencing.

Master stock, 100 μ M

$$100 \mu\text{M} = X \text{ nmoles lyophilized primer} + (X \times 10 \mu\text{l molecular grade H}_2\text{O})$$

To determine the amount of H₂O to add to the lyophilized primer multiply the number of nmol of primer in the tube by 10 and that will be the amount of H₂O to add to make a 100 μ M primer stock.

The original primer tubes are often used for this 100 μ M stock.

Master stock primers newly suspended in H₂O should be allowed to sit at room temperature for 10 minutes before they are used for working stock dilutions. Mix well before making working stock dilutions.

Working stock, 10 μ M, 5 μ M

Dilute the primer master stock in a sterile micro centrifuge tube 1:10 with molecular grade H₂O.

Dilution Calculation Example

$$\text{Final Volume} \times \text{Final Concentration} = \text{Starting Concentration} \times X$$

$$\text{Example: } 100 \text{ ml} \times 5 \mu\text{g/ml} = (x \text{ ml}) \times 100 \mu\text{g/ml}$$

$$x = 5 \text{ ml}$$

so add 5 ml of stock and 95 ml H₂O

Annexure-

1 List of 23 Y-STR Haplotypes and their respective haplogroups in five Pathan populations of Charsada and Mardan district.

	DYS576	DYS389I	DYS448	DYS389II	DYS19	DYS391	DYS481	DYS549	DYS533	DYS438	DYS437	DYS570	DYS635	DYS390	DYS439	DYS392	DYS643	DYS393	DYS458	DYS385a	DYS385b	DYS456	YGATAH4	Haplogroup
MM01	20	13	20	30	16	11	23	12	12	11	14	19	24	24	10	11	10	13	15	11	14	15	13	R1a
MM02	16	14	21	31	16	10	19	13	10	10	16	17	24	23	11	11	12	13	19	13	16	16	12	G2a
MM03	17	13	19	29	14	11	21	13	13	12	15	17	23	24	13	13	10	13	16	11	14	15	12	R1b
MM04	18	12	19	29	14	10	23	13	12	10	15	16	23	21	11	16	10	11	15	14	15	16	12	L
MM05	18	13	20	29	16	11	23	13	12	11	14	17	25	24	10	11	10	13	15	11	14	15	13	R1a
MM06	17	13	20	29	16	10	23	13	12	11	14	21	23	25	12	11	10	13	18	12	14	16	13	R1a
MM07	16	13	19	29	13	10	26	11	11	11	15	18	24	22	11	15	12	13	17	11	15	16	10	Q
MM08	20	13	20	30	16	11	25	12	12	11	14	18	23	24	10	11	10	13	15	11	14	15	11	R1a
MM09	15	12	19	28	14	10	23	13	12	11	15	14	23	22	14	14	10	11	16	13	17	16	12	L
MM10	16	13	21	30	16	11	18	13	10	10	16	17	24	23	11	11	12	13	20	13	16	16	12	G2a
MM11	20	13	20	30	15	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MM12	17	13	19	30	15	10	24	13	12	10	16	16	21	22	13	14	12	12	17	9	15	15	11	L
MM13	19	13	20	30	16	11	23	12	12	11	14	19	24	24	10	11	10	13	15	11	14	15	12	R1a
MM14	19	13	20	30	16	11	23	12	12	11	14	19	24	24	10	11	10	13	15	11	14	15	12	R1a
MM15	19	13	20	30	15	11	23	12	12	11	14	19	24	24	10	11	10	13	16	11	14	15	11	R1a
MM16	20	14	20	31	16	10	23	12	12	11	14	17	23	24	12	11	10	13	17	11	16	16	13	R1a
MM17	16	13	19	29	13	10	26	11	11	11	15	18	24	22	11	15	12	13	17	11	15	16	10	Q
MM18	19	13	20	30	15	11	25	13	12	11	14	18	23	24	10	11	10	13	15	11	14	15	12	R1a
MM19	18	12	19	29	14	10	23	13	12	10	15	16	23	21	11	16	10	11	15	14	16	16	11	L
MM20	17	13	20	30	16	11	24	12	12	11	14	21	23	25	10	11	10	13	17	11	15	15	12	R1a
MM21	16	13	21	30	15	11	19	14	10	10	16	19	24	23	11	11	12	13	20	13	16	16	12	G2a
MM22	19	12	19	28	14	10	24	1	10	11	16	21	26	23	10	10	9	13	17	11	18	15	12	Q
MM23	17	12	19	29	15	11	24	12	9	9	15	15	21	24	11	11	9	13	16	14	18	13	11	J2b
MM24	16	13	21	30	16	10	19	13	10	10	16	17	23	23	11	11	12	13	18	13	16	16	12	G2a
MM25	18	13	20	30	16	11	25	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a

MM26	18	13	20	31	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MM27	17	13	20	30	16	10	23	11	12	11	14	18	24	25	10	11	10	13	16	11	14	16	13	R1a
MM28	15	12	19	28	15	10	23	13	11	9	15	13	22	24	12	11	10	13	16	13	18	13	11	J2b
MM29	19	13	21	30	16	11	23	12	12	11	14	20	23	24	10	11	10	14	15	11	14	15	13	R1a
MM30	19	13	20	29	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a

MM31	17	12	20	30	16	11	23	12	12	11	14	19	23	25	10	11	10	13	17	11	14	17	12	R1a
MM32	18	13	20	31	16	10	24	11	12	11	14	20	24	25	10	11	10	13	15	11	14	15	12	R1a
MM33	19	13	20	30	16	11	23	12	12	11	14	19	24	24	10	11	11	13	15	11	14	14	14	R1a
MM34	16	13	21	30	16	10	19	13	10	10	16	17	24	23	11	11	12	13	19	13	16	16	12	G2a
MM35	16	13	21	30	16	10	19	12	10	10	16	17	24	23	11	11	12	13	19	13	16	16	12	G2a
MM36	19	13	20	30	16	11	22	13	12	11	14	19	24	25	10	11	10	13	15	11	14	15	13	R1a
MM37	16	13	19	28	14	11	22	13	12	12	15	17	24	24	13	14	10	12	18	11	14	16	13	R1b
MM38	18	13	20	31	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MM39	16	13	19	28	14	11	22	13	12	12	15	17	24	24	13	14	10	12	18	11	14	16	13	R1b
MM40	18	13	20	31	16	11	25	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a
MM41	19	13	20	30	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MM42	19	13	20	30	16	11	25	12	12	11	14	18	23	24	10	11	10	13	15	11	14	15	12	R1a
MM43	18	13	20	31	16	11	23	13	12	11	14	19	24	24	10	11	10	13	15	11	14	15	13	R1a
MM44	16	13	21	30	15	10	19	13	10	10	16	17	24	23	11	11	12	13	18	13	16	16	12	G2a
MM45	19	13	20	30	16	11	23	12	12	11	14	19	24	24	10	11	10	13	15	11	15	15	12	R1a
MM46	16	13	21	30	17	11	19	13	10	10	16	17	24	23	11	11	12	13	20	13	16	16	12	G2a
MM47	19	13	20	30	15	11	23	12	12	11	14	19	24	24	10	11	10	13	16	11	14	15	12	R1a
MM48	18	13	18	29	15	10	25	11	12	9	14	17	21	24	11	11	9	12	16	14	17	15	11	H
MM49	18	13	20	31	15	11	23	11	13	9	14	20	23	25	10	11	10	13	16	11	14	16	12	R1a
MM50	15	12	19	28	15	10	23	13	11	9	15	13	22	24	12	11	10	13	16	13	18	13	11	J2b

MM51	15	14	20	32	15	10	23	12	12	11	14	18	23	25	10	11	10	13	16	12	14	15	13	R1a
MM52	19	13	20	30	16	11	23	12	12	11	14	19	24	24	10	11	10	13	15	11	14	16	13	R1a
MM53	18	13	20	30	16	11	23	13	12	11	14	19	24	24	10	11	10	14	15	11	14	15	13	R1a
MM54	21	13	20	30	15	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MM55	16	13	21	30	16	11	19	13	10	10	16	17	24	23	11	11	12	13	22	13	16	16	12	G2a
MM56	16	13	21	30	16	11	19	13	10	10	16	17	24	23	11	11	12	13	20	13	16	16	12	G2a
MM57	18	13	20	30	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MM58	20	13	20	31	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MM59	19	14	21	29	14	10	24	11	12	9	15	16	23	23	12	11	10	13	17	13	16	15	11	I2a
MM60	19	13	20	30	15	11	23	12	12	11	14	19	24	24	10	11	10	13	16	11	14	15	12	R1a
MM61	19	13	20	30	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MM62	17	13	20	29	16	10	23	13	12	11	14	21	23	25	12	11	10	13	18	12	14	16	13	R1a
MM63	19	14	20	30	15	10	23	12	12	11	14	19	23	24	10	11	10	13	16	11	14	15	13	R1a
MM64	16	13	21	30	16	11	19	13	10	10	16	17	24	23	11	11	12	13	20	13	16	15	11	G2a

MM65	18	12	19	29	14	10	23	13	12	10	15	16	23	21	11	16	10	11	15	14	16	16	12	L
MM66	19	13	20	30	16	11	23	12	12	11	14	19	24	24	10	11	10	13	15	11	14	15	13	R1a
MM67	17	13	20	31	14	10	26	12	11	8	14	15	22	23	11	11	10	14	16	16	20	15	12	E1b1b
MM68	20	13	20	30	16	11	25	13	12	11	14	18	23	25	10	11	10	13	15	11	14	15	11	R1a
MM69	17	13	19	29	14	11	21	13	13	12	15	17	23	24	12	13	10	13	16	11	14	15	12	R1b
MM70	17	13	19	30	13	10	22	12	11	11	16	15	25	23	10	10	10	13	16	13	19	15	12	L
MM71	19	13	20	30	15	11	23	12	12	11	14	19	24	24	10	11	10	13	16	11	14	15	12	R1a
MM72	19	14	21	29	14	10	24	11	12	9	15	16	23	23	12	11	10	13	17	13	16	15	11	I2a
MM73	17	13	19	29	14	11	21	13	13	12	15	17	23	24	13	13	10	13	16	11	14	15	12	R1b
MM74	15	13	19	29	15	10	24	13	12	10	16	16	21	22	13	14	12	12	17	9	17	14	11	L
MM75	16	13	19	29	15	10	24	13	12	10	16	16	21	22	13	14	12	12	16	9	16	15	11	L
MZ01	18	14	20	30	15	10	23	12	12	9	14	17	20	22	12	11	9	12	18	14	19	16	12	H

MZ02	19	14	20	31	16	11	23	12	12	11	14	19	24	24	10	11	10	13	15	12	14	15	13	R1a
MZ03	18	14	19	31	14	10	22	13	12	19	14	17	21	24	12	12	10	13	17	13	16	17	11	E1b1b
MZ04	19	13	20	29	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	11	R1a
MZ05	19	13	21	29	16	11	26	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MZ06	19	14	20	31	15	10	22	12	11	9	15	15	22	23	11	11	10	12	15	14	19	15	12	L
MZ07	20	13	20	30	16	11	23	13	12	11	14	19	24	24	10	11	10	13	15	11	14	15	13	R1a
MZ08	15	12	21	28	15	10	22	12	9	10	16	16	19	22	12	10	11	14	18	15	16	15	11	G2a
MZ09	19	13	20	30	15	11	18	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a
MZ10	15	13	19	29	15	10	24	14	12	10	16	16	21	22	13	14	12	12	17	9	16	15	11	L
MZ11	16	13	21	30	16	11	19	13	10	10	16	17	24	23	11	11	12	13	19	13	16	16	12	G2a
MZ12	20	13	20	30	16	11	25	13	12	11	14	18	23	25	10	11	10	13	15	11	14	15	11	R1a
MZ13	16	13	21	30	16	10	19	13	10	10	16	17	24	23	11	11	12	13	19	13	15	16	12	G2a
MZ15	19	13	20	30	16	10	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MZ14	14	13	19	29	14	10	23	13	12	11	16	17	25	23	11	10	11	13	17	12	17	16	11	L
MZ16	14	13	19	29	14	10	23	13	12	11	16	17	25	23	11	10	11	13	17	12	17	16	11	L
MZ17	18	13	19	27	14	10	26	13	10	11	15	18	23	23	11	12	10	13	17	13	21	15	11	Q
MZ18	17	13	20	30	16	11	21	12	12	11	14	19	23	25	11	11	10	13	16	11	14	15	12	R1a
MZ19	20	13	20	30	16	11	25	12	12	11	14	19	23	23	10	11	10	13	15	11	14	15	11	R1a
MZ20	18	13	20	30	15	10	23	12	12	9	14	17	20	22	12	11	9	12	18	14	19	16	12	H
MZ21	19	14	20	31	15	10	23	12	12	11	14	19	23	25	10	?	10	13	16	11	15	15	12	R1a
MZ22	19	12	19	28	15	10	23	13	11	10	15	18	21	23	11	13	11	12	16	15	19	14	13	L
MZ23	17	13	19	31	16	9	22	13	12	11	14	18	23	25	10	11	10	13	16	11	14	15	12	R1a

MZ24	19	12	20	28	16	10	23	13	11	10	15	18	21	23	11	13	11	12	16	15	19	14	13	L
MZ25	18	13	21	30	16	11	23	12	11	11	14	20	23	26	10	11	10	13	15	11	14	16	13	R1a
MZ26	16	13	19	30	15	10	23	13	12	10	14	19	20	21	11	11	19	12	17	14	18	15	11	H
MZ27	16	13	19	30	15	10	21	11	12	10	16	19	21	22	13	11	13	14	17	14	15	17	13	G2a

MZ28	16	12	18	27	15	10	23	12	12	10	15	15	22	22	12	14	10	11	16	13	20	16	13	L
MZ29	17	14	20	31	17	10	23	12	11	11	14	19	23	25	10	11	19	13	16	11	15	15	13	R1a
MZ30	20	13	20	30	17	11	23	12	12	10	14	18	23	25	10	11	10	13	16	11	14	15	12	R1a
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MZ32	20	13	20	30	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	11	R1a
MZ33	19	13	19	28	15	10	24	12	11	11	16	18	24	23	11	10	10	14	17	12	20	16	11	L
MZ34	20	14	19	30	13	10	25	12	11	11	16	17	23	21	11	15	11	13	17	12	17	15	10	Q
MZ35	19	13	20	29	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	11	R1a
MZ36	19	13	20	30	15	11	23	12	12	11	14	19	24	24	10	11	10	13	16	11	14	15	12	R1a
MZ37	20	13	20	30	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	11	R1a
MZ38	19	13	20	31	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MZ39	20	13	20	29	15	11	25	12	12	11	14	19	23	24	10	11	10	13	16	11	14	15	11	R1a
MZ40	19	13	20	30	16	11	23	12	12	11	14	19	24	24	10	11	10	13	15	11	14	15	13	R1a
MZ41	19	13	20	30	15	11	25	12	12	11	14	19	23	25	10	11	10	13	15	11	14	15	12	R1a
MZ42	18	14	19	29	15	11	24	12	11	11	17	17	24	23	11	10	10	14	17	13	20	16	11	L
MZ43	19	13	20	30	16	11	25	12	12	11	15	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MZ44	19	13	20	30	17	11	24	12	12	11	14	19	24	25	10	11	11	13	15	11	14	15	14	R1a
MZ45	21	14	20	31	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	11	R1a
MZ46	16	13	19	29	15	10	23	12	12	10	14	18	21	22	12	11	9	12	18	16	17	15	12	H
MZ47	18	13	19	28	15	10	24	12	11	11	16	18	24	23	11	10	10	14	17	13	20	16	11	L
MZ49	18	13	20	30	16	10	24	13	12	11	14	18	24	25	10	11	8	13	16	11	14	16	13	R1a
MZ48	20	13	20	29	16	11	25	12	12	11	14	20	23	24	11	11	10	13	15	11	14	15	11	R1a
MZ50	17	13	20	30	17	11	23	11	12	11	14	18	23	25	10	11	10	13	16	11	14	15	13	R1a
MZ51	16	13	19	29	15	10	24	12	11	11	14	17	25	22	11	10	10	14	18	14	19	15	12	Q
MZ52	19	13	20	30	15	10	26	12	12	11	14	18	23	24	10	11	10	13	15	11	14	15	12	R1a
MZ53	15	13	20	30	15	7	21	13	12	8	14	11	20	21	12	11	10	12	14	13	16	15	11	G2a
MZ54	15	13	20	30	15	7	25	13	12	11	14	19	23	24	10	11	10	13	15	11	14	15	11	R1a

MZ55	16	13	19	30	15	7	24	13	12	10	16	16	21	22	13	14	12	12	17	9	16	15	11	L
MZ56	19	13	20	29	16	11	22	12	12	11	14	21	23	25	10	11	10	13	15	11	14	14	13	R1a
MZ57	18	14	20	30	16	9	28	12	13	10	14	16	21	23	12	12	10	14	19	13	15	15	12	I2a (xI2a1)

MZ58	20	13	20	29	15	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	11	R1a
MZ59	15	13	19	29	15	10	25	14	12	10	16	16	21	22	11	12	10	12	17	9	17	15	12	L
MZ60	16	13	21	30	16	11	19	13	10	10	16	17	24	23	11	11	12	13	19	13	16	16	12	G2a
MZ61	16	13	19	30	16	10	23	13	12	10	14	19	20	22	11	11	9	12	17	14	18	15	11	H
MZ62	19	13	20	30	16	11	24	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MZ63	19	13	20	30	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
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MZ65	19	12	20	28	15	10	27	12	11	11	14	20	24	24	12	11	11	12	19	12	13	15	11	Q
MZ66	20	13	20	30	15	12	25	12	12	11	14	19	23	24	10	11	10	13	15	11	15	15	12	R1a
MZ67	16	12	19	28	15	11	23	12	9	9	15	15	21	24	11	11	9	13	16	13	19	13	11	J2b
MZ68	18	10	22	27	13	10	23	12	11	10	14	14	21	24	14	11	12	13	17	16	17	16	11	E1b1b
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MZ70	18	13	20	30	16	10	23	12	11	11	14	19	23	25	10	11	10	14	16	11	14	15	12	R1a
MZ71	16	13	21	30	16	10	19	13	10	10	16	17	24	23	11	11	12	3	18	13	17	16	12	G2a
MZ72	19	13	20	32	15	10	23	12	13	11	14	18	23	24	10	11	10	13	15	11	14	15	11	R1a
MZ73	18	13	19	29	13	10	25	14	11	11	16	17	25	23	11	10	9	13	18	13	18	15	12	Q
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MD01	19	13	20	30	16	11	26	12	12	11	14	19	23	24	10	11	10	13	16	11	14	15	12	R1a
MD02	21	11	17	26	14	10	27	10	11	10	14	21	22	24	8	11	11	12	19	12	18	16	11	J1
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MD04	18	14	20	31	16	10	22	12	11	11	14	18	23	25	10	11	10	13	16	11	14	15	13	R1a
MD05	17	14	19	30	15	10	24	12	12	11	16	19	26	24	11	10	10	14	16	12	13	16	12	R1a

MD06	16	13	21	30	16	11	19	13	10	10	16	17	24	23	11	11	12	13	19	13	16	14	12	G2a
MD07	19	13	20	30	15	12	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MD08	19	13	20	30	16	11	23	12	12	11	14	18	24	24	10	11	10	13	15	11	14	15	13	R1a
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MD17	18	13	20	30	17	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
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MD21	18	12	21	32	14	10	23	12	11	10	16	19	22	22	11	11	9	13	15	14	16	15	12	G2a
MD22	19	14	18	30	14	10	24	12	12	12	16	17	27	24	11	12	10	13	14	12	13	15	12	R1b
MD23	16	13	19	29	15	10	24	13	12	10	15	16	21	22	12	14	12	12	17	9	17	16	11	L
MD24	16	13	19	29	15	10	24	11	11	10	16	16	22	22	12	14	12	12	18	7	15	13	11	L
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MD32	15	13	19	29	15	10	24	13	12	10	15	16	21	22	12	14	12	12	17	9	17	16	11	L
MD33	20	13	18	29	14	10	22	14	12	12	15	15	23	24	13	13	10	12	16	11	14	15	12	R1b
MD34	16	13	19	29	15	10	24	13	12	10	15	16	21	22	12	14	12	12	17	9	17	16	11	L
MD35	19	13	20	30	16	11	25	12	12	11	14	20	23	24	10	11	10	13	15	11	14	14	12	R1a
MD36	16	13	19	29	15	10	24	13	12	10	15	16	21	22	12	14	12	12	17	9	17	16	11	L
MD37	16	14	19	30	15	10	23	15	12	9	14	16	21	22	11	11	9	12	18	15	17	15	12	H
MD38	19	13	20	30	16	11	25	12	12	11	14	19	23	24	10	11	10	13	16	11	14	15	12	R1a
MD39	19	13	20	30	16	11	25	12	12	11	14	19	24	24	10	11	10	13	15	11	14	15	12	R1a
MD40	19	15	19	30	14	10	25	11	12	11	14	19	27	23	12	10	11	14	17	13	20	16	11	Q
MD41	19	13	20	30	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MD42	16	13	19	29	15	10	24	11	11	10	16	16	22	22	12	14	12	12	18	7	15	13	11	L
MD43	17	13	21	30	16	12	23	12	12	11	14	18	23	25	10	11	10	13	16	11	14	15	12	R1a
MD44	17	13	21	30	16	12	23	12	12	11	14	18	23	25	10	11	10	13	16	11	14	15	12	R1a
MD45	19	13	20	30	14	11	25	12	12	11	14	19	23	25	10	11	10	13	15	10	14	15	12	R1a
MD46	19	13	20	30	16	11	23	12	12	11	14	18	24	24	10	11	10	13	15	11	14	16	13	R1a
MD47	16	13	19	29	15	10	24	13	12	10	15	16	21	22	12	14	12	12	17	9	17	16	11	L
MD48	19	14	18	30	14	10	24	12	12	12	16	17	27	24	11	12	10	13	14	12	13	15	12	R1b
MD49	18	12	19	28	15	10	22	12	11	10	14	16	19	23	13	12	11	12	16	11	16	14	12	L
MD50	18	14	19	31	16	11	23	12	13	11	14	20	23	24	10	11	10	13	16	11	14	15	13	R1a

MD51	19	13	19	30	16	11	25	12	12	11	14	19	23	23	10	11	10	13	15	11	14	15	12	R1a
MD52	19	13	20	30	16	11	26	12	12	11	14	19	23	24	10	11	10	13	15	11	14	16	12	R1a
MD53	18	13	19	29	14	10	22	10	12	11	15	17	23	22	13	15	12	13	20	12	16	15	10	L
MD54	19	13	20	30	16	11	23	13	12	11	14	20	24	24	10	11	10	13	15	11	14	15	13	R1a
MD55	15	13	19	31	15	10	26	13	11	11	16	19	26	24	11	10	10	14	16	12	18	16	12	E1b1b
MD56	15	13	19	29	15	10	24	14	12	10	15	16	21	22	12	14	12	12	17	9	17	16	11	L
MD57	15	13	19	29	15	10	24	13	12	10	15	16	21	22	12	14	12	12	17	9	17	16	11	L

MD58	19	14	19	31	14	10	25	12	11	9	15	18	21	24	12	11	9	12	17	13	16	16	11	J1
MD59	19	14	19	30	14	10	25	13	11	11	15	19	25	23	10	10	10	14	16	13	22	15	11	Q
MD60	18	13	20	30	16	11	25	12	12	11	14	19	23	24	10	11	10	13	15	10	14	15	12	R1a
MD61	19	13	20	30	15	11	25	12	13	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MD62	20	11	17	26	14	10	27	10	11	10	14	20	22	24	8	11	11	12	19	12	17	16	11	J1
MD63	16	14	21	31	16	10	19	13	10	10	16	16	24	23	11	11	12	13	22	13	16	15	12	G2a
MD64	19	13	20	30	15	11	25	12	12	11	14	18	23	23	10	11	10	13	15	11	14	15	12	R1a
MD65	16	13	21	30	17	11	19	13	10	10	16	17	22	23	11	11	12	13	19	13	16	16	12	G2a
MD66	15	12	19	27	15	10	23	14	10	9	15	19	21	23	11	11	9	12	17	12	17	13	11	J2b
MD67	19	13	20	29	15	11	23	12	12	11	14	18	23	25	10	11	10	13	17	11	14	15	12	R1a
MD68	16	13	21	30	15	11	19	13	10	10	16	17	23	23	11	11	12	13	19	13	16	15	12	G2a
MD69	15	13	20	30	15	10	24	11	12	10	16	15	13	22	12	14	14	12	17	9	16	15	11	L
MD70	19	13	20	31	16	11	25	12	12	11	14	19	24	24	10	11	10	13	15	11	14	15	12	R1a
MD71	18	12	19	28	17	10	25	11	11	10	15	16	20	23	12	11	10	12	18	14	18	14	12	L
MD72	19	13	20	30	16	11	26	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
MD73	16	13	21	30	16	10	19	14	10	10	16	17	23	23	11	11	12	13	19	13	16	15	12	G2a
MD74	18	13	19	29	17	10	25	14	11	11	16	17	25	23	11	10	9	13	18	13	18	15	12	L
YS01	16	13	21	30	16	11	19	13	10	10	16	17	24	23	11	11	12	13	19	13	16	16	12	G2a
YS02	18	13	20	30		11	23	13	12	11	14	18	24	24	10	11	10	13	16	11	14	15	13	R1a
YS03	17	13	20	30	15	11	23	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
YS04	18	13	20	30		10	22	12	12	11	14	20	23	24	10	11	10	13	16	11	14	15	13	R1a
YS05	18	13	20	30	16	11	23	13	12	11	14	17	26	24	11	11	10	13	15	11	13	15	13	R1a
YS06	17	13	20	31	16	10	23	12	12	11	14	18	23	25	10	11	8	13	15	11	14	16	13	R1a
YS07	18	14	19	31	17	10	26	11	11	10	15	15	22	21	12	11	11	13	15	12	21	16	12	L
YS08	18	13	20	29	16	11	23	13	12	11	14	19	23	24	11	11	10	13	15	11	14	15	12	R1a
YS09	18	13	19	28	14	10	22	13	12	12	15	17	23	24	13	14	10	12	17	11	14	16	12	R1b

YS10	18	13	20	29	16	11	23	13	12	11	14	19	23	24	11	11	10	13	15	11	14	15	12	R1a
YS11	16	13	20	29	15	10	24	11	12	10	16	14	24	22	12	14	13	13	17	9	17	15	11	L
YS12	19	14	19	30	14	11	26	13	11	11	14	18	24	23	11	10	10	14	19	13	19	14	12	Q
YS13	18	13	20	29	16	11	23	13	12	11	14	19	23	24	11	11	10	13	15	11	14	15	12	R1a
YS14	16	12	19	28	14	11	23	13	12	10	15	14	23	22	13	14	10	11	15	13	19	16	12	L
YS15	20	14	20	31	15	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
YS16	18	13	20	31	17	11	23	13	12	11	14	20	23	25	10	11	10	13	15	11	14	15	13	R1a
YS17	18	13	20	31	15	11	23	12	12	11	14	20	23	25	10	12	11	13	16	11	14	15	12	R1a
YS18	19	12	21	31	14	11	23	12	11	10	16	20	23	22	11	11	9	13	16	14	16	15	12	G2a
YS19	19	13	20	30	15	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
YS20	17	14	20	32	15	10	23	12	13	11	14	18	23	25	10	11	10	13	15	11	14	15	11	R1a
YS21	21	13	20	30	15	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	G2a
YS22	17	13	20	30	16	10	22	11	10	10	16	19	21	22	12	11	11	13	17	13	14	15	14	R1a
YS23	18	13	20	30	16	11	23	12	12	11	14	19	23	25	10	11	10	13	14	11	14	15	12	R1a
YS24	19	13	20	30	16	11	24	13	12	11	14	18	24	24	10	11	10	13	15	11	14	15	13	R1a
YS25	16	12	19	28	14	10	23	13	12	11	15	15	25	23	13	15	10	11	14	13	16	16	12	L
YS26	18	14	18	30	14	10	23	12	13	12	15	18	25	21	12	15	13	13	17	12	15	15	10	R1b
YS27	18	13	20	31	16	11	23	13	12	11	14	19	23	25	11	11	10	15	16	11	14	15	12	R1a
YS28	20	13	20	29	14	11	22	11	12	12	15	17	23	24	13	14	10	11	16	11	14	17	12	R1b
YS29	17	13	0	30	14	10	24	12	11	10	14	17	18	23	12	11	11	14	17	14	15	15	12	E1b1a
YS30	17	14	20	32	16	10	23	12	13	11	14	18	23	25	10	11	10	13	15	11	15	15	11	R1a
YS31	18	13	19	29	15	10	27	14	11	9	15	19	21	24	11	11	11	13	16	14	16	14	12	I2a (xI2a1)
YS32	16	14	21	31	16	10	19	13	10	10	15	16	24	23	11	11	12	13	19	13	16	16	12	G2a
YS33	18	13	20	30	16	11	23	13	12	11	14	17	26	24	11	11	10	13	15	11	13	15	13	R1a
YS34	16	13	21	30	15	10	19	13	10	10	16	17	23	23	11	11	12	13	20	13	16	16	12	G2a
YS35	18	13	20	29	16	11	23	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a

YS36	19	14	21	32	15	10	24	12	12	11	14	18	24	24	10	11	10	13	17	11	15	15	12	R1a
YS37	17	12	18	30	15	10	26	13	12	11	14	18	23	24	11	11	11	14	16	13	16	15	12	Q
YS38	16	14	19	30	13	10	23	13	10	9	14	14	20	23	11	13	11	14	15	13	16	14	12	L
YS39	18	12	19	28	14	10	23	12	12	10	15	15	25	22	12	14	10	11	15	13	17	15	12	L
YS40	18	13	20	30	16	11	23	13	12	11	14	17	26	24	11	11	10	13	15	11	13	15	13	R1a
YS41	17	12	19	28	16	10	23	13	11	11	16	18	24	23	10	10	10	14	19	13	17	15	12	L
YS42	18	13	20	30	16	10	22	12	12	11	14	20	23	24	10	11	10	13	16	11	14	16	13	R1a
YS43	17	13	21	30	14	10	24	12	11	9	14	15	21	23	11	11	10	13	15	15	20	15	13	E1b1a
YS44	19	13	20	30	15	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a

YS45	18	13	20	30		10	23	12	12	11	14	18	24	25	10	11	8	13	16	11	14	16	13	R1a
YS46	20	13	20	30		11	25	12	12	11	0	19	23	24	10	11	10	13	15	11	14	15	12	R1a
YS47	19	14	21	32	15	10	24	12	12	11	14	18	24	24	10	11	11	13	17	11	14	15	12	R1a
YS48	17	13	19	32	15	10	21	11	12	10	16	16	21	23	13	11	15	14	16	14	15	17	12	G2A
YS49	18	13	20	30	16	10	23	12	12	11	14	20	23	24	10	11	10	13	17	11	14	15	11	R1a
YS50	18	13	20	30		10	23	13	12	11	14	18	24	25	10	11	8	13	16	11	14	16	13	R1a
YS51	15	13	20	29	15	10	24	12	11	10	16	15	22	20	12	14	11	12	18	9	16	15	12	L
YS52	16	13	21	30	15	10	19	13	10	10	16	17	23	23	11	11	12	13	20	13	16	16	12	G2a
YS53	17	12	18	28	15	10	26	13	12	11	14	18	23	24	11	14	11	14	16	13	16	15	11	Q
YS54	17	14	19	30	15	10	26	12	11	11	16	17	25	23	11	10	10	14	19	14	18	16	13	Q
YS55	18	13	20	29	17	11	23	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
YS56	20	13	20	30	15	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
YS57	20	13	20	30	15	11	25	12	12	11	14	19	23	24	10	11	10	13	14	11	14	15	12	R1a
YS58	18	14	20	30	15	10	24	12	12	11	14	16	23	25	10	11	10	13	17	11	16	15		R1a
YS59	19	13	21	32	15	10	24	12	12	11	14	18	24	24	10	11	10	13	17	11	14	15	12	R1a
YS60	16	12	19	28	14	11	23	13	12	10	15	15	23	22	13	14	10	11	15	12	18	16	12	L

YS61	19	14	20	30	14	10	22	11	12	10	14	18	23	23	12	13	11	12	17	15	17	14	12	L
YS62	17	12	19	28	15	10	25	14	11	11	16	19	24	23	10	10	10	14	19	13	16	15	12	L
YS63	16	13	20	29	15	10	24	11	12	10	16	14	24	22	12	14	13	13	17	9	17	15	11	L
YS64	18	14	20	31	14	10	25	13	11	10	14	16	21	24	12	11	9	12	18	11	16	15	11	J1
YS65	17	13	?	29	14	10	24	12	11	10	14	17	18	23	12	11	11	14	17	14	15	15	12	E1b1a
YS66	18	13	20	29	?	11	23	12	12	11	14	19	?	24	10	11	10	13	15	11	14	15	12	R1a
YS67	17	14	19	30	15	10	24	15	11	11	15	15	25	23	11	10	10	14	18	13	17	15	12	L
YS68	19	14	21	32	15	10	24	12	12	11	14	18	24	24	10	11	10	13	17	11	14	15	13	R1a
YS69	19	13	20	32	15	10	23	13	12	11	14	19	23	25	10	11	10	13	15	11	14	15	13	R1a
YS70	18	14	20	33	15	10	23	12	12	10	14	20	23	25	10	12	10	13	14	11	14	16	12	R1a
YS71	16	13	21	29	17	11	19	13	10	10	16	18	25	23	11	11	12	13	17	13	16	15	11	G2a
YS72	19	13	20	30	17	11	23	12	11	11	14	19	24	25	10	11	10	13	15	11	14	15	13	R1a
YS73	17	13	22	30	14	10	24	12	11	10	14	17	18	23	12	11	11	14	17	14	15	15	12	E1b1a
YS74	18	13	20	31	15	11	23	12	12	11	14	20	23	25	10	12	11	13	16	11	14	15	13	R1a
YS75	10	14	20	31	15	11	25	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
KM01	18	13	20	30	16	10	23	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a
KM02	16	12	19	29	15	11	23	12	9	9	15	15	21	24	11	11	9	13	16	13	18	13	11	J2b
KM03	17	13	19	20	15	9	23	12	12	9	15	15	21	23	11	11	11	13	15	14	19	15	12	L

KM04	18	13	21	29	15	10	23	11	12	9	15	16	21	23	12	11	10	12	15	15	18	15	12	L
KM05	18	14	19	31	15	10	24	13	11	11	15	19	26	23	10	10	10	14	18	13	19	15	11	Q
KM06	16	13	19	29	15	10	23	13	12	10	14	19	20	22	11	11	9	12	17	14	18	15	11	H
KM08	16	13	19	29	15	10	24	11	12	11	14	19	23	25	10	11	10	13	15	11	14	15	12	R1a
KM07	17	14	20	30	16	10	24	11	12	11	14	19	23	25	10	11	10	13	15	11	14	15	12	R1a
KM09	18	13	20	30	15	12	24	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a
KM10	18	13	20	31	16	11	23	11	12	11	14	19	23	24	10	11	10	13	16	10	14	17	12	R1a
KM11	18	13	20	30	16	11	24	12	12	11	14	21	23	24	10	11	10	13	15	11	14	15	12	R1a

KM12	20	14	20	31	16	10	24	12	12	11	14	19	24	24	10	11	10	13	17	12	14	15	11	R1a
KM13	19	13	20	30	16	11	23	12	12	11	14	19	25	24	10	11	10	13	15	11	14	15	13	R1a
KM14	18	13	20	30	16	11	23	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a
KM15	18	13	20	29	16	11	23	12	12	11	14	21	23	24	10	11	10	13	15	11	14	15	12	R1a
KM16	16	13	21	30	16	10	19	13	10	10	16	17	24	23	11	11	12	13	19	13	16	16	12	G2a
KM17	17	13	19	29	16	10	25	11	12	10	16	16	21	22	13	16	11	12	17	9	16	14	12	L
KM18	18	13	20	30	16	10	23	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a
KM19	17	13	19	29	16	10	25	11	12	10	16	16	21	22	13	16	11	12	17	9	16	14	12	L
KM20	18	13	20	30	16	11	23	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a
KM21	18	13	20	30	16	11	24	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a
KM22	18	13	20	30	16	11	24	12	12	11	14	21	23	24	10	11	10	13	15	11	14	15	12	R1a
KM23	18	13	20	30	16	10	24	13	11	9	14	15	21	23	11	11	9	13	16	14	20	16	11	H
KM24	18	13	20	30	16	11	24	12	12	11	14	21	23	24	10	11	10	13	15	11	14	15	12	R1a
KM25	18	13	20	30	16	11	23	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a
KM26	18	13	20	30	16	10	23	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a
KM27	16	13	19	29	16	9	24	12	11	10	14	16	18	22	10	11	12	15	18	13	20	15	10	E1b1a
KM28	14	12	18	28	15	10	23	12	11	9	15		21	24	12	11	9	12	15	14	17	13	11	J2b
KM29	18	13	20	30	16	11	23	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a
KM30	18	14	20	31	14	10	23	13	12	11	14	19	24	25	10	11	10	13	16	11	14	15	11	R1a
KM31	18	13	20	30	16	11	23	12	12	11	14	20	23	24	10	11	10	14	15	11	14	15	12	R1a
KM32	18	13	20	30	16	11	24	12	12	11	14	20	23	24	10	11	10	13	15	11	14	15	12	R1a
KM33	18	13	20	30	16	10	24	13	11	9	14	15	21	23	11	11	9	13	16	13	19	16	11	J1
KM34	18	13	20	30	0	10	24	13	11	9	14	15	21	23	11	11	9	13	16	14	20	16	11	H
KM35	18	13	20	30	0	10	24	13	11	9	14	15	21	23	11	11	9	13	16	14	20	16	11	H
KM36	19	13	20	30	0	11	23	13	12	11	14	19	24	24	10	11	10	13	15	11	14	15	13	R1a
KM37	16	12	19	28	14	10	23	13	12	10	15	15	23	22	13	14	10	11	15	13	19	16	12	L

KM38	17	12	18	28	0	9	24	13	12	11	16	17	24	24	12	10	11	14	17	14	20	15	12	Q
KM39	18	13	20	30	14	10	24	13	11	9	14	15	21	23	11	11	9	13	16	14	20	16	11	H
KM40	15	13	20	29	14	10	24	12	11	9	16	17	21	23	12	11	10	12	17	13	17	15	11	J1
KM41	18	13	20	30	15	10	24	13	11	9	14	15	21	23	11	11	9	13	16	14	20	16	11	H
KM42	18	13	20	30	16	10	24	13	11	9	14	15	21	23	11	11	9	13	16	14	20	16	11	H
KM43	18	13	20	30	14	10	24	13	11	9	14	15	21	23	11	11	9	13	16	14	20	16	11	H
KM44	16	12	19	24	14	10	23	13	12	10	15	15	23	22	13	14	10	11	15	13	19	16	12	L
KM45	16	13	21	30	16	10	19	13	10	10	16	18	23	23	11	11	12	13	19	13	16	16	12	G2a
KM46	18	13	21	30	14	10	23	13	11	9	15	16	22	23	13	11	10	12	15	13	15	15	12	J1
KM47	19	13	20	30	16	11	23	13	12	11	14	18	24	24	10	11	10	13	15	11	14	15	13	R1a
KM48	17	12	18	28	14	9	26	13	12	11	16	17	24	24	12	10	11	15	17	14	20	15	12	Q
KM49	18	13	20	30	16	11	23	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	R1a
KM50	17	12	18	28	0	9	26	13	12	11	16	17	24	24	12	10	11	15	17	14	20	15	12	Q
KM51	16	12	19	28	14	10	23	13	12	10	15	15	23	22	12	14	10	11	15	13	19	16	12	L
KM52	18	13	20	30	15	11	23	12	12	11	14	19	23	24	10	11	10	13	15	11	14	15	12	Q
KM53	16	12	19	26	14	10	23	13	12	10	15	15	23	22	13	14	10	11	15	13	19	16	12	L
KM54	17	12	18	28	14	9	26	13	12	11	16	17	24	24	12	10	11	15	17	14	20	15	12	Q
KM55	19	13	20	30	16	11	23	13	12	11	14	18	24	24	10	11	10	13	15	11	14	15	13	R1a
KM56	17	12	18	28	14	9	26	13	12	11	16	17	24	24	12	10	11	15	17	14	20	15	12	Q
KM57	17	12	18	28	14	9	26	13	12	11	16	17	24	24	12	10	11	14	17	14	20	15	12	Q
KM58	16	12	19	28	14	11	23	13	12	10	15	15	23	22	13	14	10	11	15	13	19	16	12	L
KM59	16	12	19	28	14	10	23	13	12	10	15	15	23	22	13	14	10	11	15	13	19	16	12	L
KM60	18	13	19	28	14	10	22	13	12	12	15	17	23	24	13	14	10	12	17	11	14	16	12	R1b
KM61	16	13	21	30	17	10	19	13	10	10	16	17	23	23	11	11	12	13	19	13	16	16	12	G2a
KM62	17	12	18	28	14	9	26	13	12	11	16	17	24	24	12	10	11	14	17	14	19	15	12	Q
KM63	16	12	19	28	14	11	23	13	12	10	15	15	23	22	13	14	10	11	15	13	19	16	12	L
KM64	18	13	21	30	14	10	23	13	11	9	15	16	22	23	13	11	10	12	15	13	15	15	12	J1

KM65	18	13	20	30	16	11	23	12	12	11	14	19	23	25	10	11	10	13	14	11	14	15	12	R1a
KM66	18	13	21	30	14	10	23	13	11	9	15	16	22	23	13	11	10	12	15	13	15	15	12	J1
KM67	17	12	18	28	14	9	26	13	12	11	16	17	24	24	12	10	11	15	17	14	20	15	12	Q
KM68	18	13	20	30	16	11	23	12	12	11	14	18	23	24	10	11	10	13	15	11	14	15	11	R1a
KM69	19	14	20	31	16	10	24	12	12	11	14	19	24	24	10	11	10	13	17	12	14	15	11	R1a
KM70	19	13	20	30	15	10	26	12	12	11	14	19	23	24	10	11	10	13	14	11	14	15	12	R1a

KM71	19	14	20	31	16	10	24	12	12	11	14	19	24	24	10	11	10	13	17	12	14	15	11	R1a
KM72	16	12	19	28	14	11	23	13	12	10	15	15	23	22	13	14	10	11	15	13	19	16	12	L
KM73	16	12	19	28	14	11	23	13	12	10	15	15	23	22	13	14	10	11	15	13	19	16	12	L
KM74	16	12	19	28	14	11	23	13	12	10	15	15	23	22	13	14	10	11	15	13	19	16	12	L
KM75	20	13	20	30	15	11	26	12	12	11	14	19	23	24	10	11	10	13	15	11	13	15	12	R1a

